



Webmail
Univ. Bung Hatta

hafrizal syandri <syandri_1960@bunghatta.ac.id>

Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security : Manuscript returned to draft

6 messages

Srimathi Anandan <srinathi.anandan@hindawi.com>
To: Hafrijal Syandri <syandri_1960@bunghatta.ac.id>
Cc: Deepanaa Rajadurai <deepanaa.rajadurai@hindawi.com>

Wed, Jul 12, 2023 at 6:32 PM



Hindawi



Dear Dr. Hafrijal Syandri,

We have reviewed your manuscript "Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security " with ID No. 6639837. We found the following issues which need to be solved before moving on:

- **Authors & Affiliations**

- - While checking your manuscript you have mentioned that two different email addresses for co-author are Dr. Harfiandri Damanhuri on the system "d.harfiandri@bunghatta.ac.id". Whereas in the cover letter file you have mentioned "d.harfiandri@bunghatta.ac.id". Can you please confirm your preferred email address for the further process?

- **Relevance Check**

- - You have mentioned that there is no conflict of interest in the system and the manuscript file but you have provided funding grant details too. Hence please confirm whether there is any conflict of interest for you and the co-author

“

This paper has been returned to draft for the reasons listed above. Please could you log into your account and address these points as soon as possible? Once completed, please could you resubmit the manuscript in the same manuscript number "6639837" and do not start a new submission.

Please log into your account to make the required updates. Follow the steps outlined below, and we kindly ask that you do not submit a brand new manuscript.

1. Click on the manuscript tile for Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security , which will show the status "Complete submission".
2. You will see all 4 submission steps. All the information will be pre-filled as you submitted the manuscript, so you will not need to complete the entire submission again. Click through the steps until you reach the area that needs updating.
3. If the manuscript file needs updating then go to step 4, and click the small bin icon next to the manuscript file. This will delete the file. You can then upload the new updated file.
4. When all necessary changes have been made, click the green "Submit manuscript" button at the bottom of step 4, and this will return the manuscript to the Hindawi team.

King regards,
Srimathi Anandan

This email was sent to syandri_1960@bunghatta.ac.id. You have received this email in regards to the account creation, submission, or peer review process of a paper submitted to a journal published by Hindawi Limited.

Hindawi Limited, 3rd Floor, Adam House, [1 Fitzroy Square, London, W1T 5HF, United Kingdom](#).

Hindawi respects your right to privacy. Please see our [privacy policy](#) for information on how we store, process, and safeguard your data.

[Unsubscribe](#)

hafrizal syandri <syandri_1960@bunghatta.ac.id>
To: Srimathi Anandan <srimathi.anandan@hindawi.com>

Thu, Jul 13, 2023 at 10:57 AM

Dear
Srimathi Anandan
Editor Team International Journal of Food Science

Thank you for your email on July 12, 2023; We have revised the email (d.harfiandri@bunghatta.ac.id and changed it to harfiandri@bunghatta.ac.id). Additionally, we have revised manuscript No. 6639837, especially part Acknowledgements related to the conflict of interest.

Best Regards
Hafrijal Syandri
[Quoted text hidden]

Srimathi Anandan <srimathi.anandan@hindawi.com>
Reply-To: Srimathi Anandan <srimathi.anandan@hindawi.com>
To: syandri_1960@bunghatta.ac.id
Cc: syandri_1960@bunghatta.ac.id, deepanaa.rajadurai@hindawi.com

Thu, Jul 13, 2023 at 2:24 PM

Dear Dr. Syandri,

Thank you for the email.

While checking the email address d.harfiandri@bunghatta.ac.id is verified one, could you please update this in the system as well as in the cover letter?

Best Regards,
Srimathi Anandan

Srimathi Anandan
Editorial Screener

e. srimathi.anandan@hindawi.com



[Quoted text hidden]

, hafrizal syandri <syandri_1960@bunghatta.ac.id> wrote:

[Quoted text hidden]

[Quoted text hidden]

[Quoted text hidden]

[Quoted text hidden]

Hindawi respects your right to privacy. Please see our [privacy policy](#) for information on how we store, process, and safeguard your data.

[Unsubscribe](#)

The contents of this email and any attachments are confidential and intended only for the person or entity to whom it is addressed. If you are not the intended recipient, any use, review, distribution, reproduction or any action taken in reliance upon this message is strictly prohibited. If you received this message in error, please immediately notify the sender and permanently delete all copies of the email and any attachments.

[Click here for translations of this disclaimer.](#)

Srimathi Anandan <srimathi.anandan@hindawi.com>
To: Hafrijal Syandri <syandri_1960@bunghatta.ac.id>
Cc: Deepanaa Rajadurai <deepanaa.rajadurai@hindawi.com>

Thu, Jul 13, 2023 at 2:25 PM



Dear Dr. Hafrijal Syandri,

We have reviewed your manuscript "Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security " with ID No. 6639837. We found the following issues which need to be solved before moving on:

- **Relevance Check**

- - While checking the email address d.harfiandri@bunghatta.ac.id is verified one could you please update this in the system as well as in the cover letter?

“

This paper has been returned to draft for the reasons listed above. Please could you log into your account and address these points as soon as possible? Once completed, please could you resubmit the manuscript in the same manuscript number "6639837" and do not start a new submission.

Please log into your account to make the required updates. Follow the steps outlined below, and we kindly ask that you do not submit a brand new manuscript.

1. Click on the manuscript tile for Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security , which will show the status "Complete submission".
2. You will see all 4 submission steps. All the information will be pre-filled as you submitted the manuscript, so you will not need to complete the entire submission again. Click through the steps until you reach the area that needs updating.
3. If the manuscript file needs updating then go to step 4, and click the small bin icon next to the manuscript file. This will delete the file. You can then upload the new updated file.
4. When all necessary changes have been made, click the green "Submit manuscript" button at the bottom of step 4, and this will return the manuscript to the Hindawi team.

King regards,
Srimathi Anandan

This email was sent to syandri_1960@bunghatta.ac.id. You have received this email in regards to the account creation, submission, or peer review process of a paper submitted to a journal published by Hindawi Limited.

Hindawi Limited, 3rd Floor, Adam House, 1 Fitzroy Square, London, W1T 5HF, United Kingdom.

Hindawi respects your right to privacy. Please see our [privacy policy](#) for information on how we store, process, and safeguard your data.

[Unsubscribe](#)

hafrizal syandri <syandri_1960@bunghatta.ac.id>
To: Srimathi Anandan <srimathi.anandan@hindawi.com>

Thu, Jul 13, 2023 at 4:26 PM

Dear,
Srimathi Anandan

Thank you for your email; We verify that the author Harfiandri with the email: d.harfiandri@bunghatta.ac.id, has changed to harfiandri@bunghatta.ac.id; besides that, we have changed the email d.harfiandri@bunghatta.ac.id to harfiandri@bunghatta.ac.id in the cover letter.

Best regards
Hafrijal Syandri

[Quoted text hidden]

Srimathi Anandan <srimathi.anandan@hindawi.com>
Reply-To: Srimathi Anandan <srimathi.anandan@hindawi.com>
To: syandri_1960@bunghatta.ac.id
Cc: syandri_1960@bunghatta.ac.id, deepanaa.rajadurai@hindawi.com

Fri, Jul 14, 2023 at 12:47 PM

Dear Dr. Syandri,

Thank you for the response.

We will proceed with your manuscript for further processing.

**Best Regards,
Srimathi Anandan**

Srimathi Anandan
Editorial Screener

e. srimathi.anandan@hindawi.com



Hindawi

[Quoted text hidden]

, hafrizal syandri <syandri_1960@bunghatta.ac.id> wrote:

[Quoted text hidden]

[Quoted text hidden]

The contents of this email and any attachments are confidential and intended only for the person or entity to whom it is addressed. If you are not the intended recipient, any use, review, distribution, reproduction or any action taken in reliance upon this message is strictly prohibited. If you received this message in error, please immediately notify the sender and permanently delete all copies of the email and any attachments.

[Click here for translations of this disclaimer.](#)

Research Article

Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security

Azrita Azrita¹, Hafrijal Syandri^{1*}, Hazlina Ahamad Zakeri², Harfiandri Damanhuri¹, Netti Aryani³

¹*Faculty of Fisheries and Marine Science Universitas Bung Hatta, 25131, Padang-West Sumatra Province, Indonesia*

²*Faculty of Science and Marine Environment, Universiti Malaysia Terengganu, Malaysia.*

³*Faculty of Fisheries and Marine Science Universitas Riau, 28293, Pekanbaru-Riau Province, Indonesia*

*Corresponding author: syandri_1960@bunghatta.ac.id

Abstract:

Fish has become an irreplaceable source of animal protein food, especially among households with low socioeconomic conditions in rural and urban areas in Indonesia. This study analyzes the nutrient composition of three local Bagridae fish species in Indonesia. The nutrient composition results of *Hemibagrus nemurus*, *Hemibagrus wyckii*, and *Mystus nigriceps* revealed that the protein content was 24.26%, 22.57%, and 21.39% (% of dry weight), respectively. The total lipid content varied between 6.64% and 7.75%, whereas the crude ash content ranged from 1.59% to 2.30%. Regarding mineral contents, the calcium levels ranged from 1.49 to 1.66 mg/g, iron levels varied between 28.35 and 40.36 µg/g, and zinc levels ranged from 24.03 to 54.46 µg/g. The predominant amino acids found in the three species of Bagridae fish were glutamic acid, aspartic acid, alanine, arginine, and lysine, with their concentrations ranging from 9.10% to 24.34%. Among the fatty acids, C16:0 (palmitic acid) was the most abundant in all three species, accounting for 25.59% to 30.70% of the total fatty acids. Additionally, significant amounts of C18:1 (oleic acid), C18:0 (stearic acid), and C20:4 (arachidonic acid) were also detected as primary fatty acids. The calculated AI (atherogenic index) values in the three species of Bagridae fish ranged from 0.73 to 0.99, while the TI (thrombogenic index) values varied between 0.54 and 0.75. These results indicate that consuming the meat of these three freshwater species of Bagridae fish caught in the wild does not pose any health risks to consumers. They can be considered a safe and suitable food source with good nutritional quality.

Keywords: Nutrient composition, Amino acids, Fatty acids, Freshwater fish, Kampar Kanan River,

1. Introduction

Fish serve as a high-quality protein source that is essential for satisfying dietary protein requirements, especially in areas where other protein sources could be few or difficult to get [1,2]. Their accessibility can aid in ensuring a steady supply of food, particularly for communities whose main sources of income are fishing or aquaculture [3]. Fish can contribute to a more varied selection of protein sources in communities, resulting in a diet that is more nutritious and balanced. Dietary fish can treat nutrient deficits and enhance general nutritional health. Fish are generally considered safe and offer a lower risk of contamination with harmful substances compared to certain land-based animal products. According to [Ravichandran et al. \[4\]](#), a reliable indicator of the fish's quality, nutritional value, physiological status, and environment is said to be its chemical makeup, which is found in fish flesh.

Hemibagrus nemurus, locally known as "baung", *Hemibagrus wyckii*, known as "geso" and *Mystus negriceps*, known as "ingir-ingir," are three species of catfish found in Kampar Kanan River, Indonesia [5,6]. Due to their high market value, these species have been identified as significant economic resources for the rural community [7]. Therefore, the nutritional quality of fish meat in the study area is very important for analysis that is beneficial to human health.

Knowing the composition of fatty acids and amino acids in fish meat is an essential factor that should not be ignored [8,9,10]. This information provides valuable insight into the fish's nutritional quality and values of more nutritious and healthy foods [11]. In addition, a better understanding of the composition of fatty acids and amino acids can assist in selecting a suitable feed [12], developing a balanced diet [13], and maintaining optimal fish meat nutritional [14].

Omega-3 polyunsaturated fatty acids, particularly DHA and EPA derived from fish oil, exhibit anti-inflammatory and anti-gastrointestinal cancer properties, making them crucial as immune-boosting nutrients [15,16]. Studies have demonstrated that ω -3 and ω -6 polyunsaturated fatty acids (PUFA) exhibit beneficial effects in the management of cardiovascular diseases and cancers [17,18]. The fatty acid composition of polyunsaturated fatty acids (PUFAs) can differ among different types of fish, including both freshwater and marine species, as noted in studies conducted by [Rincón-Cervera et al. \[19\]](#) and [Sun et al. \[20\]](#). Amino acids (AAs) like cysteine,

arginine, tyrosine, glycine, proline, and serine play a vital role in illness and stress situations [21], including the prevention of inflammation and the repair processes in fish intestines [22].

In addition, knowledge of the atherogenicity index (AI) and thrombogenicity (TI) of fatty acids in three local freshwater fish species of Bagridae can contribute to improving dietary guidelines. This information, as suggested by [Wen et al. \[23\]](#) and [Duarte et al. \[24\]](#), demonstrates that the value of AI and TI can be used to develop recommendations aimed at improving heart health. These recommendations may involve selecting fish with favorable fatty acid profiles, allowing individuals to choose the most suitable fish species based on their specific needs [25,19]. Therefore, this study aims to examine the nutritional composition profile, mineral content, amino acids, and fatty acids, including AI and TI indices of three freshwater fish species, namely *H. nemurus*, *H. wyckii*, and *M. nigriceps*. These three types of fish are generally consumed by people who live around the Kampar Kanan River in Kampar Regency, Riau Province, Indonesia.

2. Material and methods

2.1. Materials

In this study, a total of 27 Bagridae fish belonging to three distinct species (*H. nemurus*, *H. wyckii*, and *M. nigriceps*) were examined. The specimens were obtained from local fishermen operating in the upper stretches of the Kampar Kanan River in Kampar Regency, Riau Province (Figure 1). The sampling took place between June and August 2022. They were verified by the Fisheries Department, Faculty of Fisheries and Marine Science Universitas Bung Hatta, Indonesia. The samples were placed in plastic bags and transported to the laboratory in an insulated icebox. The samples were then grouped based on their respective sampling locations.

2.2. Biometric measurement

Upon reaching the laboratory within 10 hours, the fish samples were subjected to gutted and washing. Each specimen was weighed (TW) and measured to determine the standard length (SL) as well as the depth or maximum height (H). The length measurement was taken from the mouth's tip to the end of the upper lobe of the caudal fin, representing the total body length. The height measurement involved a vertical assessment of the body's maximum height, excluding the fins. The condition factor (CF) was calculated using the formula $CF = (TW/SL^3) \times 100$.

2.3. Proximate, amino acid, and mineral content analysis

The proximate carcass composition was assessed following the AOAC standard methods [26]. The samples were dried until a constant weight was achieved at 105°C. The crude protein content was analyzed using the standard Kjeldahl method, which involves multiplying the nitrogen content by a factor of 6.25. The Soxhlet method with ether extraction was employed to analyze crude lipids. The ash content was obtained by incinerating the samples at 550°C for 16 hours. The amino acid composition was determined using a high-performance liquid chromatography (HPLC) system, which consisted of a water 1525 binary HPLC pump, 717 autosamplers (water®), and water 2475 multi λ fluorescence detector optics (with excitation at 250 nm and emission at 395 nm). The samples were hydrolyzed in triplicate with 6 N hydrochloric acid for 24 hours at 11°C [27].

For mineral data composition (Na, Mg, Ca, K, P, Fe, Cu, Mn, and Zn), the ashed of feed sample and carcass were dissolved in 1 ml of hydrochloric acid (35% v/v Suprapur® Merck). Furthermore, it was filtered with cellulose filter paper (Watchman No 1, International Ltd; Maidstone, UK) and diluted to an appropriate concentration for each elemental mineral. P levels were analyzed with a Perkin-Elmer AA spectrophotometer mod 3110 (Norwalk, CT. USA).

2.4. Fatty acids analysis

The body meat of fish was examined utilizing the gas chromatography-mass spectrometry (GC-MS) method. The modified Folch *et al.* (1957) method, as described by Rajion [28], was employed to perform the total fat extraction. This involved utilizing a solvent system consisting of chloroform and methanol in a 2:1 (v/v) ratio. The process of transmethylation was conducted using a solution of 14% methanolic boron trifluoride. The fatty acid composition of the meat was then analyzed at the Laboratory Accredited Testing Laboratory-LP-184-IDN.

The nutritional quality of lipids AI and TI was calculated using the equations [29].

$$AI = \frac{[(C12:0 + 4 \times C14:0 + C16:0)]}{\sum MUFA + \sum n-6 + \sum n-3}$$

$$TI = \frac{[(C14:0 + C16:0 + C18:0)]}{[(0.5 \times \sum MUFA) + (0.5 \times \sum n-6) + (3 \times \sum n-3) + (\sum n-3 / \sum n-6)]}$$

Where

AI = Atherogenic index

TI = Thrombogenic index

C12:0 = lauric acid

C14:0 = myristic acid

C16:0 = palmitic acid

C18:0 = stearic acid

Σ MUFA = sum concentrations of all monosaturated fatty acid

Σ n-6 = sum concentrations of n-6 polyunsaturated fatty acid

Σ n-3 = sum concentrations of n-3 polyunsaturated fatty acid

2.5. Data analysis

Data analysis was performed using the SPSS 16.0 software package (SPSS; Chicago, IL). The homogeneity of the data was assessed using the Levine test. One-way ANOVA was carried out to determine the parameters of proximate and amino acid composition, including the composition of fatty acids and nutritional quality of lipids AI and TI for each species, followed by Duncan's multiple range test post hoc [30]. The results are reported as mean values \pm standard errors for each parameter.

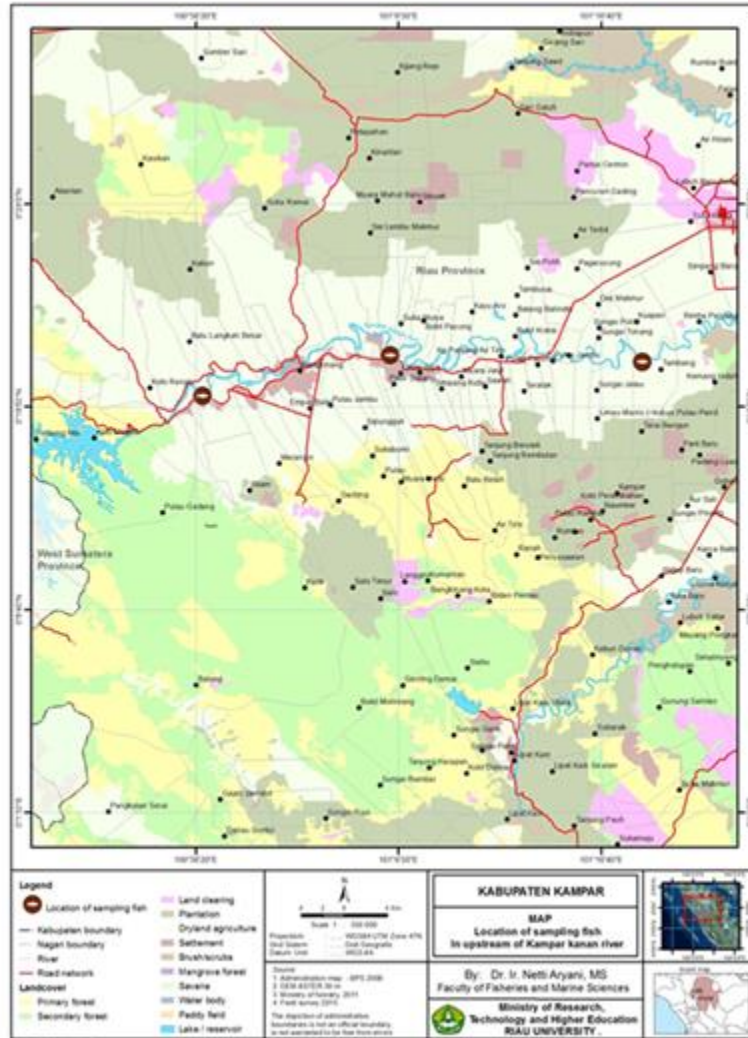


Figure 1. Map of Kampar Regency, Riau Province, and locations of three species of Bagridae fish sampling.

3. Results

Table 1 presents the average wet weight, standard length, width, and the results of meat nutritional composition analysis for three indigenous species of Indonesian freshwater Bagridae. The moisture content (% wet weight) ranges from 82.40% to 85.39%. Among the species examined, *M. nigriceps* exhibits the lowest protein content at 21.39%, while *H. nemurus* displays the highest value at 24.26%. However, *M. nigriceps* recorded has a higher mineral content such as iron and zinc.

Table 2 summarizes the fatty acid composition (% of total fatty acid) for the three species of Bagridae fish. C16:0 is the predominant fatty acid in the three species of Bagridae fish, with percentages ranging from 25.09% to 30.71%. The additional, prominent fatty acids found are C18:1, C18:0, and C22:6. *H. nemurus* exhibits a ω -6: ω -3 ratio below 1 (0.82), while *H. wyckii* and *M. nigriceps* have ratios of 1.11 and 1.02, respectively. The obtained PUFA/saturated (P/S) ratio varied between 0.61 and 0.76. In contrast, the AI values were 0.76 and 0.99, and the TI values were 0.54 and 0.75 for the respective cases (**Table 3**).

Table 4 presents the amino acid composition (% of total protein). Glutamic acid is the dominant amino acid, ranging from 19.29% to 24.34%, followed by aspartic acid, which ranges from 9.21% to 11.27%. Lysine content is consistent, ranging from 9.67% to 9.87%. In *H. nemurus*, the levels of various amino acids range from 0.86% to 24.35%. Similarly, in *H. wyckii*, the levels range from 0.76% to 21.58%, while in *M. nigriceps*, the levels range from 0.74% to 19.29%.

Table 1. Results of proximate analysis of three species of Bagridae fish

	Species		
	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
<i>Biometric measurements</i>			
Total weight (g)	389.99 ± 24.96 ^a	1,390.33 ± 168.29 ^b	17.57 ± 1.53 ^c
Standard length (cm)	28.16 ± 0.53 ^a	44.14 ± 1.98 ^b	10.29 ± 0.15 ^c
Width (cm)	8.44 ± 0.16 ^a	8.82 ± 0.39 ^b	2.57 ± 0.03 ^c
Condition factor	1.63 ± 0.08	1.58 ± 0.07	1.56 ± 0.08
<i>Proximate composition</i>			
Crude protein (% DW)	24.26 ± 0.87 ^a	22.57 ± 0.37 ^b	21.39 ± 0.15 ^c
Crude fat (% DW)	6.64 ± 0.03 ^a	7.47 ± 0.02 ^b	7.75 ± 0.40 ^c
Crude ash (% DW)	1.94 ± 0.02 ^a	2.30 ± 0.09 ^b	1.59 ± 0.02 ^c
Moisture (% WW)	82.40 ± 2.51	85.39 ± 2.36	83.75 ± 2.21
<i>Mineral composition</i>			
Macromineral (mg/g)			
Sodium	0.99 ± 0.00 ^a	1.61 ± 0.01 ^b	1.61 ± 0.02 ^c
Magnesium	1.12 ± 0.01 ^a	0.57 ± 0.02 ^b	1.15 ± 0.01 ^c
Calcium	1.66 ± 0.00 ^a	1.55 ± 0.02 ^b	1.49 ± 0.04 ^c
Potassium	0.71 ± 0.00 ^a	0.55 ± 0.02 ^b	0.43 ± 0.01 ^c
Phosphorous	7.03 ± 0.03 ^a	2.74 ± 0.02 ^b	6.45 ± 0.07 ^c
Microminerals (µg/g)			
Iron	28.30 ± 0.11 ^a	28.73 ± 0.08 ^b	40.36 ± 0.55 ^c
Copper	8.93 ± 0.03 ^a	7.46 ± 0.09 ^b	6.21 ± 0.32 ^c

Manganese	1.64 ± 0.02 ^a	2.64 ± 0.02 ^b	2.84 ± 0.02 ^c
Zinc	24.03 ± 0.45 ^a	24.61 ± 0.19 ^b	54.46 ± 0.17 ^c

Values are % ± SE; *Values in the same row with a different superscript a significantly different (p<0.05)

Table 2. Fatty acid composition of three species of Bagridae fish

Fatty acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
C12:0 (lauric acid)	1.97 ± 0.05 ^a	3.94 ± 0.01 ^b	4.27 ± 0.01 ^c
C14:0 (Meristic acid)	2.38 ± 0.03 ^a	2.72 ± 0.08 ^b	2.89 ± 0.01 ^c
C16:0 (Palmitic acid)	27.23 ± 0.06 ^a	30.70 ± 0.06 ^b	25.59 ± 0.01 ^c
C18:0 (Stearic acid)	16.17 ± 0.02 ^a	13.54 ± 0.02 ^b	10.64 ± 0.01 ^c
C20:0 (Arachidic acid)	0.23 ± 0.01 ^a	0.14 ± 0.01 ^b	0.37 ± 0.01 ^c
C16:1 (Palmitoleic acid)	1.78 ± 0.00 ^a	1.27 ± 0.04 ^b	4.93 ± 0.01 ^c
C18:1 (Oleic acid)	16.86 ± 0.01 ^a	16.18 ± 0.01 ^b	16.97 ± 0.01 ^c
C18:2 (Linoleic acid)	3.24 ± 0.02 ^a	3.84 ± 0.33 ^b	4.55 ± 0.001 ^c
C18:3 (Linolenic acid)	1.12 ± 0.01 ^a	0.82 ± 0.01 ^b	1.57 ± 0.00 ^c
C20:4 (Arachidonic acid)	12.13 ± 0.01 ^a	12.49 ± 0.09 ^b	12.41 ± 0.02 ^c
C20:5 (Eicosapentaenoic acid; EPA)	2.06 ± 0.08 ^a	2.34 ± 0.04 ^b	1.35 ± 0.01 ^c
C22:6 (Docosahexaenoic acid; DHA)	15.65 ± 0.01 ^a	11.56 ± 0.04 ^b	13.64 ± 0.02 ^c

Values are % ± SE; *Values in the same row with a different superscript a significantly different (p<0.05)

Table 3. Fatty acid ω-6/ω-3 and polyunsaturated/saturated fatty acid ratio of three species of Bagridae fish

	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
ω-6/ω-3	0.82 ± 0.00 ^a	1.11 ± 0.02 ^b	1.02 ± 0.01 ^c
Ratio of P/S	0.69 ± 0.01 ^a	0.61 ± 0.06 ^b	0.76 ± 0.00 ^c
AI	0.87 ± 0.01 ^a	0.99 ± 0.06 ^b	0.73 ± 0.00 ^c
TI	0.63 ± 0.01 ^a	0.75 ± 0.03 ^b	0.54 ± 0.00 ^c

P/S, polyunsaturated/saturated fatty acid; AI, Atherogenic index; TI, Thrombogenic index

Values are % ± SE; *Values in the same row with a different superscript a significantly different (p<0.05)

Table 4. Amino acid composition of three species of Bagridae fish

Amino acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
Aspartic acid	10.39 ± 0.01 ^a	9.61 ± 0.04 ^b	11.28 ± 0.02 ^c
Glutamic acid	24.34 ± 0.00 ^a	21.58 ± 0.41 ^b	19.29 ± 0.03 ^c
Serine	3.74 ± 0.00 ^a	5.40 ± 0.03 ^b	7.50 ± 0.06 ^c
Glycine	4.02 ± 0.00 ^a	5.77 ± 0.04 ^b	4.69 ± 0.02 ^c
Histidine	2.39 ± 0.00 ^a	2.90 ± 0.03 ^b	2.21 ± 0.02 ^c
Arginine	6.17 ± 0.01 ^a	8.27 ± 0.03 ^b	6.22 ± 0.03 ^c
Threonine	4.49 ± 0.01 ^a	4.19 ± 0.03 ^b	4.39 ± 0.04 ^c
Alanine	7.03 ± 0.00 ^a	6.04 ± 0.02 ^b	7.89 ± 0.01 ^c
Proline	2.40 ± 0.00 ^a	2.71 ± 0.01 ^b	2.21 ± 0.02 ^c
Tyrosine	3.91 ± 0.01 ^a	3.29 ± 0.06 ^b	3.82 ± 0.04 ^c
Valine	3.73 ± 0.00 ^a	4.02 ± 0.01 ^b	3.85 ± 0.04 ^c
Methionine	3.42 ± 0.00 ^a	1.96 ± 0.02 ^b	1.27 ± 0.03 ^c
Cystine	0.87 ± 0.00 ^a	0.76 ± 0.02 ^b	0.75 ± 0.03 ^c
Isoleucine	3.82 ± 0.001 ^a	3.96 ± 0.003 ^b	3.66 ± 0.002 ^c
Leucine	6.15 ± 0.000 ^a	6.69 ± 0.002 ^b	6.28 ± 0.004 ^c
Phenylalanine	5.13 ± 0.000 ^a	6.03 ± 0.005 ^b	5.18 ± 0.027 ^c
Lysine	9.61 ± 0.001 ^a	9.82 ± 0.026 ^b	9.87 ± 0.003 ^c

Values are % ± SE; *Values in the same row with a different superscript a significantly different (p<0.05)

4. Discussion

The results of this study provided significant new information about the nutritional makeup of the studied species of Bagridae fish. The fish's biometric measures yielded important information about its physical traits and health. Researchers are able to determine the average weight of fish at various lengths by examining the link between length and weight for a particular species. This knowledge is essential for understanding the trends in fish population growth, spotting potential shifts in growth rates over time, and assessing the general health and welfare of the fish [31]. A variety of factors, including sex, age, maturity level, size, level of

stomach fullness, sampling strategies, sample sizes, and environmental circumstances, have an impact on fish health and the parameters that determine length-weight connections in fish [32]. However, none of these factors are considered in our study. In general, according to the theory put forth by [Bagenal and Tesch \[33\]](#), heavier fish of a given length are in better physiological condition. The condition factor is also used in fisheries science to compare the health, fatness, and condition of fish [34]. [Ujjania et al. \[35\]](#) stated that a condition factor of ≥ 1 is favourable, indicating a good level of nutrition and a suitable habitat. Thus, from the findings in this study, it may be said that the Kampar Kanan River provides a suitable habitat for the fish, ensuring their accessibility and availability for community consumption.

The fish samples exhibited good nutritional profiles, with high amounts of proteins and lipids as well as moisture content contributing to their overall nutritional value, according to the results of proximate analysis and minerals composition (Table 1). These results imply that the fish species studied may provide a possible source of nutrients to solve issues with food security. The study of the minerals composition revealed the existence of other critical minerals in the fish samples in addition to proteins and lipids. For total health and wellbeing, enough mineral consumption is required. Fish with a high level of crude ash can help increase dietary mineral intake and may even contain vital minerals including phosphorus (P), calcium (Ca), magnesium (Mg) and trace elements such as iron (Fe) and zinc (Zn). All the three Bagridae fish are significantly differ from each other in terms of their crude ash content and minerals profile (Table 1). However, in general, P and Ca are the two highest minerals found in all fish. Both these macrominerals are good for human's bone and teeth health, as key components of hydroxyapatite, the mineral matrix that provides strength and structure to bones and teeth [36]. P also contributes to a number of activities that include the synthesis and utilisation of ATP, as well as the breakdown of carbs, proteins, and fats, helping the body's ability to produce and use energy [37]. Cellular communication, enzyme activation, and the control of cell membrane integrity are all affected by the presence of P [38]. Ca on the other hand, is crucial for muscle contraction and relaxation, nerve impulse transmission, and neurotransmitter release, enabling proper muscle function [39]. It also aids in the activation of clotting factors and the production of fibrin, a protein that forms a meshwork to halt bleeding from wounds or traumas [40].

In the current study, the meat of three species of Bagridae fish from the Kampar Kanan River Indonesia was found to contain abundance of microminerals such as Fe and Zn. Insufficient intake of Fe and Zn has been linked to major developmental health problems, such as Fe deficiency which is one of the major causes of anemia in pregnant and lactating women, especially in rural communities [41,42]. Lacking in Zn, on the other hand, can lead to numerous immunological disorders, such as metabolic and chronic illnesses, as well as infectious diseases including respiratory infections, malaria, HIV, or tuberculosis [43]. *M. nigriceps* that has been recorded exhibits higher mineral content, including iron and zinc. This species is categorized as a small indigenous fish (SIF) with a maximum standard length of approximately 10.29 cm. According to Mohanty et al. [44], SIFs, in general, are known to be rich in Ca, Fe, and manganese (Mn). The variation in mineral content among the three species of Bagridae fish is comparable to that of *Clarias gariepinus* and *Tilapia zilli* meat from the Owan River in Edo State, Nigeria [13].

In general, fish consists of 70-84% moisture [45]. High moisture content is often associated with freshness in fish. Compared to fish that has been processed or stored, fresh fish often has a higher moisture content, the factor that contributes to a moist and succulent texture. Fish with high moisture content may retain more water-soluble nutrients during cooking or processing as proven by Gupta et al. [46] with green leafy vegetables. This can help preserve the nutritional value of the fish, ensuring that important vitamins and minerals are retained. However, due to the high moisture content in fish, a processing technique that changes the moisture content will also change the fish's properties, such as the texture and protein gelation [47].

The amount of crude fats in the fish is a good indicator of how much lipids or fats are present. In addition to acting as transporters for fat-soluble vitamins, fats offer a concentrated source of energy to the body. Additionally, they improve the fish's flavour, texture, and palatability [48]. Numerous health advantages, including improved cardiovascular health, improved brain function, and anti-inflammatory effects, have been linked to specific fatty acids present in fish, particularly omega fatty acids. So a fish with a high amount of crude fat might be good, especially if it has good fatty acids. From the findings, all three Bagridae fish had a total fat content of around 7-8% by weight (Table 1). According to Fuentes et al. [49], fish with lower fat content exhibit higher water content, which is also shown in our study. On the other hand,

Nurnadia et al. [50] reported that the fish species can be grouped into different categories, including high-fat fish (>8%), medium-fat fish (4-8%), low-fat fish (2-4%), and lean fish (<2%). According to the established classification, the three indigenous Bagridae fish species are characterized as having a moderate level of fat content, which is reflected by the high levels of DHA and EPA in the meat of *H. nemurus*, *H. wyckii*, and *M. nigriceps* fish (Table 2). It is still unclear whether consuming the fish meat of these three species can indirectly help lower blood pressure, reduce inflammation in blood vessels, improve lipid profiles by increasing levels of good cholesterol (HDL) and decreasing levels of bad cholesterol (LDL), and reduce blood clotting in individuals. Nonetheless, the fat content of fish is determined by various factors, such as species, habitat ecosystems, feeding patterns, and other biological attributes, including the distinctive flavor characteristics of the fish [8,49,51,52].

Each of the three Bagridae fish species analyzed in the study contained arachidonic acid (C20:4), which serves as a precursor for synthesizing prostaglandins and leukotrienes [53]. Leukotrienes are crucial in allergic responses, inducing smooth muscle contraction and increasing vascular permeability [54]. In this study, the arachidonic acid content in *H. nemurus*, *H. wyckii*, and *M. nigriceps* was determined to be 12.13%, 12.49%, and 12.41%, respectively. Conversely, the levels of arachidonic acid in *Channa striatus*, *Channa micropeltes*, and *Channa lucius* were reported as 19.02%, 4.71%, and 12.41%, respectively [55]. According to Carballo-Casla et al. [56], increased consumption of marine omega-3 fatty acids, such as EPA and DHA, has been linked to a decreased risk of pain development in older individuals. Additionally, DHA and EPA have demonstrated a preventive effect on human coronary arteries and Alzheimer's disease [57,58]. In this study, our hypothesis is that the levels of EPA and DHA found in the meat of three species of Bagridae fish in the Kampar Kanan River, Indonesia, may also have the potential in reducing muscle pain and inflammation. Additionally, as a primary food source, fish plays a role in the prevention and treatment of chronic pain [59]. As determined by the current study, the ω -6: ω -3 ratios of three Bagridae fish species ranged from 1:0.8 to 1:0.97. These varying ω -6: ω -3 ratios can have diverse implications for treating or preventing diseases [60]. It was found that all three fish species had ω -6: ω -3 ratios within the recommended range for a typical Indonesian diet. In contrast, a typical Western diet is characterized by a high ω -6: ω -3 ratio of 10:1 to 20:1, while a Mediterranean diet has a ratio of 4:1 to 5:1 [61]. The P/S ratio

analysis revealed that all three Bagridae fish species were considered an average source of PUFA. Additionally, it has been reported that marine fish serve as a good source of PUFA [62].

The atherogenic index (AI) and thrombogenic index (TI) indices are directly influenced by the levels of C14:0, C16:0, and C18:0, all of which are known to promote thrombogenicity [24]. AI and TI indices indicate the potential to induce platelet aggregation. A lower value in AI and TI indicates a more significant protective potential against coronary artery disease [63]. The Food and Agriculture Organization (FAO) and the World Health Organization (WHO) have recommended AI and TI values ranging from 0.4 to 0.5 for promoting human health [64]. The AI and TI values of various marine fish species, such as Sardine (*Sardinella hualiensis*), being 0.60 and 0.20, respectively, and Mackerel (*Scomber japonicus*) having values of 0.48 and 0.24, respectively [65]. In the current study, the observed AI values in three freshwater species of Bagridae fish ranged from 0.73 to 0.99, while the TI values varied between 0.54 and 0.75. This finding is associated with a notable disparity in the values of saturated fatty acids (SFA) among the three species of Bagridae fish. Based on these findings, we certify that consuming the meat of three freshwater species of Bagridae fish caught in the wild does not endanger consumers' health and can serve as a suitable source of food safety and nutritional quality.

A high value for crude protein shows that the fish contains a considerable amount of protein. Since fish protein is easier to digest than protein found in meat and includes less collagen fibre, the body can use its protein more effectively [66]. The body's tissues need protein to develop, mend, and remain healthy. The absence of amino acids and fatty acids elements will impede the recovery process [67]. Protein is a crucial ingredient for human health, contributing to a number of physiological functions like immunological function, muscular growth, and enzyme synthesis. Since protein is a crucial part of a balanced diet, fish with a high crude protein content, as also been shown by the three Bagridae fish studied (Table 1), are regarded as being nutritionally useful. According to Pawar and Sonaware [68], protein and amino acid content can directly indicate the nutritional quality of meat. In Table 4, *H. wykii* had higher quantities of essential amino acids (EAA) than *H. nemurus* and *M. nigriceps*. Since EAA cannot be synthesised by the body yet are necessary for physiologic function, they are significant for humans. They are thus derived through EAA-rich foods such as fish. Among the EAA, lysine (Lys) was found to be present in the highest amount in all three species. Lys is necessary for tissue growth and repair

[69]. Furthermore, Lys can reduced stress-induced anxiety [70] and has antiviral properties [71]. Lys also plays a role in metabolic activities, including the creation of carnitine for mitochondrial energy generation and the conversion of fatty acids into energy [72].

More than 60% of the amino acid composition in all three Brigade fish studied comes from the non-essential amino acids (NEAA), particularly glutamic acid (Glu) (Table 4). [Watford \[73\]](#) and [Mohanty et al. \[62\]](#), however, classified Glu in addition to Arg, glycine (Gly), proline (Pro) and taurine as conditionally EAA, amino acids which are usually not essential except in times of illness and stress. Glu and its amine, glutamine (Gln) are highly abundant amino acids found in fish in the free and protein-bound forms [74]. As functional amino acids, Glu and Gln are significant metabolic fuels for fish tissues, such as the gut, liver, kidneys, and skeletal muscle, and they play crucial roles in protein synthesis as well as glutathione formation and anti-oxidative responses [75]. In humans, Glu functions as an excitatory neurotransmitter involved in memory, cognition and mood regulation [76]. Glu is also responsible for the “umami” or savory taste in protein-rich food such as fish [77]. The perception of this taste enhances food flavour and, when used as salt, such as monosodium glutamate, stands out for having only a third of the sodium found in table salt. This salt is therefore thought to be an option to help people consume less sodium [78]. Additionally, umami chemicals have been linked to the control of obesity and the post-meal feeling of satiety [79]. However, the association between the Glu content and umami taste in the three Bagridae fish studied has yet to be determined and proven. Other amino acids found abundant in the three fish also play an important physiologic as well biochemical functions. The NEAA, arginine (Arg), for example, are involved in protein synthesis, creatine formation, and various other biochemical processes within the body. The EAAs, leucine (Leu), phenylalanine (Phe) and methionine (Met), play important roles in the synthesis of nitric oxide (NO), a molecule that can expand blood vessels [75].

4. Conclusions

The present study offers insightful information about the fatty acid and amino acid compositions, biometric measurements, proximate analysis, and mineral composition of three local freshwater species of Indonesian Bagridae fish. The findings show their potential as a nutrient-dense, sustainable food source that can support efforts to increase food security. These

fish species might be incorporated into the local diet to increase nutrient intake and alleviate nutritional inadequacies, which would ultimately improve Indonesia's overall food security status.

Data Availability

The data utilized in this research has not been previously released or published in any form. The data sets employed and/or analyzed during the present study can be obtained by contacting the corresponding author.

Conflicts of Interest

The authors confirm that they do not have any conflicting interests.

Acknowledgements

The author acknowledges and expresses gratitude for the financial support received from the Ministry of Education, Culture, Research, and Technology of the Republic of Indonesia, specifically through grant no. 076/E5/PG. 02.00. PT/2022, which funded the research. The author affirms that there is no conflict of interest associated with the utilization of these research funds. Furthermore, the author extends appreciation to the students and fishermen involved in the research conducted in the upper reaches of the Kampar Kanan River, Indonesia.

References

- [1] A.G. Hasselberg, I.Aakre, J. Scholtens *et al.*, “Fish for food and nutrition security in Ghana: Challenges and opportunities,” *Global Food Security*. vol 26: 100380, 2020 <https://doi.org/10.1016/j.gfs.2020.100380>
- [2] N.Tran, U.P, Rodriguez, C.Y.Chan., *et al.*, “Future cenarios of fish supply and demand for food and nutrition security in Bangladesh: An analysis with the AsiaFish model”, *Aquaculture*. Vol.568: 739288, 2023. <https://doi.org/10.1016/j.aquaculture.2023.739288>
- [3] C.Z. Levkoe, K. Lowitt, C. Nelson, (2017). “Fish as food: Exploring a food sovereignty approach to small-scale fisheries”. *Marine Policy*. vol 85: pp.65-70, 2023. <https://doi.org/10.1016/j.marpol.2017.08.018>
- [4] M. Ravichandran, N.S. Hettiarachchy, V. Ganesh *et al.*, “Enhancement Of Antimicrobial Activities Of Naturally Occurring Phenolic Compounds By Nanoscale Delivery Against Listeria Monocytogenes, Escherichia Coli O157:H7 And Salmonella Typhimurium In Broth And Chicken Meat System”. *Journal of Food Safety*. vol 31, no. 4, pp. 462-471,2011. <https://doi.org/10.1111/j.1745-4565.2011.00322.x>

- [5] N. Aryani, I. Suharman, Azrita, *et al.*, “Diversity and distribution of fish fauna of upstream and downstream areas at Koto Panjang Reservoir, Riau Province, Indonesia”. *F1000 Research*. vol 8, pp. 1435. 2020. <https://f1000research.com/articles/8-1435/v2>
- [6] Syafrialdi, Dahelmi., D.I. Roesma, *et al.*, “Length-weight relationships and condition factor of Two-Spot Catfish (*Mystus nigriceps* [Valenciennes, 1840]) Pisces, Bagridae], from Kampar Kanan River and Kampar Kiri River in Indonesia”. *Pakistan Journal of Biological Science*, vol. 23, no. 12, pp. 1636-1642, 2020. <https://scialert.net/fulltext/?doi=pjbs.2020.1636.1642>
- [7] N. Aryani, “Native species in Kampar Kanan River, Riau Province Indonesia. *International Journal of Fisheries and Aquatic Studies*”, vol.2, no.5, pp. 213-217. 2015. <https://www.fisheriesjournal.com/archives/2015/vol2issue5/PartD/2-5-63.pdf>
- [8] Azrita, H. Syandri, N. Aryani, *et al.*, “The utilization of new products formulated from water coconut, palm sap sugar, and fungus to increase nutritional feed quality, feed efficiency, growth, and carcass of gurami sago (*Osphronemus goramy* Lacepède, 1801) juvenile”. *F1000 Research*. vol. 10. :1121, pp. 1-16, 2021. <https://f1000research.com/articles/10-1121/v1>
- [9] Azrita, H. Syandri, N. Aryani *et al.*, “Effect of feed enriched by products formulated from coconut water, palm sap sugar, and mushroom on the chemical composition of feed and carcass, growth performance, body indices, and gut micromorphology of giant gourami, *Osphronemus goramy* (Lacepède, 1801), juveniles”. *F1000 Research*. vol 12. No.140, pp. 1-19, 2023. <https://f1000research.com/articles/12-140/v1>
- [10] L.F. Montenegro, A.M. Descalzo, R. Rizzo, *et al.*, “Improving the antioxidant status, fat-soluble vitamins, fatty acid composition, and lipid stability in the meat of Grass carp (*Ctenopharyngodon idella* Val) fed fresh ryegrass (*Lolium multiflorum* Lam)”. *Aquaculture*, 553, 738067, 2022. <https://doi.org/10.1016/j.aquaculture.2022.738067>
- [11] C. Bene, R. Arthur, H. Norbury *et al.*, “Contribution of Fisheries and Aquaculture to Food Security and Poverty Reduction: Assessing the Current Evidence”. *World Development*, 79: 177–196. 2016. <http://dx.doi.org/10.1016/j.worlddev.2015.11.007>
- [12] N. Aryani, I. Suharman, S. Hasibuan, *et al.*, “Fatty acid composition on diet and carcasses, growth, body indices and profile serum of Asian redbtail catfish (*Hemibagrus nemurus*) fed a diet containing different levels of EPA and DHA”. *F1000Research*. 11:1409.pp.1-23, 2023. <https://doi.org/10.12688/f1000research.126487.2>
- [13] E.N. Kingsley, O.O. Cyril, O.I. Patience, “Potential contribution of selected wild fish species to the minerals intake of Pregnant and Lactating Women, Children and Adults in Rural Riverine Communities of Edo State: Insights and Outcomes”. *Measurement: Food*, 8, 100063 : 1-7, 2022. <https://doi.org/10.1016/j.meafoo.2022.100063>
- [14] A.A. Sausa, L. Nora, D.L.A. Lopes, *et al.*, Vegetable choline in feed for Nile tilapia (*Oreochromis niloticus*) raised in a biofloc technology system (BFT): Biofloc composition, chemical composition, and fatty acid profiles in meat“. *Aquaculture*. vol. 545. 737174, 2021. <https://doi.org/10.1016/j.aquaculture.2021.737174>
- [15] A.M. Eltweri, A.L. Thomas, M. Metcalfe, *et al.*, 2017. Potential applications of fish oils rich in omega-3 polyunsaturated fatty acids in the management of gastrointestinal cancer. *Clinical Nutrition*. vol. 36, no. 1, pp. 65-78. 2017. <https://doi.org/10.1016/j.clnu.2016.01.007>

- [16] P.C. Haycock, M.C. Borges, K. Burrows, *et al.*, “The association between genetically elevated polyunsaturated fatty acids and risk of cancer”. *eBiomedicine*. vol.91: 104510, 2023. <https://doi.org/10.1016/j.ebiom.2023.104510>
- [17] E.K. Aglago, I. Huybrechts, N. Murphy, *et al.*, “Consumption of Fish and Long-chain n-3 Polyunsaturated Fatty Acids Is Associated With Reduced Risk of Colorectal Cancer in a Large European Cohort”. *Clinical Gastroenterology and Hepatology*. vol. 18, no. 3, pp. 654-666.e6. 2020. <https://doi.org/10.1016/j.cgh.2019.06.031>
- [18] H. Jiang, X. Shi, T.Y. Fan, *et al.*, “Dietary omega-3 polyunsaturated fatty acids and fish intake and risk of age-related macular degeneration” . *Clinical Nutrition*. vol.40, no. 12, pp: 5662-5673. 2021. [https://www.clinicalnutritionjournal.com/article/S0261-5614\(21\)00474-X/fulltext](https://www.clinicalnutritionjournal.com/article/S0261-5614(21)00474-X/fulltext)
- [19] M.A. Rincón-Cervera, NV. González-Barriga, R. Valenzuela, *et al.*, “Profile and distribution of fatty acids in edible parts of commonly consumed marine fishes in Chile”, *Food Chemistry*, vol. 274, pp. 123-129, 2018. <https://doi.org/10.1016/j.foodchem.2018.08.113>
- [20] S. Sun, T. Ren, X. Li., *et al.*, “Polyunsaturated fatty acids synthesized by freshwater fish: A new insight to the roles of elovl2 and elovl5 in vivo”, *Biochemical and Biophysical Research Communications*, vol. 532, pp. 414-419, 2020. <https://doi.org/10.1016/j.bbrc.2020.08.074>
- [21] M.B. Behyar, M. Hasanzadeh, F. Seidi, *et al.*, “Sensing of amino acids: Critical role of nanomaterials for the efficient biomedical analysis”, *Microchemical Journal*, vol. 188, pp. 108452, 2023 <https://doi.org/10.1016/j.microc.2023.108452>
- [22] P. Shi, K. Liao, J. Xu, *et al.*, “Eicosapentaenoic acid mitigates palmitic acid-induced heat shock response, inflammation and repair processes in fish intestine”, *Fish & Shellfish Immunology*, vol. 124, pp. 362-371, 2022. <https://doi.org/10.1016/j.fsi.2022.04.011>
- [23] J. Wen, L. Zeng, Y. Xu, *et al.*, “Proximate composition, amino acid and fatty acid composition of fish maws”, *Natural Product Research: Formerly*, vol. 30, no. 2, pp. 214-217, 2015. <https://doi.org/10.1080/14786419.2015.1040790>
- [24] F.O.S. Duarte, F.G. de Paula, C.S. Prado, *et al.*, “Better fatty acids profile in fillets of Nile Tilapia (*Oreochromis niloticus*) supplemented with fish oil”, *Aquaculture*, Vol. 534, pp. 736241, 2021. <https://doi.org/10.1016/j.aquaculture.2020.736241>
- [25] K.L. Weaver, P. Ivester, J.A.Chilton, *et al.*, “The Content of Favorable and Unfavorable Polyunsaturated Fatty Acids Found in Commonly Eaten Fish”, *Journal of the American Dietetic Association*, vol. 108, No. 7 , pp. 1178–1185, 2008. <https://doi.org/10.1016/j.jada.2008.04.023>
- [26] AOAC, “Official Methods of Analysis. 15th Edn., Association of Official Analytical Chemists (AOAC)”, Washington, DC., USA.1990. Reference Source
- [27] S.A.Cohen, “Amino acid analysis using pre-column derivatization with 6-aminoquinoly1-N-hydroxysuccinimidyl carbamate”, *Protein Sequencing Protocols*, vol. 211, pp. 143-54, 2003. <https://doi.org/10.1385/1-59259-047-0:039>
- [28] M.A. Rajion, J.G. McLean, R.N. Cahill, “Essential fatty acid metabolism in the fetal and neonatal lamb”, *Australian Journal of Biological Sciences*, vol. 38, no. 1, pp. 33-40, 1985. [Essential fatty acids in the fetal and newborn lamb - PubMed \(nih.gov\)](https://pubmed.ncbi.nlm.nih.gov/)

- [29] T.L. Ulbricht, D.A. Southgate, “Coronary heart disease: Seven dietary factors”, *The Lancet*, Vol 338, no. 8773, pp. 985–992, 1991. [https://doi.org/10.1016/0140-6736\(91\)91846-M](https://doi.org/10.1016/0140-6736(91)91846-M)
- [30] D.B. Duncan, “Multiple ranges and multiple F tests”, *Biometrics*, vol. 11, pp. 1–42, 1955. <https://doi.org/10.2307/3001478>
- [31] R. Froese, (). “Cube law, condition factor and weight–length relationships: History, meta-analysis and recommendations”. *Journal of Applied Ichthyology*, vol. 22, no. 4, pp. 241–253, 2006. <https://doi.org/10.1111/j.1439-0426.2006.00805.x>
- [32] O.O.Famoofo, & W.O. Abdul, “Biometry, condition factors and length-weight relationships of sixteen fish species in Iwopin fresh-water ecotype of Lekki Lagoon, Ogun State, Southwest Nigeria”, *Heliyon*, vol. 6, no. 1, 2020. <https://doi.org/10.1016/j.heliyon.2019.e02957>
- [33] T.B. Bagenal, F.W. Tesch, “Age and Growth. In: Bagenal, T., Ed., Methods for Assessment of Fish Production in Fresh Waters”, *Blackwell Science Publications*, pp. 101-136,1978.
- [34] E.O. Ahmed, M.E. Ali, A.A. Aziz, “Length-weight relationships and Condition factors of six fish species in Atbara River and Khashm el-girba Reservoir, Sudan”, *International Journal of Agriculture Sciences*, vol. 3, no.1, pp. 65–70, 2011. <http://dx.doi.org/10.9735/0975-3710.3.1.65-70>
- [35] N.C. Ujjania, M.P.S. Kohli, L.L. Sharma, “Length-weight relationship and condition factors of Indian major carps (C. catla, L. rohita and C. mrigala) in Mahi Bajaj Sagar, India”, *Research Journal of Biology*, vol. 2, no.1, pp. 30-36, 2012. <https://doi.org/10.21077/ijf.2017.64.special-issue.76263-27>
- [36] J.P. Bonjour, “Calcium and phosphate: A duet of ions playing for bone health”, *Journal of the American College of Nutrition*, vol. 30, no. 5, pp. 438S-48S, 2011. <https://doi.org/10.1080/07315724.2011.10719988>
- [37] T. Tian, Y. Chu, Y. Yang, *et al.*, “Phosphates as Energy Sources to Expand Metabolic Networks”, *Life*, Vol. 9, no. 2, pp. 43, 2019. <https://doi.org/10.3390/life9020043>
- [38] K. Kritmetapak, R. Kumar, “Phosphate as a Signaling Molecule”, *Calcified tissue international*, Vol. 108, no. 1, pp. 16, 2021. <https://doi.org/10.1007/s00223-019-00636-8>
- [39] M.W. Berchtold, H. Brinkmeier, M. Müntener, “Calcium Ion in Skeletal Muscle: Its Crucial Role for Muscle Function, Plasticity, and Disease”, *Physiological Reviews*, Vol. 80, no. 3, 2000. <https://doi.org/10.1152/physrev.2000.80.3.1215>
- [40] S. Singh, J. Dodt, P.Volkers, *et al.*, “Structure functional insights into calcium binding during the activation of coagulation factor XIII A”. *Scientific Reports*, vol. 9, no. 11324, 2019. <https://doi.org/10.1038/s41598-019-47815-z>
- [41] A.A. Hassan, A.I. Mamman, , S. Adaji, *et al.*, “Anemia and iron deficiency in pregnant women in Zaria, Nigeria”, *Sub-Saharan African Journal of Medicine*, vo.1, no.1, pp. 36-39, 2014. <http://dx.doi.org/10.4103/2384-5147.129311>
- [42] I.M. Okafor, D.C. Okpokam, A.B. Antai, *et al.*, “Iron status of pregnant women in rural and urban communities of cross river state, South-South Nigeria”. *Nigerian journal of physiological sciences : official publication of the Physiological Society of Nigeria*, vol. 31, pp. 121-125, 2016. <https://pubmed.ncbi.nlm.nih.gov/28262847/>
- [43] M. Maywald, L. Rink, “Zinc in Human Health and Infectious Diseases”, *Biomolecules*, vol. 12 no. 12, pp. 1748, 2022. <https://doi.org/10.3390/biom12121748>

- [44] B.P. Mohanty, T.V. Sankar, S. Ganguly, *et al.*, “Micronutrient composition of 35 food fishes from India and their significance in human nutrition”, *Biological Trace Element Research*, vol. 174, pp. 448-458, 2016. <https://doi.org/10.1007/s12011-016-0714-3>
- [45] B. Abraha, H. Admassu, A. Mahmud, *et al.*, “Effect of processing methods on nutritional and physico-chemical composition of fish: a review”, *Food Processing and Technology*, vol. 6, no. 4, pp. 376-382, 2018. <https://doi.org/10.15406/mojfpt.2018.06.00191>
- [46] S. Gupta, B.S. Gowri, A.J. Lakshmi, *et al.*, “Retention of nutrients in green leafy vegetables on dehydration”, *Journal of Food Science and Technology*, vol. 50, no. 5, pp. 918-925, 2013. <https://doi.org/10.1007/s13197-011-0407-z>
- [47] S. NurSyahirah, A. Rozzamri, “Effects of frying on fish, fish products and frying oil – a review”, *Food Research*, vol. 6, no. 5, pp. 14-32, 2022. [https://doi.org/10.26656/fr.2017.6\(5\).608](https://doi.org/10.26656/fr.2017.6(5).608)
- [48] F. Shahidi, A. Hossain, “Role of Lipids in Food Flavor Generation”. *Molecules*, vol. 27, no.15, 2022. <https://doi.org/10.3390/molecules27155014>
- [49] E.F. Fernandes, , M.A.S. Vasconcelos, , M.d.A. Ribeiro, *et al.*, “Nutritional and lipid profiles in marine fish species from Brazil”, *Food Chemistri*, vol. 160, pp. 67-71, 2014. <https://doi.org/10.1016/j.foodchem.2014.03.055>
- [50] A.A. Nurnadia, , A. Azrina, I. Amin, “Proximate composition and energetic value of selected marine fish and shellfish from the west coast of peninsular Malaysia”. *International Food Research Journal*. Vol 18, pp. 137-148, 2011. <https://core.ac.uk/download/pdf/153807129.pdf>
- [51] B.P. Mohanty, A. Mahanty, S. Ganguly, *et al.*, “Nutritional composition of food fishes and their importance in providing food and nutritional security”, *Food Chemistry*, vol. 293, pp. 561-570, 2019. <https://doi.org/10.1016/j.foodchem.2017.11.039>
- [52] F. Jabeen, , A.A. Chudhry, “Chemical composition and fatty acid profiles of three freshwater species”, *Food Chemistry*, vol. 125, pp. 991-996, 2011. <https://doi.org/10.1016/j.foodchem.2010.09.103>
- [53] S.C. Dyall, , L. Balas, , N.G. Bazan, *et al.*, “Polyunsaturated fatty acids and fatty acid-derived lipid mediators: Recent advances in the understanding of their biosynthesis, structures and functions”, *Progress in Lipid Research*. vol. 86, pp. 101165, 2022. <https://doi.org/10.1016/j.plipres.2022.101165>
- [54] N. Kim, , U.D.Sohn, , V. Mangannan, *et al.*, “Leukotrienes in Acetylcholine-Induced Contraction of Esophageal Circular Smooth Muscle in Experimental”, *Gastroenterology*, vol. 112, no.5, pp. 1548–1558, 1997. [https://doi.org/10.1016/S0016-5085\(97\)70036-2](https://doi.org/10.1016/S0016-5085(97)70036-2)
- [55] A. Zuraini, M.N. Somchit, M.H. Solihah, *et al.*, “Fatty acid and amino acid composition of three local Malaysian Channa spp. Fish”, *Food Chemistry*, vol. 97, no. 4, pp. 674-678, 2005. <https://doi.org/10.1016/j.foodchem.2005.04.031>
- [56] Carballo-Casla, Garcia-Esguines, J.R. Banegas, *et al.*, “Fish consumption, omega-3 fatty acid intake, and risk of pain: the Seniors-ENRICA-1 cohort”, *Clinical Nutrition*, vol. 41, no. 11, pp. 2587-2595, 2022. <https://doi.org/10.1016/j.clnu.2022.09.007>
- [57] D. Swanson, R. Block, , S.A.Mousa, “Omega-3 Fatty Acids EPA and DHA: Health Benefits Throughout Life”. *Advances in Nutrition*, vol. 3, no.1, pp. 1-7,2012. <https://doi.org/10.3945/an.111.000893>

- [58] R.P.Mason , S.C.R. Sherratt, , L.H. Eckel, “Omega-3-fatty acids: Do they prevent cardiovascular disease?”, *Best Practice & Research Clinical Endocrinology & Metabolism*, vol.37, no 3, 101681, 2023. <https://doi.org/10.1016/j.beem.2022.101681>.
- [59] M. Rondanelli, M.A. Faliva, , A. Miccono, *et al.*, “Food pyramid for subjects with chronic pain: foods and dietary constituents as anti-inflammatory and antioxidant agents”, *Nutrition Research Reviews*, vol. 31, no. 1, pp. 131-151, 2018. <https://doi.org/10.1016/j.endien.2020.01.010>
- [60] D. Karageorgou, U. Rova, , P. Christakopoulos, *et al.*, “Benefits of supplementation with microbial omega-3 fatty acids on human health and the current market scenario for fish-free omega-3 fatty acid”, *Trends in Food Science & Technology*, vol. 136, pp. 169-180, 2023. <https://doi.org/10.1016/j.tifs.2023.04.018>
- [61] J. M. Seddon, , J. Cote, , N. Davis, *et al.*, “Progression of age-related macular degeneration. Progression of age-related macular degeneration”. *Arch Ophthalmol*. Vol. 121, no. 6, pp. 785-792, 2003. <https://doi.org/10.1001/archophth.121.6.785>
- [62] B.P. Mohanty, A. Mahanty, , S. Ganguly, *et al.* “Amino Acid compositions of 27 food fishes and their importance in clinical nutrition”, *Journal Amino Acids*, vol. 2014, 2014. <https://doi.org/10.1155/2014/269797>
- [63] H. Turan, G. Sonmez, Y. Kaya, “Fatty acid profile and proximate composition of the thornback ray (*Raja clavata*, L. 1758) from the Sinop coast in the Black Sea”, *Journal of Fisheries Sciences*, vol. 1, no. 2, pp. 97–103, 2007. <https://doi.org/10.3153/jfscm.2007012>
- [64] FAO/WHO: Fat and Oils in Human Nutrition: Report of a Joint Expert Consultation. FAO/WHO, Rome, Italy.
- [65] C.E. Fernandes, M.A.da.S. Vasconcelos, , M.L.A. Sarubbo, *et al.*, Nutritional and lipid profiles in marine fish species from Brazil, *Food Chemistry*, vol. 160, no. 67-71, 2014. <https://doi.org/10.1016/j.foodchem.2014.03.055>
- [66] L. Day, J.A. Cakebread, S.M. Loveday, “Food proteins from animals and plants: Differences in the nutritional and functional properties”, *Trends in Food Science & Technology*, vol. 119, pp. 428-442, 2022. <https://doi.org/10.1016/j.tifs.2021.12.020>
- [67] L. Wu, A. Wang, , R. Shen, *et al.*, “Effect of processing on the contents of amino acids and fatty acids, and glucose release from the starch of quinoa”, *Food Science & Nutrition*, vol. 8, no. 9, pp. 4877-4887, 2020. <https://doi.org/10.1002/fsn3.1775>
- [68] S.M. Pawar, S.R. Sonaware, “Fish muscle protein highest source of energy”. *International Journal of Biodiversity and Conservation*. Vol. 5, no. 7, pp. 433-435, 2013. <https://doi.org/10.5897/IJBC12.043>
- [69] F. Spallotta, , C. Cencioni, , S. Straino, *et al.*, “Enhancement of lysine acetylation accelerates wound repair”. *Communicative & Integrative Biology*, vol. 6, no. 5, 2013. <https://doi.org/10.4161/cib.25466>
- [70] M. Smriga, T. Ando, M. Akutsu, *et al.*, “Oral treatment with L-lysine and L-arginine reduces anxiety and basal cortisol levels in healthy humans”, *Biomedical Research*, vol. 28, no.2, pp. 85-90, 2007. <https://doi.org/10.2220/biomedres.28.85>
- [71] M.C. Pedrazini, M.H. da Silva, F.C. Groppo, “L-lysine: Its antagonism with L-arginine in controlling viral infection. Narrative literature review, *British Journal of Clinical Pharmacology*. vol. 88, no. 11, pp. 4708-4723, 2022. <https://doi.org/10.1111/bcp.15444>

- [72] D. Tomé, C. Bos, “Lysine requirement through the human life cycle”, *The Journal of Nutrition*, vol. 137, no.6, pp. 1642S–1645S, 2007. <https://doi.org/10.1093/jn/137.6.1642S>
- [73] M. Watford, “Glutamine and glutamate: Nonessential or essential amino acids?”, *Animal Nutrition*, vol. 1, no. 3, pp. 119-122, 2015. <https://doi.org/10.1016/j.aninu.2015.08.008>
- [74] X. Li, S. Zheng, G. Wu, “Nutrition and metabolism of glutamate and glutamine in fish”, *Amino Acids*. vol.52 , no.5, pp. 671-691, 2020. <https://doi.org/10.1007/s00726-020-02851-2>
- [75] M. Li, , Y. Wu, , L. Ye, “ The Role of Amino Acids in Endothelial Biology and Function”, *Cells*, vol.11, no.8, pp. 1372, 2022. <https://doi.org/10.3390/cells11081372>
- [76] M.M. Pal, “Glutamate: The Master Neurotransmitter and Its Implications in Chronic Stress and Mood Disorders”, *Frontiers in Human Neuroscience*, vol. 15, pp. 722323. <https://doi.org/10.3389/fnhum.2021.722323>
- [77] W. Wang, X. Zhou, Y. Liu, “Characterization and evaluation of umami taste: A review”, *TrAC Trends in Analytical Chemistry*, vol. 127, pp. 115876, 2020. <https://doi.org/10.1016/j.trac.2020.115876>
- [78] C. Marques, A. Reis, C. Moura, *et al.*, “Consumer insight into the monosodium glutamate”, *Acta Scientiarum. Technology*. vol. 40, 2018. <https://doi.org/10.4025/actascitechnol.v40i1.30838>
- [79] K. Stańska, , A. Krzeski, “The umami taste: from discovery to clinical use”. *Polish Journal of Otolaryngology*. vol. 70, no.4, pp. 10–15, 2016. <http://dx.doi.org/10.5604/00306657.1199991>

**6639837: Revision requested**

5 messages

International Journal of Food Science <ijfs@hindawi.com>

Sat, Sep 16, 2023 at 8:53 PM

Reply-To: International Journal of Food Science <deepanaa.rajadurai@hindawi.com>

To: Hafrijal Syandri <syandri_1960@bunghatta.ac.id>



Dear Dr. Syandri,

In order for your submission "Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security " to "International Journal of Food Science" to proceed to the review process, there needs to be a revision.

Reason & Details:

“

Dear authors,

We have received the reports from our advisers on your manuscript, "Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security". Based on the advice received, I have decided that your manuscript can be accepted for publication after a major revision, as suggested by the reviewer.

Attached, please find the reviewers' comments for your perusal.

For more information about what is required, please click the link below.

MANUSCRIPT DETAILS

Kind regards,

Deepanaa Rajadurai

on behalf of "International Journal of Food Science"

This email was sent to syandri_1960@bunghatta.ac.id. You have received this email in regards to the account creation, submission, or peer review process of a paper submitted to a journal published by Hindawi Limited.

Hindawi Limited, 3rd Floor, Adam House, [1 Fitzroy Square, London, W1T 5HF, United Kingdom](#).

Hindawi respects your right to privacy. Please see our [privacy policy](#) for information on how we store, process, and safeguard your data.

[Unsubscribe](#)

hafrizal syandri <syandri_1960@bunghatta.ac.id>
To: International Journal of Food Science <deepanaa.rajadurai@hindawi.com>

Mon, Sep 18, 2023 at 6:41 AM

Dear
Deepanaa Rajadurai
on behalf of "International Journal of Food Science"

Thank you for your email on September 17, 2023. We have read the reviewer's report for manuscript No. 6639837. Five points need to be revised. Therefore, we ask for seven days to make edits to this manuscript. We will request assistance from American Journal Experts (AJE) to make edits for language and grammatical errors in some sections.

Best regards
Hafrijal Syandri
[Quoted text hidden]

Phenom Emails <phenom.emails@hindawi.com>
Reply-To: Phenom Emails <phenom.emails@hindawi.com>
To: syandri_1960@bunghatta.ac.id

Mon, Sep 18, 2023 at 3:19 PM

Dear Dr. Syandri,

Hope you are good and well.

Thank you for your email and we are happy to grant your request for an extension during this uncertain time. Please note that you may receive some reminder emails from our system but please ignore these for the time being.

We hope to hear from you again soon.

Best Regards,

Deepanaa

Deepanaa Rajadurai
Editorial Assistant



Hindawi

[Hindawi.com](https://www.hindawi.com) | [Twitter](#) | [Facebook](#) | [LinkedIn](#) | [YouTube](#)

[Quoted text hidden]

, hafrizal syandri <syandri_1960@bunghatta.ac.id> wrote:

[Quoted text hidden]

[Quoted text hidden]

The contents of this email and any attachments are confidential and intended only for the person or entity to whom it is addressed. If you are not the intended recipient, any use, review, distribution, reproduction or any action taken in reliance upon this message is strictly prohibited. If you received this message in error, please immediately notify the sender and permanently delete all copies of the email and any attachments.

[Click here for translations of this disclaimer.](#)

hafrizal syandri <syandri_1960@bunghatta.ac.id>
To: Phenom Emails <phenom.emails@hindawi.com>

Thu, Sep 21, 2023 at 5:19 PM

Dear
Deepanaa Rajadurai
Editorial Assistant

We want to inform you that we have made revisions to manuscript No. 6639837, as follows:

1. The manuscript has been thoroughly edited, including correcting language and grammatical errors by American Journal Experts (AJE) with an attached certificate.
2. We have reduced the number of references according to the revised manuscript.
3. The manuscript has been narrowed down based on research information that we have investigated.

We appreciate your attention and look forward to hearing good news from you. Thank You.

Best regards

Hafrijal Syandri
[Quoted text hidden]

 **Sertificate AJE.pdf**
79K

Phenom Emails <phenom.emails@hindawi.com>
Reply-To: Phenom Emails <phenom.emails@hindawi.com>
To: syandri_1960@bunghatta.ac.id

Thu, Sep 21, 2023 at 6:27 PM

Dear Dr. Syandri,

Thank you for your email.

Please be informed that for your manuscript 6639837, your revised manuscript has been successfully submitted. Once the decision is made you will be notified.

Best Regards,

Deepanaa

Deepanaa Rajadurai
Editorial Assistant



Hindawi

[Hindawi.com](https://www.hindawi.com) | [Twitter](#) | [Facebook](#) | [LinkedIn](#) | [YouTube](#)

On Mon, 18 Sep at 9:19 AM , Phenom Emails <phenom.emails@hindawi.com> wrote:
Dear Dr. Syandri,

Hope you are good and well.

Thank you for your email and we are happy to grant your request for an extension during this uncertain time. Please note that you may receive some reminder emails from our system but please ignore these for the time being.

We hope to hear from you again soon.

Best Regards,

Deepanaa

Deepanaa Rajadurai
Editorial Assistant



Hindawi

Hindawi.com | [Twitter](#) | [Facebook](#) | [LinkedIn](#) | [YouTube](#)

On Mon, 18 Sep at 12:41 AM , hafrizal syandri <syandri_1960@bunghatta.ac.id> wrote:

Dear

Deepanaa Rajadurai

on behalf of "International Journal of Food Science"

Thank you for your email on September 17, 2023. We have read the reviewer's report for manuscript No. 6639837. Five points need to be revised. Therefore, we ask for seven days to make edits to this manuscript. We will request assistance from American Journal Experts (AJE) to make edits for language and grammatical errors in some sections.

Best regards
Hafrijal Syandri

[Quoted text hidden]

The contents of this email and any attachments are confidential and intended only for the person or entity to whom it is addressed. If you are not the intended recipient, any use, review, distribution, reproduction or any action taken in reliance upon this message is strictly prohibited. If you received this message in error, please immediately notify the sender and permanently delete all copies of the email and any attachments.

[Click here for translations of this disclaimer.](#)

Research Article

Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security

Azrita Azrita¹, Hafrijal Syandri^{1*}, Hazlina Ahamad Zakeri², Harfiandri Damanhuri¹, Netti Aryani³

¹*Faculty of Fisheries and Marine Science Universitas Bung Hatta, 25131, Padang-West Sumatra Province, Indonesia*

²*Faculty of Science and Marine Environment, Universiti Malaysia Terengganu, Malaysia.*

³*Faculty of Fisheries and Marine Science Universitas Riau, 28293, Pekanbaru-Riau Province, Indonesia*

*Corresponding author: syandri_1960@bunghatta.ac.id

Abstract:

Fish has become an irreplaceable source of animal protein food, especially among households with low socioeconomic conditions in rural and urban areas in Indonesia. This study analyzes the nutrient composition of three local Bagridae fish species in Indonesia. The nutrient composition results of *Hemibagrus nemurus*, *Hemibagrus wyckii*, and *Mystus nigriceps* revealed that the protein content was 24.26%, 22.57%, and 21.39% (% of dry weight), respectively. The total lipid content varied between 6.64% and 7.75%, whereas the crude ash content ranged from 1.59% to 2.30%. Regarding mineral contents, the calcium levels ranged from 1.49 to 1.66 mg/g, iron levels varied between 28.35 and 40.36 µg/g, and zinc levels ranged from 24.03 to 54.46 µg/g. The predominant amino acids found in the three species of Bagridae fish were glutamic acid, aspartic acid, alanine, arginine, and lysine, with their concentrations ranging from 9.10% to 24.34%. Among the fatty acids, C16:0 (palmitic acid) was the most abundant in all three species, accounting for 25.59% to 30.70% of the total fatty acids. Additionally, significant amounts of C18:1 (oleic acid), C18:0 (stearic acid), and C20:4 (arachidonic acid) were also detected as primary fatty acids. The calculated AI (atherogenic index) values in the three species of Bagridae fish ranged from 0.73 to 0.99, while the TI (thrombogenic index) values varied between 0.54 and 0.75. These results indicate that consuming the meat of these three freshwater species of Bagridae fish caught in the wild does not pose any health risks to consumers. They can be considered a safe and suitable food source with good nutritional quality.

Keywords: Nutrient composition, Amino acids, Fatty acids, Freshwater fish, Kampar Kanan River,

1. Introduction

Fish serve as a high-quality protein source that is essential for satisfying dietary protein requirements, especially in areas where other protein sources could be few or difficult to get [1,2]. Their accessibility can aid in ensuring a steady supply of food, particularly for communities whose main sources of income are fishing or aquaculture [3]. Fish can contribute to a more varied selection of protein sources in communities, resulting in a diet that is more nutritious and balanced. Dietary fish can treat nutrient deficits and enhance general nutritional health. Fish are generally considered safe and offer a lower risk of contamination with harmful substances compared to certain land-based animal products. According to Ravichandran et al. [4], a reliable indicator of the fish's quality, nutritional value, physiological status, and environment is said to be its chemical makeup, which is found in fish flesh.

Hemibagrus (H) nemurus, locally known as "baung", *Hemibagrus (H) wyckii*, known as "geso" and *Mystus (M) negriceps*, known as "ingir-ingir," are three species of catfish found in Kampar Kanan River, Indonesia [5,6]. Due to their high market value, these species have been identified as significant economic resources for the rural community [7]. Therefore, the nutritional quality of fish meat in the study area is very important for analysis that is beneficial to human health.

Knowing the composition of fatty acids and amino acids in fish meat is an essential factor that should not be ignored [8]. This information provides valuable insight into the fish's nutritional quality and values of more nutritious and healthy foods [9]. In addition, a better understanding of the composition of fatty acids and amino acids can assist in selecting a suitable feed [10], developing a balanced diet [11], and maintaining optimal fish meat nutritional [12].

Omega-3 polyunsaturated fatty acids, particularly DHA and EPA derived from fish oil, exhibit anti-inflammatory and anti-gastrointestinal cancer properties, making them crucial as immune-boosting nutrients [13,14]. Studies have demonstrated that ω -3 and ω -6 polyunsaturated fatty acids (PUFA) exhibit beneficial effects in the management of cardiovascular diseases and cancers [15,16]. The fatty acid composition of polyunsaturated fatty acids (PUFAs) can differ among different types of fish, including both freshwater and marine species [17, 18]. Amino acids (AAs) like cysteine, arginine, tyrosine, glycine, proline, and serine play a vital role in

illness and stress situations and the prevention of inflammation and the repair processes in fish intestines [19, 20].

Furthermore, knowledge of the atherogenicity index (AI) and thrombogenicity (TI) of fatty acids in three local freshwater fish species of Bagridae can contribute to improving dietary guidelines. AI and TI indexes can be used to develop recommendations aimed at improving heart health [21,22]. These recommendations may involve selecting fish with favorable fatty acid profiles, allowing individuals to choose the most suitable fish species based on their specific needs [17, 23]. Therefore, this study aims to examine the nutritional composition profile, mineral content, amino acids, and fatty acids, including AI and TI indices of three freshwater fish species, namely *H. nemurus*, *H. wyckii*, and *M. nigriceps*. These three types of fish are generally consumed by people who live around the Kampar Kanan River in Kampar Regency, Riau Province, Indonesia.

2. Material and methods

2.1. Materials

In this study, a total of 27 Bagridae fish belonging to three distinct species (*H. nemurus*, *H. wyckii*, and *M. nigriceps*) were examined. The specimens were obtained from local fishermen operating in the upper stretches of the Kampar Kanan River in Kampar Regency, Riau Province (Figure 1). The sampling took place between June and August 2022. They were verified by the Fisheries Department, Faculty of Fisheries and Marine Science Universitas Bung Hatta, Indonesia. The samples were placed in plastic bags and transported to the laboratory in an insulated icebox. The samples were then grouped based on their respective sampling locations.

2.2. Biometry measurement

Upon reaching the laboratory within 10 hours, the fish samples were subjected to gutted and washing. Each specimen was weighed (TW) and measured to determine the standard length (SL) as well as the depth or maximum height (H). TW was measured using 600i-scales with a precision of 0.001 gram. The length measurement was taken from the mouth's tip to the end of the upper lobe of the caudal fin, representing the total body length using a meter ruler with an accuracy of 1 millimeter. The height measurement involved a vertical assessment of the body's maximum height measured using a Digital Sketmat Sigmat Vernier Caliper with an accuracy of 1 millimeter. The condition factor (CF) was calculated using the formula $CF = (TW/SL^3) \times 100$.

2.3. Proximate, amino acid, and mineral content analysis

The proximate carcass composition was assessed following the AOAC standard methods [24]. The samples were dried until a constant weight was achieved at 105°C. The crude protein content was analyzed using the standard Kjeldahl method, which involves multiplying the nitrogen content by a factor of 6.25. The Soxhlet method with ether extraction was employed to analyze crude lipids. The ash content was obtained by incinerating the samples at 550°C for 16 hours. The amino acid composition was determined using a high-performance liquid chromatography (HPLC) system, which consisted of a water 1525 binary HPLC pump, 717 autosamplers (water®), and water 2475 multi λ fluorescence detector optics (with excitation at 250 nm and emission at 395 nm). The samples were hydrolyzed in triplicate with 6 N hydrochloric acid for 24 hours at 11°C [25].

For mineral data composition (Na, Mg, Ca, K, P, Fe, Cu, Mn, and Zn), the ashed of feed sample and carcass were dissolved in 1 ml of hydrochloric acid (35% v/v Suprapur® Merck). Furthermore, it was filtered with cellulose filter paper (Watchman No 1, International Ltd; Maidstone, UK) and diluted to an appropriate concentration for each elemental mineral. P levels were analyzed with a Perkin-Elmer AA spectrophotometer mod 3110 (Norwalk, CT. USA).

2.4. Fatty acids analysis

The body meat of fish was examined utilizing the gas chromatography-mass spectrometry (GC-MS) method. The modified Folch *et al.* (1957) method, as described by Rajion [26], was employed to perform the total fat extraction. This involved utilizing a solvent system consisting of chloroform and methanol in a 2:1 (v/v) ratio. The process of transmethylation was conducted using a solution of 14% methanolic boron trifluoride. The fatty acid composition of the meat was then analyzed at the Laboratory Accredited Testing Laboratory-LP-184-IDN.

The nutritional quality of lipids AI and TI was calculated using the equations [27].

$$AI = \frac{[(C12:0 + 4 \times C14:0 + C16:0)]}{\sum MUFA + \sum n - 6 + \sum n - 3}$$

$$TI = \frac{[(C14:0 + C16:0 + C18:0)]}{[(0.5 \times \sum MUFA) + (0.5 \times \sum n - 6) + (3 \times \sum n - 3) + (\sum n - 3 / \sum n - 6)]}$$

Where

AI = Atherogenic index

TI = Thrombogenic index

C12:0 = lauric acid

C14:0 = myristic acid

C16:0 = palmitic acid

C18:0 = stearic acid

\sum MUFA = sum concentrations of all monosaturated fatty acid

\sum n-6 = sum concentrations of n-6 polyunsaturated fatty acid

\sum n-3 = sum concentrations of n-3 polyunsaturated fatty acid

2.5. Data analysis

Data analysis was performed using the SPSS 16.0 software package (SPSS; Chicago, IL). The homogeneity of the data was assessed using the Levine test. One-way ANOVA was carried out to determine the parameters of proximate and amino acid composition, including the composition of fatty acids and nutritional quality of lipids AI and TI for each species, followed by Duncan's multiple range test post hoc [28]. The results are reported as mean values \pm standard errors for each parameter.

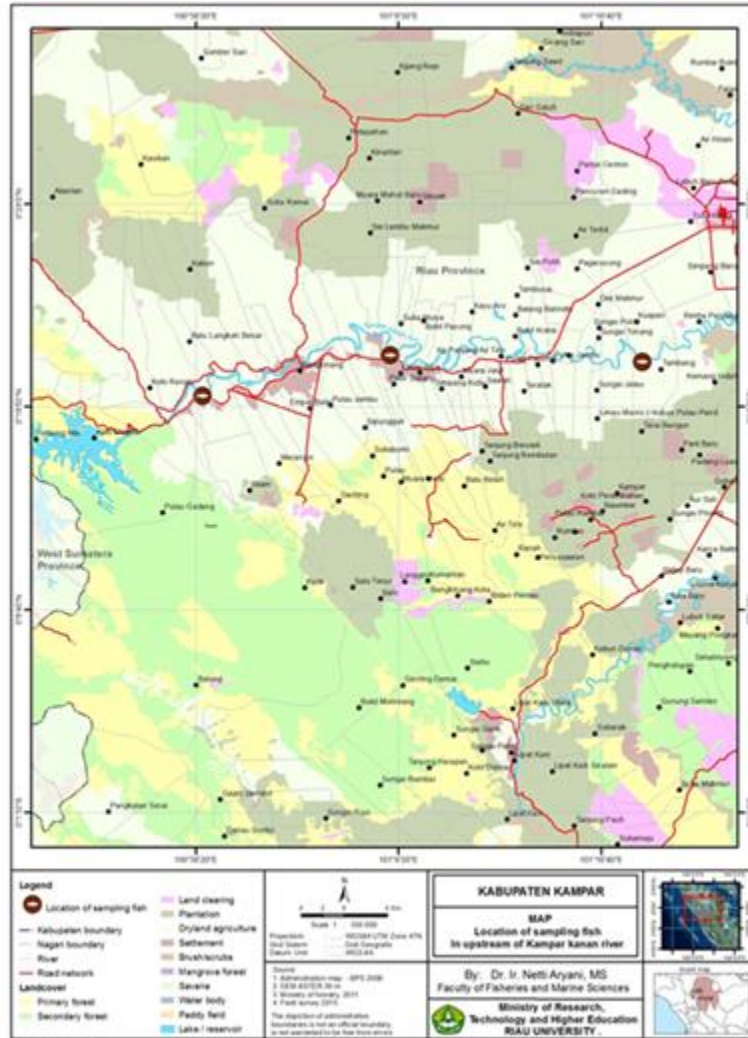


Figure 1. Map of Kampar Regency, Riau Province, and locations of three species of Bagridae fish sampling.

3. Results

Table 1 presents the average wet weight, standard length, width, and the results of meat nutritional composition analysis for three indigenous species of Indonesian freshwater Bagridae. The moisture content (% wet weight) ranges from 82.40% to 85.39%. Among the species examined, *M. nigriceps* exhibits the lowest protein content at 21.39%, while *H. nemurus* displays the highest value at 24.26%. However, *M. nigriceps* recorded has a higher mineral content such as iron and zinc.

Table 2 summarizes the fatty acid composition (% of total fatty acid) for the three species of Bagridae fish. C16:0 is the predominant fatty acid in the three species of Bagridae fish, with percentages ranging from 25.09% to 30.71%. The additional, prominent fatty acids found are C18:1, C18:0, and C22:6. *H. nemurus* exhibits a ω -6: ω -3 ratio below 1 (0.82), while *H. wyckii* and *M. nigriceps* have ratios of 1.11 and 1.02, respectively. The obtained PUFA/saturated (P/S) ratio varied between 0.61 and 0.76. In contrast, the AI values were 0.76 and 0.99, and the TI values were 0.54 and 0.75 for the respective cases (**Table 3**).

Table 4 presents the amino acid composition (% of total protein). Glutamic acid is the dominant amino acid, ranging from 19.29% to 24.34%, followed by aspartic acid, which ranges from 9.21% to 11.27%. Lysine content is consistent, ranging from 9.67% to 9.87%. In *H. nemurus*, the levels of various amino acids range from 0.86% to 24.35%. Similarly, in *H. wyckii*, the levels range from 0.76% to 21.58%, while in *M. nigriceps*, the levels range from 0.74% to 19.29%.

Table 1. Results of proximate analysis of three species of Bagridae fish

	Species		
	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
<i>Biometric measurements</i>			
Total weight (g)	389.99 ± 24.96 ^a	1,390.33 ± 168.29 ^b	17.57 ± 1.53 ^c
Standard length (cm)	28.16 ± 0.53 ^a	44.14 ± 1.98 ^b	10.29 ± 0.15 ^c
Height (cm)	8.44 ± 0.16 ^a	8.82 ± 0.39 ^b	2.57 ± 0.03 ^c
Condition factor	1.63 ± 0.08	1.58 ± 0.07	1.56 ± 0.08
<i>Proximate composition</i>			
Crude protein (% DW)	24.26 ± 0.87 ^a	22.57 ± 0.37 ^b	21.39 ± 0.15 ^c
Crude fat (% DW)	6.64 ± 0.03 ^a	7.47 ± 0.02 ^b	7.75 ± 0.40 ^c
Crude ash (% DW)	1.94 ± 0.02 ^a	2.30 ± 0.09 ^b	1.59 ± 0.02 ^c
Moisture (% WW)	82.40 ± 2.51	85.39 ± 2.36	83.75 ± 2.21
<i>Mineral composition</i>			
Macromineral (mg/g)			
Sodium	0.99 ± 0.00 ^a	1.61 ± 0.01 ^b	1.61 ± 0.02 ^c
Magnesium	1.12 ± 0.01 ^a	0.57 ± 0.02 ^b	1.15 ± 0.01 ^c
Calcium	1.66 ± 0.00 ^a	1.55 ± 0.02 ^b	1.49 ± 0.04 ^c
Potassium	0.71 ± 0.00 ^a	0.55 ± 0.02 ^b	0.43 ± 0.01 ^c
Phosphorous	7.03 ± 0.03 ^a	2.74 ± 0.02 ^b	6.45 ± 0.07 ^c
Microminerals (µg/g)			
Iron	28.30 ± 0.11 ^a	28.73 ± 0.08 ^b	40.36 ± 0.55 ^c
Copper	8.93 ± 0.03 ^a	7.46 ± 0.09 ^b	6.21 ± 0.32 ^c
Manganese	1.64 ± 0.02 ^a	2.64 ± 0.02 ^b	2.84 ± 0.02 ^c

Zinc	24.03 ± 0.45 ^a	24.61 ± 0.19 ^b	54.46 ± 0.17 ^c
------	---------------------------	---------------------------	---------------------------

Values are % ± SE; *Values in the same row with a different superscript a significantly different (p<0.05)

Table 2. Fatty acid composition of three species of Bagridae fish

Fatty acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
C12:0 (lauric acid)	1.97 ± 0.05 ^a	3.94 ± 0.01 ^b	4.27 ± 0.01 ^c
C14:0 (Meristic acid)	2.38 ± 0.03 ^a	2.72 ± 0.08 ^b	2.89 ± 0.01 ^c
C16:0 (Palmitic acid)	27.23 ± 0.06 ^a	30.70 ± 0.06 ^b	25.59 ± 0.01 ^c
C18:0 (Stearic acid)	16.17 ± 0.02 ^a	13.54 ± 0.02 ^b	10.64 ± 0.01 ^c
C20:0 (Arachidic acid)	0.23 ± 0.01 ^a	0.14 ± 0.01 ^b	0.37 ± 0.01 ^c
C16:1 (Palmitoleic acid)	1.78 ± 0.00 ^a	1.27 ± 0.04 ^b	4.93 ± 0.01 ^c
C18:1 (Oleic acid)	16.86 ± 0.01 ^a	16.18 ± 0.01 ^b	16.97 ± 0.01 ^c
C18:2 (Linoleic acid)	3.24 ± 0.02 ^a	3.84 ± 0.33 ^b	4.55 ± 0.001 ^c
C18:3 (Linolenic acid)	1.12 ± 0.01 ^a	0.82 ± 0.01 ^b	1.57 ± 0.00 ^c
C20:4 (Arachidonic acid)	12.13 ± 0.01 ^a	12.49 ± 0.09 ^b	12.41 ± 0.02 ^c
C20:5 (Eicosapentaenoic acid; EPA)	2.06 ± 0.08 ^a	2.34 ± 0.04 ^b	1.35 ± 0.01 ^c
C22:6 (Docosahexaenoic acid; DHA)	15.65 ± 0.01 ^a	11.56 ± 0.04 ^b	13.64 ± 0.02 ^c

Values are % ± SE; *Values in the same row with a different superscript a significantly different (p<0.05)

Table 3. Fatty acid ω-6/ω-3 and polyunsaturated/saturated fatty acid ratio of three species of Bagridae fish

	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
ω-6/ω-3	0.82 ± 0.00 ^a	1.11 ± 0.02 ^b	1.02 ± 0.01 ^c
Ratio of P/S	0.69 ± 0.01 ^a	0.61 ± 0.06 ^b	0.76 ± 0.00 ^c
AI	0.87 ± 0.01 ^a	0.99 ± 0.06 ^b	0.73 ± 0.00 ^c
TI	0.63 ± 0.01 ^a	0.75 ± 0.03 ^b	0.54 ± 0.00 ^c

P/S, polyunsaturated/saturated fatty acid; AI, Atherogenic index; TI, Thrombogenic index

Values are % ± SE; *Values in the same row with a different superscript a significantly different (p<0.05)

Table 4. Amino acid composition of three species of Bagridae fish

Amino acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
Aspartic acid	10.39 ± 0.01 ^a	9.61 ± 0.04 ^b	11.28 ± 0.02 ^c
Glutamic acid	24.34 ± 0.00 ^a	21.58 ± 0.41 ^b	19.29 ± 0.03 ^c
Serine	3.74 ± 0.00 ^a	5.40 ± 0.03 ^b	7.50 ± 0.06 ^c
Glycine	4.02 ± 0.00 ^a	5.77 ± 0.04 ^b	4.69 ± 0.02 ^c
Histidine	2.39 ± 0.00 ^a	2.90 ± 0.03 ^b	2.21 ± 0.02 ^c
Arginine	6.17 ± 0.01 ^a	8.27 ± 0.03 ^b	6.22 ± 0.03 ^c
Threonine	4.49 ± 0.01 ^a	4.19 ± 0.03 ^b	4.39 ± 0.04 ^c
Alanine	7.03 ± 0.00 ^a	6.04 ± 0.02 ^b	7.89 ± 0.01 ^c
Proline	2.40 ± 0.00 ^a	2.71 ± 0.01 ^b	2.21 ± 0.02 ^c
Tyrosine	3.91 ± 0.01 ^a	3.29 ± 0.06 ^b	3.82 ± 0.04 ^c
Valine	3.73 ± 0.00 ^a	4.02 ± 0.01 ^b	3.85 ± 0.04 ^c
Methionine	3.42 ± 0.00 ^a	1.96 ± 0.02 ^b	1.27 ± 0.03 ^c
Cystine	0.87 ± 0.00 ^a	0.76 ± 0.02 ^b	0.75 ± 0.03 ^c
Isoleucine	3.82 ± 0.001 ^a	3.96 ± 0.003 ^b	3.66 ± 0.002 ^c
Leucine	6.15 ± 0.000 ^a	6.69 ± 0.002 ^b	6.28 ± 0.004 ^c
Phenylalanine	5.13 ± 0.000 ^a	6.03 ± 0.005 ^b	5.18 ± 0.027 ^c
Lysine	9.61 ± 0.001 ^a	9.82 ± 0.026 ^b	9.87 ± 0.003 ^c

Values are % ± SE; *Values in the same row with a different superscript a significantly different (p<0.05)

4. Discussion

The results of this study provided significant new information about the nutritional makeup of the studied species of Bagridae fish. The fish's biometric measures yielded important information about its physical traits and health. Researchers are able to determine the average weight of fish at various lengths by examining the link between length and weight for a particular species. This knowledge is essential for understanding the trends in fish population growth, spotting potential shifts in growth rates over time, and assessing the general health and welfare of the fish [29]. A variety of factors, including sex, age, maturity level, size, level of

stomach fullness, sampling strategies, sample sizes, and environmental circumstances, have an impact on fish health and the parameters that determine length-weight connections in fish [30]. However, none of these factors are considered in our study. In general, according to the theory put forth by Bagenal and Tesch [31], heavier fish of a given length are in better physiological condition. The condition factor is also used in fisheries science to compare the health, fatness, and condition of fish [32]. A condition factor of > 1 is favorable, indicating a good level of feed nutrition and suitable habitat for living [33]. Thus, from the findings in this study, it may be said that the Kampar Kanan River provides a suitable habitat for the fish, ensuring their accessibility and availability for community consumption.

The fish samples exhibited good nutritional profiles, with high amounts of proteins and lipids as well as moisture content contributing to their overall nutritional value, according to the results of proximate analysis and minerals composition (see Table 1). These results imply that the fish species studied may provide a possible source of nutrients to solve issues with food security. The study of the mineral's composition revealed the existence of critical minerals such as phosphorus (P), calcium (Ca), magnesium (Mg), iron (Fe), and zinc (Zn) in the fish samples in addition to proteins and lipids. All three Bagridae fish significantly differ from each other in terms of their crude ash content and minerals profile (see Table 1). However, in general, P and Ca are the two highest minerals found in all fish. Both these macro minerals are good for human bone and teeth health, as key components of hydroxyapatites, the mineral matrix that provides strength and structure to bones and teeth [34].

In the current study, the meat of three species of Bagridae fish from the Kampar Kanan River in Indonesia was found to contain an abundance of microminerals such as Fe and Zn. *M. nigriceps* which has been recorded to exhibit higher mineral content, including Fe and Zn. This species is categorized as a small indigenous fish (SIF) with a maximum standard length of approximately 10.29 cm. According to [35], SIFs, in general, are known to be rich in Ca, Fe, and Mn.

~~In the current study, the meat of three species of Bagridae fish from the Kampar Kanan River Indonesia was found to contain abundance of microminerals such as Fe and Zn. Insufficient intake of Fe and Zn has been linked to major developmental health problems, such as~~

Fe deficiency which is one of the major causes of anemia in pregnant and lactating women, especially in rural communities [41,42]. Lacking in Zn, on the other hand, can lead to numerous immunological disorders, such as metabolic and chronic illnesses, as well as infectious diseases including respiratory infections, malaria, HIV, or tuberculosis [43]. *M. nigriceps* that has been recorded exhibits higher mineral content, including iron and zinc. This species is categorized as a small indigenous fish (SIF) with a maximum standard length of approximately 10.29 cm. According to Mohanty et al. [44], SIFs, in general, are known to be rich in Ca, Fe, and manganese (Mn). The variation in mineral content among the three species of Bagridae fish is comparable to that of *Clarias gariepinus* and *Tilapia zilli* meat from the Owan River in Edo State, Nigeria [13].

In general, fish consists of 70% to 84% moisture [45]. High moisture content is often associated with freshness in fish. Compared to fish that has been processed or stored, fresh fish often has a higher moisture content, the factor that contributes to a moist and succulent texture. Fish with high moisture content may retain more water-soluble nutrients during cooking or processing as proven by Gupta et al. [46] with green leafy vegetables. This can help preserve the nutritional value of the fish, ensuring that important vitamins and minerals are retained. However, due to the high moisture content in fish, a processing technique that changes the moisture content will also change the fish's properties, such as the texture and protein gelation [47].

The amount of crude fats in the fish is a good indicator of how much lipids or fats are present. In addition to acting as transporters for fat-soluble vitamins, fats offer a concentrated source of energy to the body. Additionally, they improve the fish's flavour, texture, and palatability [48]. Numerous health advantages, including improved cardiovascular health, improved brain function, and anti-inflammatory effects, have been linked to specific fatty acids present in fish, particularly omega fatty acids. So a fish with a high amount of crude fat might be good, especially if it has good fatty acids. From the findings, all three Bagridae fish had a total fat content of around 7-8% by weight (Table 1). According to Fuentes et al. [49], fish with lower fat content exhibit higher water content, which is also shown in our study. On the other hand, Nurnadia et al. [50] reported that the fish species can be grouped into different categories,

including high-fat fish (>8%), medium-fat fish (4-8%), low-fat fish (2-4%), and lean fish (<2%). According to the established classification, the three indigenous Bagridae fish species are characterized as having a moderate level of fat content, which is reflected by the high levels of DHA and EPA in the meat of *H. nemurus*, *H. wyckii*, and *M. nigriceps* fish (Table 2). It is still unclear whether consuming the fish meat of these three species can indirectly help lower blood pressure, reduce inflammation in blood vessels, improve lipid profiles by increasing levels of good cholesterol (HDL) and decreasing levels of bad cholesterol (LDL), and reduce blood clotting in individuals. Nonetheless, the fat content of fish is determined by various factors, such as species, habitat ecosystems, feeding patterns, and other biological attributes, including the distinctive flavor characteristics of the fish [8,49,51,52].

In this study, all three Bagridae fish had a total fat content of around 7% to 8% by weight, with a higher water content ranging from 82.40% to 85.39% (Table 1). Similarly, for wild and cultured Sea bass, *Dicentrarchus labrax* [36]. 49]. Additionally, the fish species can be grouped into different categories, including high-fat fish (>8%), medium-fat fish (4-8%), low-fat fish (2-4%), and lean fish (<2%) [37].- 49]. According to the established classification by [37] 49], the three indigenous Bagridae fish species are characterized as having a moderate level of fat content, which is reflected by the high levels of DHA and EPA in the meat of *H. nemurus*, *H. wyckii*, and *M. nigriceps* (Table 2). Therefore, we have recommended consuming the fish meat of these three species can indirectly help for food security in rural and urban communities.

Each of the three Bagridae fish species analyzed in the study contained arachidonic acid (C20:4) 12.13% for *H.nemerus*, 12.49 % for *H. wyckii*, and 12.41% for *M. nigriceps*. Arachidonic acid serves as a precursor for synthesizing prostaglandins and leukotrienes [38, 53]. Leukotrienes are crucial in allergic responses, inducing smooth muscle contraction and increasing vascular permeability [39, 54]. Additionally, all three species also contained EPA ranging from 1.35% and 2.34%, and DHA ranging from 11.56% to 15.65%. The increased consumption of omega-3 fatty acids, such as EPA and DHA, has been linked to a decreased risk of pain development in older individuals and preventive Alzheimer's disease [40,41, 55,56].

~~Each of the three Bagridae fish species analyzed in the study contained arachidonic acid (C20:4), which serves as a precursor for synthesizing prostaglandins and leukotrienes [53]. Leukotrienes are crucial in allergic responses, inducing smooth muscle contraction and~~

increasing vascular permeability [54]. In this study, the arachidonic acid content in *H. nemurus*, *H. wyckii*, and *M. negriceps* was determined to be 12.13%, 12.49%, and 12.41%, respectively. Conversely, the levels of arachidonic acid in *Channa striatus*, *Channa micropeltes*, and *Channa lucius* were reported as 19.02%, 4.71%, and 12.41%, respectively [55]. According to Carballo-Casla et al. [56], increased consumption of marine omega-3 fatty acids, such as EPA and DHA, has been linked to a decreased risk of pain development in older individuals. Additionally, DHA and EPA have demonstrated a preventive effect on human coronary arteries and Alzheimer's disease [57,58].

As determined by the current study, the ω -6: ω -3 ratios of three Bagridae fish species ranged from 1:0.8 to 1:0.97. It was found that all three fish species had ω -6: ω -3 ratios within the recommended range for a typical Indonesian diet. In contrast, a typical Western diet is characterized by a high ω -6: ω -3 ratio of 10:1 to 20:1, while a Mediterranean diet has a ratio of 4:1 to 5:1 [42, 61]. On the other hand, P/S ratio analysis shows that the three Bagridae fish species are contained sources of PUFA that meet the requirements for food safety.

In this study, our hypothesis is that the levels of EPA and DHA found in the meat of three species of Bagridae fish in the Kampar Kanan River, Indonesia, may also have the potential in reducing muscle pain and inflammation. Additionally, as a primary food source, fish plays a role in the prevention and treatment of chronic pain [59]. As determined by the current study, the ω -6: ω -3 ratios of three Bagridae fish species ranged from 1:0.8 to 1:0.97. These varying ω -6: ω -3 ratios can have diverse implications for treating or preventing diseases [60]. It was found that all three fish species had ω -6: ω -3 ratios within the recommended range for a typical Indonesian diet. In contrast, a typical Western diet is characterized by a high ω -6: ω -3 ratio of 10:1 to 20:1, while a Mediterranean diet has a ratio of 4:1 to 5:1 [61]. The P/S ratio analysis revealed that all three Bagridae fish species were considered an average source of PUFA. Additionally, it has been reported that marine fish serve as a good source of PUFA [62].

The atherogenic index (AI) and thrombogenic index (TI) indices are directly influenced by the levels of C14:0, C16:0, and C18:0, all of which are known to promote thrombogenicity [24]. AI and TI indices indicate the potential to induce platelet aggregation. A lower value in AI and TI indicates a more significant protective potential against coronary artery disease [63]. The Food and Agriculture Organization (FAO) and the World Health Organization (WHO) have

~~recommended AI and TI values ranging from 0.4 to 0.5 for promoting human health [64]. The AI and TI values of various marine fish species, such as Sardine (*Sardinella hualiensis*), being 0.60 and 0.20, respectively, and Mackerel (*Scomber japonicus*) having values of 0.48 and 0.24, respectively [65]. In the current study, the observed AI values in three freshwater species of Bagridae fish ranged from 0.73 to 0.99, while the TI values varied between 0.54 and 0.75. This finding is associated with a notable disparity in the values of saturated fatty acids (SFA) among the three species of Bagridae fish. Based on these findings, we certify that consuming the meat of three freshwater species of Bagridae fish caught in the wild does not endanger consumers' health and can serve as a suitable source of food safety and nutritional quality.~~

The Food and Agriculture Organization (FAO) and the World Health Organization (WHO) have recommended AI and TI values ranging from 0.4 to 0.5 for promoting human health [43, 64]. In the current study, the observed AI values in three freshwater species of Bagridae fish ranged from 0.73 to 0.99, while the TI values varied between 0.54 and 0.75. This finding is associated with a notable disparity in the values of saturated fatty acids (SFA) among the three species of Bagridae fish. Based on these findings, we certify that consuming the meat of three freshwater species of Bagridae fish caught in the wild does not endanger consumers' health and can serve as a suitable source of food safety and nutritional quality.

Fish with a high crude protein content, as also shown by the three Bagridae fish studied (Table 1), are regarded as being nutritionally useful for human health. According to Pawar and Sonaware [44, 68], protein and amino acid content can directly indicate the nutritional quality of meat. In Table 4, *H. wykii* had higher content of essential amino acids (EAA) than *H. nemurus* and *M. nigriceps*. Among the EAA, lysine (Lys) was found to be present in the highest amount in all three species. Lys is necessary for tissue growth and repair, reduced stress-induced anxiety, and the conversion of fatty acids into energy [45-47, 69,71,72].

~~A high value for crude protein shows that the fish contains a considerable amount of protein. Since fish protein is easier to digest than protein found in meat and includes less collagen fibre, the body can use its protein more effectively [66]. The body's tissues need protein to develop, mend, and remain healthy. The absence of amino acids and fatty acids elements will impede the recovery process [67]. Protein is a crucial ingredient for human health, contributing to a number~~

of physiological functions like immunological function, muscular growth, and enzyme synthesis. Since protein is a crucial part of a balanced diet, fish with a high crude protein content, as also been shown by the three Bagridae fish studied (Table 1), are regarded as being nutritionally useful. According to Pawar and Sonaware [68], protein and amino acid content can directly indicate the nutritional quality of meat. In Table 4, *H. wykii* had higher quantities of essential amino acids (EAA) than *H. nemurus* and *M. nigriceps*. Since EAA cannot be synthesised by the body yet are necessary for physiologic function, they are significant for humans. They are thus derived through EAA rich foods such as fish. Among the EAA, lysine (Lys) was found to be present in the highest amount in all three species. Lys is necessary for tissue growth and repair [69]. Furthermore, Lys can reduced stress induced anxiety [70] and has antiviral properties [71]. Lys also plays a role in metabolic activities, including the creation of carnitine for mitochondrial energy generation and the conversion of fatty acids into energy [72].

More than 60% of the amino acid composition in all three Brigade fish studied comes from the non-essential amino acids (NEAA), particularly glutamic acid (Table 4). Glutamic and its amine, glutamine are highly abundant amino acids found in fish in the free and protein-bound forms [48,74]. Other amino acids found abundant in the three fish also play important physiologic as well as biochemical functions. The NEAA, arginine, for example, is involved in protein synthesis, creatine formation, and various other biochemical processes within the body. The EAAs, leucine, phenylalanine, and methionine, play important roles in the synthesis of nitric oxide, a molecule that can expand blood vessels [49, 75].

4. Conclusions

The present study offers insightful information about the fatty acid and amino acid compositions, biometric measurements, proximate analysis, and mineral composition of three local freshwater species of Indonesian Bagridae fish. The findings show their potential as a nutrient-dense, sustainable food source that can support efforts to increase food security. These fish species might be incorporated into the local diet to increase nutrient intake and alleviate nutritional inadequacies, which would ultimately improve Indonesia's overall food security status.

Data Availability

The data utilized in this research has not been previously released or published in any form. The data sets employed and/or analyzed during the present study can be obtained by contacting the corresponding author.

Conflicts of Interest

The authors confirm that they do not have any conflicting interests.

Acknowledgements

The author acknowledges and expresses gratitude for the financial support received from the Ministry of Education, Culture, Research, and Technology of the Republic of Indonesia, specifically through grant no. 076/E5/PG. 02.00. PT/2022, which funded the research. The author affirms that there is no conflict of interest associated with the utilization of these research funds. Furthermore, the author extends appreciation to the students and fishermen involved in the research conducted in the upper reaches of the Kampar Kanan River, Indonesia.

References

- [1] A.G. Hasselberg, I.Aakre, J. Scholtens *et al.*, “Fish for food and nutrition security in Ghana: Challenges and opportunities,” *Global Food Security*. vol 26: 100380, 2020 <https://doi.org/10.1016/j.gfs.2020.100380>
- [2] N.Tran, U.P, Rodriguez, C.Y.Chan., *et al.*, “Future scenarios of fish supply and demand for food and nutrition security in Bangladesh: An analysis with the AsiaFish model”, *Aquaculture*. Vol.568: 739288, 2023. <https://doi.org/10.1016/j.aquaculture.2023.739288>
- [3] C.Z. Levkoe, K. Lowitt, C. Nelson, (2017). “Fish as food: Exploring a food sovereignty approach to small-scale fisheries”. *Marine Policy*. vol 85: pp.65-70, 2023. <https://doi.org/10.1016/j.marpol.2017.08.018>
- [4] M. Ravichandran, N.S. Hettiarachchy, V. Ganesh *et al.*, “Enhancement Of Antimicrobial Activities Of Naturally Occurring Phenolic Compounds By Nanoscale Delivery Against Listeria Monocytogenes, Escherichia Coli O157:H7 And Salmonella Typhimurium In Broth And Chicken Meat System”. *Journal of Food Safety*. vol 31, no. 4, pp. 462-471,2011. <https://doi.org/10.1111/j.1745-4565.2011.00322.x>
- [5] N. Aryani, I. Suharman, Azrita, *et al.*, “Diversity and distribution of fish fauna of upstream and downstream areas at Koto Panjang Reservoir, Riau Province, Indonesia”. *F1000 Research*. vol 8, pp. 1435. 2020. <https://f1000research.com/articles/8-1435/v2>
- [6] Syafrialdi, Dahelmi., D.I. Roesma, *et al.*, “Length-weight relationships and condition factor of Two-Spot Catfish (*Mystus nigriceps* [Valenciennes, 1840]) Pisces, Bagridae], from Kampar Kanan River and Kampar Kiri River in Indonesia”. *Pakistan Journal of Biological Science*, vol. 23, no. 12, pp. 1636-1642, 2020. <https://scialert.net/fulltext/?doi=pjbs.2020.1636.1642>

- [7] N. Aryani, "Native species in Kampar Kanan River, Riau Province Indonesia. *International Journal of Fisheries and Aquatic Studies*", vol.2, no.5, pp. 213-217. 2015. <https://www.fisheriesjournal.com/archives/2015/vol2issue5/PartD/2-5-63.pdf>
- [8] Azrita, H. Syandri, N. Aryani, *et al.*, "The utilization of new products formulated from water coconut, palm sap sugar, and fungus to increase nutritional feed quality, feed efficiency, growth, and carcass of gurami sago (*Osphronemus goramy* Lacepède, 1801) juvenile". *F1000 Research*. vol. 10. :1121, pp. 1-16, 2021. <https://f1000research.com/articles/10-1121/v1>
- [9] Azrita, H. Syandri, N. Aryani *et al.*, "Effect of feed enriched by products formulated from coconut water, palm sap sugar, and mushroom on the chemical composition of feed and carcass, growth performance, body indices, and gut micromorphology of giant gourami, *Osphronemus goramy* (Lacepède, 1801), juveniles". *F1000 Research*. vol 12. No.140, pp. 1-19, 2023. <https://f1000research.com/articles/12-140/v1>
- [10] L.F. Montenegro, A.M. Descalzo, R. Rizzo, *et al.*, "Improving the antioxidant status, fat-soluble vitamins, fatty acid composition, and lipid stability in the meat of Grass carp (*Ctenopharyngodon idella* Val) fed fresh ryegrass (*Lolium multiflorum* Lam)". *Aquaculture*, 553, 738067, 2022. <https://doi.org/10.1016/j.aquaculture.2022.738067>
- [11] C. Bene, R. Arthur, H. Norbury *et al.*, "Contribution of Fisheries and Aquaculture to Food Security and Poverty Reduction: Assessing the Current Evidence". *World Development*, 79: 177–196. 2016. <http://dx.doi.org/10.1016/j.worlddev.2015.11.007>
- [12] N. Aryani, I. Suharman, S. Hasibuan, *et al.*, "Fatty acid composition on diet and carcasses, growth, body indices and profile serum of Asian redbtail catfish (*Hemibagrus nemurus*) fed a diet containing different levels of EPA and DHA". *F1000Research*. 11:1409.pp.1-23, 2023. <https://doi.org/10.12688/f1000research.126487.2>
- [13] E.N. Kingsley, O.O. Cyril, O.I. Patience, "Potential contribution of selected wild fish species to the minerals intake of Pregnant and Lactating Women, Children and Adults in Rural Riverine Communities of Edo State: Insights and Outcomes". *Measurement: Food*, 8, 100063 : 1-7, 2022. <https://doi.org/10.1016/j.meafao.2022.100063>
- [14] A.A. Sausa, L. Nora, D.L.A. Lopes, *et al.*, "Vegetable choline in feed for Nile tilapia (*Oreochromis niloticus*) raised in a biofloc technology system (BFT): Biofloc composition, chemical composition, and fatty acid profiles in meat". *Aquaculture*. vol. 545. 737174, 2021. <https://doi.org/10.1016/j.aquaculture.2021.737174>
- [15] A.M. Eltweri, A.L. Thomas, M. Metcalfe, *et al.*, 2017. Potential applications of fish oils rich in omega-3 polyunsaturated fatty acids in the management of gastrointestinal cancer. *Clinical Nutrition*. vol. 36, no. 1, pp. 65-78. 2017. <https://doi.org/10.1016/j.clnu.2016.01.007>
- [16] P.C. Haycock, M.C. Borges, K. Burrows, *et al.*, "The association between genetically elevated polyunsaturated fatty acids and risk of cancer". *eBiomedicine*. vol.91: 104510, 2023. <https://doi.org/10.1016/j.ebiom.2023.104510>
- [17] E.K. Aglago, I. Huybrechts, N. Murphy, *et al.*, "Consumption of Fish and Long-chain n-3 Polyunsaturated Fatty Acids Is Associated With Reduced Risk of Colorectal Cancer in a Large European Cohort". *Clinical Gastroenterology and Hepatology*. vol. 18, no. 3, pp. 654-666.e6. 2020. <https://doi.org/10.1016/j.cgh.2019.06.031>

- [18] H. Jiang, X. Shi, T.Y. Fan, *et al.*, “Dietary omega-3 polyunsaturated fatty acids and fish intake and risk of age-related macular degeneration” . *Clinical Nutrition*. vol.40, no. 12, pp: 5662-5673. 2021. [https://www.clinicalnutritionjournal.com/article/S0261-5614\(21\)00474-X/fulltext](https://www.clinicalnutritionjournal.com/article/S0261-5614(21)00474-X/fulltext)
- [19] M.A. Rincón-Cervera, N.V. González-Barriga, R. Valenzuela, *et al.*, “Profile and distribution of fatty acids in edible parts of commonly consumed marine fishes in Chile”, *Food Chemistry*, vol. 274, pp. 123-129, 2018. <https://doi.org/10.1016/j.foodchem.2018.08.113>
- [20] S. Sun, T. Ren, X. Li., *et al.*, “Polyunsaturated fatty acids synthesized by freshwater fish: A new insight to the roles of elovl2 and elovl5 in vivo”, *Biochemical and Biophysical Research Communications*, vol. 532, pp. 414-419, 2020. <https://doi.org/10.1016/j.bbrc.2020.08.074>
- [21] M.B. Behyar, M. Hasanzadeh, F. Seidi, *et al.*, “Sensing of amino acids: Critical role of nanomaterials for the efficient biomedical analysis”, *Microchemical Journal*, vol. 188, pp. 108452, 2023 <https://doi.org/10.1016/j.microc.2023.108452>
- [22] P. Shi, K. Liao, J. Xu, *et al.*, “Eicosapentaenoic acid mitigates palmitic acid-induced heat shock response, inflammation and repair processes in fish intestine”, *Fish & Shellfish Immunology*, vol. 124, pp. 362-371, 2022. <https://doi.org/10.1016/j.fsi.2022.04.011>
- [23] J. Wen, L. Zeng, Y. Xu, *et al.*, “Proximate composition, amino acid and fatty acid composition of fish maws”, *Natural Product Research: Formerly*, vol. 30, no. 2, pp. 214-217, 2015. <https://doi.org/10.1080/14786419.2015.1040790>
- [24] F.O.S. Duarte, F.G. de Paula, C.S. Prado, *et al.*, “Better fatty acids profile in fillets of Nile Tilapia (*Oreochromis niloticus*) supplemented with fish oil”, *Aquaculture*, Vol. 534, pp. 736241, 2021. <https://doi.org/10.1016/j.aquaculture.2020.736241>
- [25] K.L. Weaver, P. Ivester, J.A.Chilton, *et al.*, “The Content of Favorable and Unfavorable Polyunsaturated Fatty Acids Found in Commonly Eaten Fish”, *Journal of the American Dietetic Association*, vol. 108, No. 7 , pp. 1178–1185, 2008. <https://doi.org/10.1016/j.jada.2008.04.023>
- [26] AOAC, “Official Methods of Analysis. 15th Edn., Association of Official Analytical Chemists (AOAC)”, Washington, DC., USA.1990. Reference Source
- [27] S.A.Cohen, “Amino acid analysis using pre-column derivatization with 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate”, *Protein Sequencing Protocols*, vol. 211, pp. 143-54, 2003. <https://doi.org/10.1385/1-59259-047-0:039>
- [28] M.A. Rajion, J.G. McLean, R.N. Cahill, “Essential fatty acid metabolism in the fetal and neonatal lamb”, *Australian Journal of Biological Sciences*, vol. 38, no. 1, pp. 33-40, 1985. [Essential fatty acids in the fetal and newborn lamb - PubMed \(nih.gov\)](https://pubmed.ncbi.nlm.nih.gov/2708000/)
- [29] T.L. Ulbricht, D.A. Southgate, “Coronary heart disease: Seven dietary factors”, *The Lancet*, Vol 338, no. 8773, pp. 985–992, 1991. [https://doi.org/10.1016/0140-6736\(91\)91846-M](https://doi.org/10.1016/0140-6736(91)91846-M)
- [30] D.B. Duncan, “Multiple ranges and multiple F tests”, *Biometrics*, vol. 11, pp. 1–42, 1955. <https://doi.org/10.2307/3001478>
- [31] R. Froese, (). “Cube law, condition factor and weight–length relationships: History, meta-analysis and recommendations”. *Journal of Applied Ichthyology*, vol. 22, no. 4, pp. 241-253, 2006. <https://doi.org/10.1111/j.1439-0426.2006.00805.x>

- [32] O.O.Famoofo, & W.O. Abdul, “Biometry, condition factors and length-weight relationships of sixteen fish species in Iwopin fresh-water ecotype of Lekki Lagoon, Ogun State, Southwest Nigeria”, *Heliyon*, vol. 6, no. 1, 2020. <https://doi.org/10.1016/j.heliyon.2019.e02957>
- [33] T.B. Bagenal, F.W. Tesch, “Age and Growth. In: Bagenal, T., Ed., Methods for Assessment of Fish Production in Fresh Waters”, *Blackwell Science Publications*, pp. 101-136, 1978.
- [34] E.O. Ahmed, M.E. Ali, A.A. Aziz, “Length-weight relationships and Condition factors of six fish species in Atbara River and Khashm el-girba Reservoir, Sudan”, *International Journal of Agriculture Sciences*, vol. 3, no.1, pp. 65–70, 2011. <http://dx.doi.org/10.9735/0975-3710.3.1.65-70>
- [35] N.C. Ujjania, M.P.S. Kohli, L.L. Sharma, “Length-weight relationship and condition factors of Indian major carps (C. catla, L. rohita and C. mrigala) in Mahi Bajaj Sagar, India”, *Research Journal of Biology*, vol. 2, no.1, pp. 30-36, 2012. <https://doi.org/10.21077/ijf.2017.64.special-issue.76263-27>
- [36] J.P. Bonjour, “Calcium and phosphate: A duet of ions playing for bone health”, *Journal of the American College of Nutrition*, vol. 30, no. 5, pp. 438S-48S, 2011. <https://doi.org/10.1080/07315724.2011.10719988>
- [37] T. Tian, Y. Chu, Y. Yang, *et al.*, “Phosphates as Energy Sources to Expand Metabolic Networks”, *Life*, Vol. 9, no. 2, pp. 43, 2019. <https://doi.org/10.3390/life9020043>
- [38] K. Kritmetapak, R. Kumar, “Phosphate as a Signaling Molecule”, *Calcified tissue international*, Vol. 108, no. 1, pp. 16, 2021. <https://doi.org/10.1007/s00223-019-00636-8>
- [39] M.W. Berchtold, H. Brinkmeier, M. Müntener, “Calcium Ion in Skeletal Muscle: Its Crucial Role for Muscle Function, Plasticity, and Disease”, *Physiological Reviews*, Vol. 80, no. 3, 2000. <https://doi.org/10.1152/physrev.2000.80.3.1215>
- [40] S. Singh, J. Dodt, P.Volkers, *et al.*, “Structure functional insights into calcium binding during the activation of coagulation factor XIII A”. *Scientific Reports*, vol. 9, no. 11324, 2019. <https://doi.org/10.1038/s41598-019-47815-z>
- [41] A.A. Hassan, A.I. Mamman, , S. Adaji, *et al.*, “Anemia and iron deficiency in pregnant women in Zaria, Nigeria”, *Sub-Saharan African Journal of Medicine*, vo.1, no.1, pp. 36-39, 2014. <http://dx.doi.org/10.4103/2384-5147.129311>
- [42] I.M. Okafor, D.C. Okpokam, A.B. Antai, *et al.*, “Iron status of pregnant women in rural and urban communities of cross river state, South-South Nigeria”. *Nigerian journal of physiological sciences : official publication of the Physiological Society of Nigeria*, vol. 31, pp. 121-125, 2016. <https://pubmed.ncbi.nlm.nih.gov/28262847/>
- [43] M. Maywald, L. Rink, “Zinc in Human Health and Infectious Diseases”, *Biomolecules*, vol. 12 no. 12, pp. 1748, 2022. <https://doi.org/10.3390/biom12121748>
- [44] B.P. Mohanty, T.V. Sankar, S. Ganguly, *et al.*, “Micronutrient composition of 35 food fishes from India and their significance in human nutrition”, *Biological Trace Element Research*, vol. 174, pp. 448-458, 2016. <https://doi.org/10.1007/s12011-016-0714-3>
- [45] B. Abraha, H. Admassu, A. Mahmud, *et al.*, “Effect of processing methods on nutritional and physico-chemical composition of fish: a review”, *Food Processing and Technology*, vol. 6, no. 4, pp. 376-382, 2018. <https://doi.org/10.15406/mojfpt.2018.06.00191>

- [46] S. Gupta, B.S. Gowri, A.J. Lakshmi, *et al.*, “Retention of nutrients in green leafy vegetables on dehydration”, *Journal of Food Science and Technology*, vol. 50, no. 5, pp. 918-925, 2013. <https://doi.org/10.1007/s13197-011-0407-z>
- [47] S. NurSyahirah, A. Rozzamri, “Effects of frying on fish, fish products and frying oil – a review”, *Food Research*, vol. 6, no. 5, pp. 14-32, 2022. [https://doi.org/10.26656/fr.2017.6\(5\).608](https://doi.org/10.26656/fr.2017.6(5).608)
- [48] F. Shahidi, A. Hossain, “Role of Lipids in Food Flavor Generation”. *Molecules*, vol. 27, no.15, 2022. <https://doi.org/10.3390/molecules27155014>
- [49] E.F. Fernandes, , M.A.S. Vasconcelos, , M.d.A. Ribeiro, *et al.*, “Nutritional and lipid profiles in marine fish species from Brazil”, *Food Chemistri*, vol. 160, pp. 67-71, 2014. <https://doi.org/10.1016/j.foodchem.2014.03.055>
- [50] A.A. Nurnadia, , A. Azrina, I. Amin, “Proximate composition and energetic value of selected marine fish and shellfish from the west coast of peninsular Malaysia”. *International Food Research Journal*. Vol 18, pp. 137-148, 2011. <https://core.ac.uk/download/pdf/153807129.pdf>
- [51] B.P. Mohanty, A. Mahanty, S. Ganguly, *et al.*, “Nutritional composition of food fishes and their importance in providing food and nutritional security”, *Food Chemistry*, vol. 293, pp. 561-570, 2019. <https://doi.org/10.1016/j.foodchem.2017.11.039>
- [52] F. Jabeen, , A.A. Chudhry, “Chemical composition and fatty acid profiles of three freshwater species”, *Food Chemistry*, vol. 125, pp. 991-996, 2011. <https://doi.org/10.1016/j.foodchem.2010.09.103>
- [53] S.C. Dyall, , L. Balas, , N.G. Bazan, *et al.*, “Polyunsaturated fatty acids and fatty acid-derived lipid mediators: Recent advances in the understanding of their biosynthesis, structures and functions”, *Progress in Lipid Research*. vol. 86, pp. 101165, 2022. <https://doi.org/10.1016/j.plipres.2022.101165>
- [54] N. Kim, , U.D.Sohn, , V. Mangannan, *et al.*, “Leukotrienes in Acetylcholine-Induced Contraction of Esophageal Circular Smooth Muscle in Experimental”, *Gastroenterology*, vol. 112, no.5, pp. 1548–1558, 1997. [https://doi.org/10.1016/S0016-5085\(97\)70036-2](https://doi.org/10.1016/S0016-5085(97)70036-2)
- [55] A. Zuraini, M.N. Somchit, M.H. Solihah, *et al.*, “Fatty acid and amino acid composition of three local Malaysian Channa spp. Fish”, *Food Chemistry*, vol. 97, no. 4, pp. 674-678, 2005. <https://doi.org/10.1016/j.foodchem.2005.04.031>
- [56] Carballo-Casla, Garcia-Esguines, J.R. Banegas, *et al.*, “Fish consumption, omega-3 fatty acid intake, and risk of pain: the Seniors-ENRICA-1 cohort”, *Clinical Nutrition*, vol. 41, no. 11, pp. 2587-2595, 2022. <https://doi.org/10.1016/j.clnu.2022.09.007>
- [57] D. Swanson, R. Block, , S.A.Mousa, “Omega-3 Fatty Acids EPA and DHA: Health Benefits Throughout Life”. *Advances in Nutrition*, vol. 3, no.1, pp. 1-7,2012. <https://doi.org/10.3945/an.111.000893>
- [58] R.P.Mason , S.C.R. Sherratt, , L.H. Eckel, “Omega-3-fatty acids: Do they prevent cardiovascular disease?”, *Best Practice & Research Clinical Endocrinology & Metabolism*, vol.37, no 3, 101681, 2023. <https://doi.org/10.1016/j.beem.2022.101681>.
- [59] M. Rondanelli, M.A. Faliva, , A. Miccono, *et al.*, “Food pyramid for subjects with chronic pain: foods and dietary constituents as anti-inflammatory and antioxidant agents”, *Nutrition*

- Research Reviews*, vol. 31, no. 1, pp. 131-151, 2018. <https://doi.org/10.1016/j.endien.2020.01.010>
- [60] D. Karageorgou, U. Rova, , P. Christakopoulos, *et al.*, “Benefits of supplementation with microbial omega-3 fatty acids on human health and the current market scenario for fish-free omega-3 fatty acid”, *Trends in Food Science & Technology*, vol. 136, pp. 169-180, 2023. <https://doi.org/10.1016/j.tifs.2023.04.018>
- [61] J. M. Seddon, , J. Cote, , N. Davis, *et al.*, “Progression of age-related macular degeneration. Progression of age-related macular degeneration”. *Arch Ophthalmol*. Vol. 121, no. 6, pp. 785-792, 2003. <https://doi.org/10.1001/archophth.121.6.785>
- [62] B.P. Mohanty, A. Mahanty, , S. Ganguly, *et al.* “Amino Acid compositions of 27 food fishes and their importance in clinical nutrition”, *Journal Amino Acids*, vol. 2014, 2014. <https://doi.org/10.1155/2014/269797>
- [63] H. Turan, G. Sonmez, Y. Kaya, “Fatty acid profile and proximate composition of the thornback ray (*Raja clavata*, L. 1758) from the Sinop coast in the Black Sea”, *Journal of Fisheries Sciences*, vol. 1, no. 2, pp. 97–103, 2007. <https://doi.org/10.3153/jfscm.2007012>
- [64] FAO/WHO: Fat and Oils in Human Nutrition: Report of a Joint Expert Consultation. FAO/WHO, Rome, Italy.
- [65] C.E. Fernandes, M.A.da.S. Vasconcelos, , M.L.A. Sarubbo, *et al.*, Nutritional and lipid profiles in marine fish species from Brazil, *Food Chemistry*, vol. 160, no. 67-71, 2014. <https://doi.org/10.1016/j.foodchem.2014.03.055>
- [66] L. Day, J.A. Cakebread, S.M. Loveday, “Food proteins from animals and plants: Differences in the nutritional and functional properties”, *Trends in Food Science & Technology*, vol. 119, pp. 428-442, 2022. <https://doi.org/10.1016/j.tifs.2021.12.020>
- [67] L. Wu, A. Wang, , R. Shen, *et al.*, “Effect of processing on the contents of amino acids and fatty acids, and glucose release from the starch of quinoa”, *Food Science & Nutrition*, vol. 8, no. 9, pp. 4877-4887, 2020. <https://doi.org/10.1002/fsn3.1775>
- [68] S.M. Pawar, S.R. Sonaware, “Fish muscle protein highest source of energy”. *International Journal of Biodiversity and Conservation*. Vol. 5, no. 7, pp. 433-435, 2013. <https://doi.org/10.5897/IJBC12.043>
- [69] F. Spallotta, , C. Cencioni, , S. Straino, *et al.*, “Enhancement of lysine acetylation accelerates wound repair”. *Communicative & Integrative Biology*, vol. 6, no. 5, 2013. <https://doi.org/10.4161/cib.25466>
- [70] M. Smriga, T. Ando, M. Akutsu, *et al.*, “Oral treatment with L-lysine and L-arginine reduces anxiety and basal cortisol levels in healthy humans”, *Biomedical Research*, vol. 28, no.2, pp. 85-90, 2007. <https://doi.org/10.2220/biomedres.28.85>
- [71] M.C. Pedrazini, M.H. da Silva, F.C. Groppo, “L-lysine: Its antagonism with L-arginine in controlling viral infection. Narrative literature review, *British Journal of Clinical Pharmacology*. vol. 88, no. 11, pp. 4708-4723, 2022. <https://doi.org/10.1111/bcp.15444>
- [72] D. Tomé, C. Bos, “Lysine requirement through the human life cycle”, *The Journal of Nutrition*, vol. 137, no.6, pp. 1642S–1645S, 2007. <https://doi.org/10.1093/jn/137.6.1642S>
- [73] M. Watford, “Glutamine and glutamate: Nonessential or essential amino acids?”, *Animal Nutrition*, vol. 1, no. 3, pp. 119-122, 2015. <https://doi.org/10.1016/j.aninu.2015.08.008>

- [74] X. Li, S. Zheng, G. Wu, “Nutrition and metabolism of glutamate and glutamine in fish”, *Amino Acids*. vol.52 , no.5, pp. 671-691, 2020.
<https://doi.org/10.1007/s00726-020-02851-2>
- [75] M. Li, , Y. Wu, , L. Ye, “ The Role of Amino Acids in Endothelial Biology and Function”, *Cells*, vol.11, no.8, pp. 1372, 2022. <https://doi.org/10.3390/cells11081372>
- [76] M.M. Pal, “Glutamate: The Master Neurotransmitter and Its Implications in Chronic Stress and Mood Disorders”, *Frontiers in Human Neuroscience*, vol. 15, pp. 722323.
<https://doi.org/10.3389/fnhum.2021.722323>
- [77] W. Wang, X. Zhou, Y. Liu, “Characterization and evaluation of umami taste: A review”, *TrAC Trends in Analytical Chemistry*, vol. 127, pp. 115876, 2020.
<https://doi.org/10.1016/j.trac.2020.115876>
- [78] C. Marques, A. Reis, C. Moura, *et al.*, “Consumer insight into the monosodium glutamate”, *Acta Scientiarum. Technology*. vol. 40, 2018.
<https://doi.org/10.4025/actascitechnol.v40i1.30838>
- [79] K. Stańska, , A. Krzeski, “The umami taste: from discovery to clinical use”. *Polish Journal of Otolaryngology*. vol. 70, no.4, pp. 10–15, 2016.
<http://dx.doi.org/10.5604/00306657.1199991>



Webmail
Univ. Bung Hatta

hafrizal syandri <syandri_1960@bunghatta.ac.id>

6639837: Update on manuscript

1 message

International Journal of Food Science <ijfs@hindawi.com>

Fri, Nov 17, 2023 at 12:57 PM

Reply-To: International Journal of Food Science <deepanaa.rajadurai@hindawi.com>

To: Hafrijal Syandri <syandri_1960@bunghatta.ac.id>



Hindawi

Dear Dr. Hafrijal Syandri,

I am writing regarding your manuscript 6639837, entitled Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security . It is taking us longer than expected to send you an editorial decision for your manuscript.

As standard practice, we perform rigorous peer review integrity checks as part of the editorial process. These checks can sometimes incur delays.

We wanted to inform you that we are still working to send you an editorial decision as soon as possible. You can track the status of your manuscript at the link below:

[MANUSCRIPT DETAILS](#)

Please accept our apologies for any inconvenience.

Kind regards,
Rajadurai Deepanaa
International Journal of Food Science

This email was sent to syandri_1960@bunghatta.ac.id. You have received this email in regards to the account creation, submission, or peer review process of a paper submitted to a journal published by Hindawi Limited.

Hindawi Limited, 3rd Floor, Adam House, [1 Fitzroy Square, London, W1T 5HF, United Kingdom](#).

Hindawi respects your right to privacy. Please see our [privacy policy](#) for information on how we store, process, and safeguard your data.

[Unsubscribe](#)

Research Article

Analysis of Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security

Azrita Azrita¹, Hafrijal Syandri^{*}, Hazlina Ahamad Zakeri², Harfiandri Damanhuri¹,
Netti Aryani³

¹Faculty of Fisheries and Marine Science, Universitas Bung Hatta, 25131, Padang-West Sumatra Province, Indonesia

²Faculty of Science and Marine Environment, Universiti Malaysia Terengganu, Malaysia.

³Faculty of Fisheries and Marine Science, Universitas Riau, 28293, Pekanbaru-Riau Province, Indonesia

*Corresponding author: syandri_1960@bunghatta.ac.id

Abstract

Fish have become an irreplaceable dietary source of animal protein, especially among households with low socioeconomic status in rural and urban areas of Indonesia. This study analysed the nutrient composition of three local Bagridae fish species in Indonesia. The nutrient composition results of *Hemibagrus nemurus*, *Hemibagrus wyckii*, and *Mystus nigriceps* revealed that the protein content was 24.26%, 22.57%, and 21.39% (% dry weight), respectively. The total lipid content varied between 6.64–7.75%, whereas the crude ash content ranged from 1.59–2.30%. Regarding mineral contents, the calcium levels ranged from 1.49–1.66 mg/g, iron levels from 28.35–40.36 µg/g, and zinc levels from 24.03–54.46 µg/g. The predominant amino acids found in the three species of Bagridae fish were glutamic acid, aspartic acid, alanine, arginine, and lysine, with their concentrations ranging from 9.10–24.34%. Among the fatty acids, C16:0 (palmitic acid) was the most abundant in all three species, accounting for 25.59–30.70% of the total fatty acids. Additionally, significant amounts of C18:1 (oleic acid), C18:0 (stearic acid), and C20:4 (arachidonic acid) were also detected as primary fatty acids. The calculated atherogenic index values in the three species of Bagridae fish ranged from 0.73–0.99, while the thrombogenic index values varied between 0.54–0.75. These results indicate that consuming the meat of these three freshwater Bagridae fish species caught in the wild does not pose any health risks to consumers. They can be considered a safe and suitable food source with good nutritional quality.

Keywords: Nutrient composition, Amino acids, Fatty acids, Freshwater fish, Kampar Kanan River of Indonesia

1. Introduction

Fish serve as a high-quality protein source that is essential for satisfying dietary protein requirements, especially in areas where other protein sources are few or difficult to obtain [1,2]. Their accessibility can aid in ensuring a steady supply of food, particularly for communities whose main source of income is fishing or aquaculture [3]. Fish can contribute to a more varied selection of protein sources in communities, resulting in a diet that is more nutritious and balanced. Dietary fish can treat nutrient deficits and enhance general nutritional health. Fish are generally considered safe and offer a lower risk of contamination with harmful substances compared to some land-based animal products. A reliable indicator of a fish's quality, nutritional value, physiological status, and environment is its chemical makeup, which is found in fish flesh [4].

Hemibagrus nemurus, locally known as "baung", *Hemibagrus wyckii*, known as "geso" and *Mystus nigriceps*, known as "ingir-ingir," are three species of catfish found in the Kmpar Kanan River, Indonesia [5,6]. Due to their high market value, these species have been identified as significant economic resources for rural communities [7]. Therefore, the analysis of the nutritional quality of fish meat in the study area is very important to determine whether it is beneficial to human health.

Knowing the composition of fatty acids and amino acids in fish meat is an essential factor that should not be ignored [8]. This information provides valuable insight into the nutritional quality of fish and the values of more nutritious and healthy foods [9]. In addition, a better understanding of the composition of fatty acids and amino acids can assist in selecting a suitable feed [10], developing a balanced diet [11], and maintaining optimal fish meat nutrition [12].

Omega-3 polyunsaturated fatty acids (PUFAs), particularly docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA) derived from fish oil, exhibit anti-inflammatory and anti-gastrointestinal cancer properties, making them crucial as immune-boosting nutrients [13,14]. Studies have demonstrated that ω -3 and ω -6 PUFAs exhibit beneficial effects in the management of cardiovascular diseases and cancers [15,16]. The fatty acid composition of PUFAs can differ among different types of fish, including both freshwater and marine species [17, 18]. Amino acids (AAs), such as cysteine, arginine, tyrosine, glycine, proline, and serine,

play a vital role in illness and stress situations and in the prevention of inflammation and repair processes in fish intestines [19, 20].

Furthermore, knowledge of the atherogenicity index (AI) and thrombogenicity index (TI) of fatty acids in the three local freshwater fish species of Bagridae can contribute to improving dietary guidelines. The AI and TI can be used to develop recommendations aimed at improving heart health [21,22]. These recommendations may involve selecting fish with favourable fatty acid profiles, allowing individuals to choose the most suitable fish species based on their specific needs [17, 23]. Therefore, this study aims to examine the nutritional composition profile, mineral content, amino acids, and fatty acids, and AI and TI of three freshwater fish species, namely, *H. nemurus*, *H. wyckii*, and *M. nigriceps*. These three fish species are generally consumed by people who live around the Kampar Kanan River in Kampar Regency, Riau Province, Indonesia.

2. Material and methods

2.1. Materials

In this study, a total of 27 Bagridae fish belonging to three distinct species (*H. nemurus*, *H. wyckii*, and *M. nigriceps*) were examined. The specimens were obtained from local fishers operating in the upper stretches of the Kampar Kanan River in Kampar Regency, Riau Province (Figure 1). Sampling took place between June and August 2022 and was verified by the Fisheries Department, Faculty of Fisheries and Marine Science, Universitas Bung Hatta, Indonesia. The samples were placed in plastic bags and transported to the laboratory in an insulated icebox. The samples were then grouped based on their respective sampling locations.

2.2. Biometry measurement

Upon reaching the laboratory within 10 hours, the fish samples were subjected to gutting and washing. Each specimen was weighed (TW) and measured to determine the standard length (SL) as well as the depth or maximum height (H). TW was measured using balance scale (OHAUS Model CT 6000 USA) with a precision of 0.01 g. The length measurement was taken from the tip of the mouth to the end of the upper lobe of the caudal fin, representing the total body length, using a metre ruler with an accuracy of 1 mm. The height measurement involved a vertical assessment of the body's maximum height measured using a

Digital Sekhmet Sigma Vernier calliper with an accuracy of 1 mm. The condition factor (CF) was calculated using the formula $CF = (TW/SL^3) \times 100$.

2.3. Proximate, amino acid, and mineral content analysis

The proximate carcass composition was assessed following the AOAC standard methods [24]. The samples were dried to a constant weight at 105 °C. The crude protein content was analysed using the standard Kjeldahl method, which involves multiplying the nitrogen content by a factor of 6.25. The Soxhlet method with ether extraction was employed to analyse crude lipids. The ash content was obtained by incinerating the samples at 550 °C for 16 h. The amino acid composition was determined using a high-performance liquid chromatography (HPLC) system, which consisted of a Waters® 1525 binary HPLC pump, Waters® 717 autosamplers, and Waters® 2475 multi- λ fluorescence detector (with excitation at 250 nm and emission at 395 nm). The samples were hydrolysed in triplicate with 6 N hydrochloric acid for 24 h at 11 °C [25].

For mineral data composition (Na, Mg, Ca, K, P, Fe, Cu, Mn, and Zn), the ashed feed sample and carcass were dissolved in 1 ml of hydrochloric acid (35% v/v Suprapur®, Merck) and then filtered with cellulose filter paper (Watchman No 1, International Ltd; Maidstone, UK) and diluted to an appropriate concentration for each elemental mineral. P levels were analysed with a Perkin-Elmer AA spectrophotometer mod 3110 (Norwalk, CT, USA).

2.4. Fatty acid analysis

The fish meat was examined utilizing the gas chromatography–mass spectrometry (GC–MS) method. The method of Folch et al. (1957) modified by Rajion et al. [26] was employed to perform the total fat extraction. This involved using a solvent system consisting of chloroform and methanol at a 2:1 (v/v) ratio. The process of transmethylation was conducted using a solution of 14% methanolic boron trifluoride. The fatty acid composition of the meat was then analysed at the SIG Laboratory, Accredited Testing Laboratory-LP-184-IDN.

The nutritional quality of lipids (AI and TI) was calculated using the following equations [27]:

$$AI = \frac{[(C12:0 + 4 \times C14:0 + C16:0)]}{\sum MUFA + \sum n-6 + \sum n-3}$$

$$TI = \frac{[C14:0 + C16:0 + C18:0]}{[(0.5 \times \sum MUFA) + (0.5 \times \sum n-6) + (3 \times \sum n-3) + (\sum n-3 / \sum n-6)]}$$

where:

AI = Atherogenic index

TI = Thrombogenic index

C12:0 = lauric acid

C14:0 = myristic acid

C16:0 = palmitic acid

C18:0 = stearic acid

\sum MUFA = sum of the concentrations of all monosaturated fatty acids

\sum n-6 = sum of the concentrations of n-6 polyunsaturated fatty acids

\sum n-3 = sum of the concentrations of n-3 polyunsaturated fatty acids

2.5. Data analysis

Data analysis was performed using the SPSS 16.0 software package (SPSS; Chicago, IL). The homogeneity of variances was assessed using Levene's test. One-way ANOVA was carried out to determine the parameters of proximate and amino acid composition, including the composition of fatty acids and nutritional quality of lipids (AI and TI) for each species, followed by the post hoc Duncan's multiple range test [28]. The results are reported as the mean values \pm standard errors for each parameter.

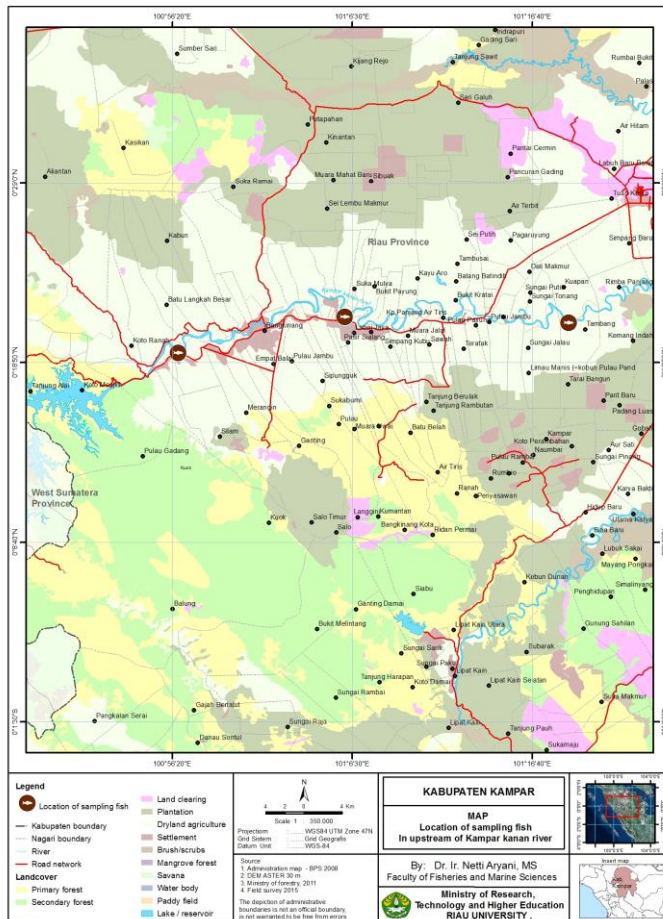


Figure 1. Map of Kampar Regency, Riau Province, and sampling locations of the three species of Bagridae fish.

Formatted: Highlight

Commented [S1]: Figure 1 has been revised

Formatted: Highlight

3. Results

Table 1 presents the average wet weight, standard length, height, and the results of meat nutritional composition analysis for three indigenous Indonesian freshwater Bagridae species. The moisture content (% wet weight) ranged from 82.40– 85.39%. Among the species examined, *M. nigriceps* exhibited the lowest protein content at 21.39%, while *H. nemurus* displayed the highest value at 24.26%. However, *M. nigriceps* had a higher mineral content, including of iron and zinc.

Table 2 summarizes the fatty acid composition (%total fatty acid) for the three Bagridae fish species. C16:0 was the predominant fatty acid in the three species of Bagridae fish, with percentages ranging from 25.09–30.71%. The additional prominent fatty acids found were C18:1, C18:0, and C22:6. *H. nemurus* exhibited a ω -6/ ω -3 ratio below 1 (0.82), while *H. wyckii* and *M. nigriceps* had ratios of 1.11 and 1.02, respectively. The obtained PUFA/saturated fatty acid (SFA) (P/S) ratio varied between 0.61 and 0.76. The AI values varied between 0.76 and 0.99 and the TI between 0.54 and 0.75 (**Table 3**).

Table 4 presents the amino acid composition (% of total protein) of the three Bagridae species. Glutamic acid was the dominant amino acid, ranging from 19.29–24.34%, followed by aspartic acid, which ranged from 9.21–11.27%. The lysine content was consistent, ranging from 9.67–9.87%. In *H. nemurus*, the levels of various amino acids ranged from 0.86–24.35%. Similarly, in *H. wyckii*, the amino acid levels ranged from 0.76–21.58%, while in *M. nigriceps*, the levels ranged from 0.74–19.29%.

Table 1. Results of biometric, proximate and mineral composition of three species of Bagridae fish

	Species		
	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
<i>Biometric measurements</i>			
Total weight (g)	389.99 ± 24.96 ^a	1,390.33 ± 168.29 ^b	17.57 ± 1.53 ^c
Standard length (cm)	28.16 ± 0.53 ^a	44.14 ± 1.98 ^b	10.29 ± 0.15 ^c
Height (cm)	8.44 ± 0.16 ^a	8.82 ± 0.39 ^b	2.57 ± 0.03 ^c
Condition factor	1.63 ± 0.08	1.58 ± 0.07	1.56 ± 0.08
<i>Proximate composition</i>			
Crude protein (% DW)	24.26 ± 0.87 ^a	22.57 ± 0.37 ^b	21.39 ± 0.15 ^c
Crude fat (% DW)	6.64 ± 0.03 ^a	7.47 ± 0.02 ^b	7.75 ± 0.40 ^c
Crude ash (% DW)	1.94 ± 0.02 ^a	2.30 ± 0.09 ^b	1.59 ± 0.02 ^c
Moisture (% WW)	82.40 ± 2.51	85.39 ± 2.36	83.75 ± 2.21
<i>Mineral composition</i>			
Macrominerals (mg/g)			
Sodium	0.99 ± 0.00 ^a	1.61 ± 0.01 ^b	1.61 ± 0.02 ^c
Magnesium	1.12 ± 0.01 ^a	0.57 ± 0.02 ^b	1.15 ± 0.01 ^c
Calcium	1.66 ± 0.00 ^a	1.55 ± 0.02 ^b	1.49 ± 0.04 ^c
Potassium	0.71 ± 0.00 ^a	0.55 ± 0.02 ^b	0.43 ± 0.01 ^c
Phosphorous	7.03 ± 0.03 ^a	2.74 ± 0.02 ^b	6.45 ± 0.07 ^c
Microminerals (µg/g)			
Iron	28.30 ± 0.11 ^a	28.73 ± 0.08 ^b	40.36±0.55 ^c
Copper	8.93 ± 0.03 ^a	7.46 ± 0.09 ^b	6.21 ± 0.32 ^c

Manganese	1.64 ± 0.02 ^a	2.64 ± 0.02 ^b	2.84 ± 0.02 ^c
Zinc	24.03 ± 0.45 ^a	24.61 ± 0.19 ^b	54.46 ± 0.17 ^c

Values are mean % ± SE; *Values in the same row followed by different letters are significantly different (p<0.05).

Table 2. Fatty acid composition of three species of Bagridae fish

Fatty acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
C12:0 (lauric acid)	1.97 ± 0.05 ^a	3.94 ± 0.01 ^b	4.27 ± 0.01 ^c
C14:0 (Meristic acid)	2.38 ± 0.03 ^a	2.72 ± 0.08 ^b	2.89 ± 0.01 ^c
C16:0 (Palmitic acid)	27.23 ± 0.06 ^a	30.70 ± 0.06 ^b	25.59 ± 0.01 ^c
C18:0 (Stearic acid)	16.17 ± 0.02 ^a	13.54 ± 0.02 ^b	10.64 ± 0.01 ^c
C20:0 (Arachidic acid)	0.23 ± 0.01 ^a	0.14 ± 0.01 ^b	0.37 ± 0.01 ^c
C16:1 (Palmitoleic acid)	1.78 ± 0.00 ^a	1.27 ± 0.04 ^b	4.93 ± 0.01 ^c
C18:1(Oleic acid)	16.86 ± 0.01 ^a	16.18 ± 0.01 ^b	16.97 ± 0.01 ^c
C18:2 (Linoleic acid)	3.24 ± 0.02 ^a	3.84 ± 0.33 ^b	4.55 ± 0.001 ^c
C18:3 (Linolenic acid)	1.12 ± 0.01 ^a	0.82 ± 0.01 ^b	1.57 ± 0.00 ^c
C20:4 (Arachidonic acid)	12.13 ± 0.01 ^a	12.49 ± 0.09 ^b	12.41 ± 0.02 ^c
C20:5 (Eicosapentaenoic acid; EPA)	2.06 ± 0.08 ^a	2.34 ± 0.04 ^b	1.35 ± 0.01 ^c
C22:6 (Docosahexaenoic acid; DHA)	15.65 ± 0.01 ^a	11.56 ± 0.04 ^b	13.64 ± 0.02 ^c

Values are mean % ± SE; *Values in the same row with a different superscript are significantly different (p<0.05).

Table 3. Fatty acid ω-6/ω-3 and polyunsaturated/saturated fatty acid ratio of three species of Bagridae fish

	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
ω-6/ω-3 ratio	0.82 ± 0.00 ^a	1.11 ± 0.02 ^b	1.02 ± 0.01 ^c
P/S ratio	0.69 ± 0.01 ^a	0.61 ± 0.06 ^b	0.76 ± 0.00 ^c
AI	0.87 ± 0.01 ^a	0.99 ± 0.06 ^b	0.73 ± 0.00 ^c
TI	0.63 ± 0.01 ^a	0.75 ± 0.03 ^b	0.54 ± 0.00 ^c

P/S, polyunsaturated/saturated fatty acid; AI, atherogenic index; TI, thrombogenic index
Values are mean % ± SE; *Values in the same row followed by different letters are significantly different (p<0.05).

Table 4. Amino acid composition of three species of Bagridae fish

Amino acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
Aspartic acid	10.39 ± 0.01 ^a	9.61 ± 0.04 ^b	11.28 ± 0.02 ^c
Glutamic acid	24.34 ± 0.00 ^a	21.58 ± 0.41 ^b	19.29 ± 0.03 ^c
Serine	3.74 ± 0.00 ^a	5.40 ± 0.03 ^b	7.50 ± 0.06 ^c
Glycine	4.02 ± 0.00 ^a	5.77 ± 0.04 ^b	4.69 ± 0.02 ^c
Histidine	2.39 ± 0.00 ^a	2.90 ± 0.03 ^b	2.21 ± 0.02 ^c
Arginine	6.17 ± 0.01 ^a	8.27 ± 0.03 ^b	6.22 ± 0.03 ^c
Threonine	4.49 ± 0.01 ^a	4.19 ± 0.03 ^b	4.39 ± 0.04 ^c
Alanine	7.03 ± 0.00 ^a	6.04 ± 0.02 ^b	7.89 ± 0.01 ^c
Proline	2.40 ± 0.00 ^a	2.71 ± 0.01 ^b	2.21 ± 0.02 ^c
Tyrosine	3.91 ± 0.01 ^a	3.29 ± 0.06 ^b	3.82 ± 0.04 ^c
Valine	3.73 ± 0.00 ^a	4.02 ± 0.01 ^b	3.85 ± 0.04 ^c
Methionine	3.42 ± 0.00 ^a	1.96 ± 0.02 ^b	1.27 ± 0.03 ^c
Cystine	0.87 ± 0.00 ^a	0.76 ± 0.02 ^b	0.75 ± 0.03 ^c
Isoleucine	3.82 ± 0.001 ^a	3.96 ± 0.003 ^b	3.66 ± 0.002 ^c
Leucine	6.15 ± 0.000 ^a	6.69 ± 0.002 ^b	6.28 ± 0.004 ^c
Phenylalanine	5.13 ± 0.000 ^a	6.03 ± 0.005 ^b	5.18 ± 0.027 ^c
Lysine	9.61 ± 0.001 ^a	9.82 ± 0.026 ^b	9.87 ± 0.003 ^c

Values are mean % ± SE; *Values in the same row followed by different letters are significantly different (p<0.05).

4. Discussion

The results of this study provide important new information about the nutritional makeup of the studied species of Bagridae fish. The biometric measures of the fish yielded important information about their physical traits and health. Researchers are able to determine the average weight of fish at various lengths by examining the link between length and weight for a particular species. This knowledge is essential for understanding the trends in fish population growth, identifying potential shifts in growth rates over time, and assessing the general health and welfare of fish [29]. A variety of factors, including sex, age, maturity level, size, stomach fullness level, sampling strategies, sample sizes, and environmental

circumstances, have an impact on fish health and the parameters that determine length–weight connections in fish [30]. However, none of these factors were considered in our study. In general, according to the theory put forth by [Bagenal and Tesch \[31\]](#), heavier fish of a given length are in better physiological condition. The condition factor is also used in fisheries science to compare the health, fatness, and condition of fish [32]. A condition factor of > 1 is favourable, indicating a good feed nutrition level and suitable living habitat [33]. Thus, based on the findings in this study, the Kampar Kanan River provides a suitable habitat for the fish, ensuring their accessibility and availability for community consumption.

The fish samples exhibited good nutritional profiles, with high amounts of proteins and lipids as well as moisture content contributing to their overall nutritional value, according to the results of proximate analysis and mineral composition (see Table 1). These results imply that the fish species studied provide a possible source of nutrients to solve issues with food security. The study of the mineral composition revealed the existence of critical minerals such as phosphorus (P), calcium (Ca), magnesium (Mg), iron (Fe), and zinc (Zn) in the fish samples in addition to proteins and lipids. All three Bagridae fish significantly differed from each other in terms of their crude ash content and mineral profile (see Table 1). However, in general, P and Ca are the two highest minerals found in all fish. Both of these macro minerals are good for human bone and tooth health, as key components of hydroxyapatite, the mineral matrix that provides strength and structure to bones and teeth [34].

In the current study, the meat of three species of Bagridae fish from the Kampar Kanan River in Indonesia was found to contain an abundance of microminerals such as Fe and Zn. *M. nigriceps* has been reported to exhibit higher mineral content, including of Fe and Zn. This species is categorized as a small indigenous fish (SIF) with a maximum standard length of approximately 10.29 cm. According to [35], SIFs are generally known to be rich in Ca, Fe, and Mn.

In this study, all three Bagridae fish had a total fat content of approximately 7–8% by weight, with a higher water content ranging from 82.40–85.39% (Table 1). These values are similar to those found in wild and cultured sea bass *Dicentrarchus labrax* [36]. Additionally, the fish species can be grouped into different categories, including high-fat fish ($>8\%$), medium-fat fish (4–8%), low-fat fish (2–4%), and lean fish ($<2\%$) [37]. According to the classification established by [37], the three indigenous Bagridae fish species are characterized as having a moderate fat content level, which is reflected by the high levels of DHA and EPA

in the meat of *H. nemurus*, *H. wyckii*, and *M. nigriceps* (Table 2). Therefore, we recommend that consuming the meat of these three fish species can indirectly boost food security in rural and urban communities.

All three Bagridae fish species analysed in the study contained arachidonic acid (C20:4), at 12.13% for *H. nemurus*, 12.49% for *H. wyckii*, and 12.41% for *M. nigriceps*. Arachidonic acid serves as a precursor for synthesizing prostaglandins and leukotrienes [38]. Leukotrienes are crucial in allergic responses, inducing smooth muscle contraction and increasing vascular permeability [39]. Additionally, all three species also contained EPA ranging from 1.35–2.34% and DHA ranging from 11.56–15.65%. The increased consumption of omega-3 fatty acids, such as EPA and DHA, has been linked to a decreased risk of pain development in older individuals and to the prevention of Alzheimer's disease [40,41].

As determined by the current study, the ω -6/ ω -3 ratios of the three Bagridae fish species ranged from 1:0.8 to 1:0.97. It was found that all three fish species had ω -6/ ω -3 ratios within the recommended range for a typical Indonesian diet. In contrast, a typical Western diet is characterized by a high ω -6/ ω -3 ratio of 10:1 to 20:1, while a Mediterranean diet has a ratio of 4:1 to 5:1 [42]. Additionally, the P/S ratio analysis showed that the three Bagridae fish species are sources of PUFAs that meet the requirements for food safety.

The Food and Agriculture Organization (FAO) and the World Health Organization (WHO) have recommended AI and TI values ranging from 0.4 to 0.5 for promoting human health [43]. In the current study, the observed AI values in the three freshwater species of Bagridae ranged from 0.73 to 0.99, while the TI values varied between 0.54 and 0.75. This finding is associated with a notable disparity in the values of SFAs among the three Bagridae fish species. Based on these findings, we confirm that consuming the meat of the three freshwater species of Bagridae fish caught in the wild does not endanger consumer health and can serve as a suitable source of food safety and nutritional quality.

Fish with a high crude protein content, as shown by the three studied Bagridae fish (Table 1), are regarded as being nutritionally useful for human health. The protein and amino acid contents can directly indicate the nutritional quality of meat [44]. As shown in Table 4, *H. wyckii* had a higher content of essential amino acids (EAAs) than *H. nemurus* and *M. nigriceps*. Among the EAAs, lysine (Lys) was found to be present in the highest amount in all

three species. Lys is necessary for tissue growth and repair, reduced stress-induced anxiety, and the conversion of fatty acids into energy [45-47].

More than 60% of the amino acid composition in all three Brigade fish studied comes from nonessential amino acids (NEAAs), particularly glutamic acid (Table 4). Glutamic acid and its amine, glutamine, are highly abundant amino acids found in fish in the free and protein-bound forms [48]. Other amino acids found at abundant levels in the three fish also play important physiological and biochemical functions. The NEAA arginine, for example, is involved in protein synthesis, creatine formation, and various other biochemical processes within the body. The EAAs leucine, phenylalanine, and methionine play important roles in the synthesis of nitric oxide, a molecule that can expand blood vessels [48].

4. Conclusions

The present study provides insightful information about the fatty acid and amino acid compositions, biometric measurements, proximate analysis, and mineral composition of three local freshwater species of Indonesian Bagridae fish. The findings show their potential as a nutrient-dense, sustainable food source that can support efforts to increase food security. These fish species might be incorporated into the local diet to increase nutrient intake and alleviate nutritional inadequacies, which would ultimately improve Indonesia's overall food security status.

Data Availability

The data utilized in this research has not been previously released or published in any form. The datasets employed and/or analysed during the present study can be obtained by contacting the corresponding author.

Conflicts of Interest

The authors confirm that they do not have any conflicting interests.

Acknowledgements

The authors acknowledge and express their gratitude for the financial support received from the Ministry of Education, Culture, Research, and Technology of the Republic of Indonesia, specifically through grant no. 076/E5/PG. 02.00. PT/2022, which funded the

research. The authors affirm that there is no conflict of interest associated with the utilization of these research funds. Furthermore, the authors extend their appreciation to the students and fishers involved in the research conducted in the upper reaches of the Kampar Kanan River, Indonesia.

References

- [1] A.G. Hasselberg, I.Aakre, J. Scholtens *et al.*, “Fish for food and nutrition security in Ghana: Challenges and opportunities,” *Global Food Security*. vol 26: 100380, 2020 <https://doi.org/10.1016/j.gfs.2020.100380>
- [2] N.Tran, U.P, Rodriguez, C.Y.Chan., *et al.*, “Future scenarios of fish supply and demand for food and nutrition security in Bangladesh: An analysis with the AsiaFish model”, *Aquaculture*. Vol.568: 739288, 2023. <https://doi.org/10.1016/j.aquaculture.2023.739288>
- [3] C.Z. Levkoe, K. Lowitt, C. Nelson. “Fish as food: Exploring a food sovereignty approach to small-scale fisheries”. *Marine Policy*. vol 85: pp.65-70, 2017. <https://doi.org/10.1016/j.marpol.2017.08.018>
- [4] M. Ravichandran, N.S. Hettiarachchy, V. Ganesh *et al.*, “Enhancement of antimicrobial activities of naturally occurring phenolic compounds by nanoscale delivery against listeria monocytogenes, escherichia coli O157:H7 and salmonella typhimurium in broth and chicken meat system”. *Journal of Food Safety*. vol 31, no. 4, pp. 462-471,2011. <https://doi.org/10.1111/j.1745-4565.2011.00322.x>
- [5] N. Aryani, I. Suharman, Azrita, *et al.*, “Diversity and distribution of fish fauna of upstream and downstream areas at Koto Panjang Reservoir, Riau Province, Indonesia”. *F1000 Research*. vol 8, pp. 1435. 2020. <https://f1000research.com/articles/8-1435/v2>
- [6] Syafrialdi, Dahelmi, D.I. Roesma, *et al.*, “Length-weight relationships and condition factor of Two-Spot Catfish (*Mystus nigriceps* [Valenciennes, 1840]) Pisces, Bagridae], from Kampar Kanan River and Kampar Kiri River in Indonesia”. *Pakistan Journal of Biological Science*, vol. 23, no. 12, pp. 1636-1642, 2020. <https://scialert.net/fulltext/?doi=pjbs.2020.1636.1642>
- [7] N. Aryani, “Native species in Kampar Kanan River, Riau Province Indonesia. *International Journal of Fisheries and Aquatic Studies*”, vol.2, no.5, pp. 213-217. 2015. <https://www.fisheriesjournal.com/archives/2015/vol2issue5/PartD/2-5-63.pdf>
- [8] Azrita, H. Syandri, N. Aryani, *et al.*, “The utilization of new products formulated from water coconut, palm sap sugar, and fungus to increase nutritional feed quality, feed efficiency, growth, and carcass of gurami sago (*Osphronemus goramy* Lacepède, 1801) juvenile”. *F1000 Research*. vol. 10. :1121, pp. 1-16, 2021. <https://f1000research.com/articles/10-1121/v1>
- [9] C. Bene, R. Arthur, H. Norbury *et al.*, “Contribution of Fisheries and Aquaculture to Food Security and Poverty Reduction: Assessing the Current Evidence”. *World Development*, 79: 177–196. 2016. <http://dx.doi.org/10.1016/j.worlddev.2015.11.007>
- [10] N. Aryani, I. Suharman, S. Hasibuan, *et al.*, “Fatty acid composition on diet and carcasses, growth, body indices and profile serum of Asian redtail catfish (*Hemibagrus nemurus*) fed a diet containing different levels of EPA and DHA”. *F1000Research*. 11:1409.pp.1-23, 2023. <https://doi.org/10.12688/f1000research.126487.2>

- [11] E.N. Kingsley, O.O. Cyril, O.I. Patience, "Potential contribution of selected wild fish species to the minerals intake of Pregnant and Lactating Women, Children and Adults in Rural Riverine Communities of Edo State: Insights and Outcomes". *Measurement: Food*, 8, 100063 : 1-7, 2022. <https://doi.org/10.1016/j.meafoo.2022.100063>
- [12] A.A. Sausa, L. Nora, D.L.A. Lopes, *et al.*, Vegetable choline in feed for Nile tilapia (*Oreochromis niloticus*) raised in a biofloc technology system (BFT): Biofloc composition, chemical composition, and fatty acid profiles in meat". *Aquaculture*. vol. 545. 737174, 2021. <https://doi.org/10.1016/j.aquaculture.2021.737174>
- [13] A.M. Eltweri, A.L. Thomas, M. Metcalfe, *et al.*, Potential applications of fish oils rich in omega-3 polyunsaturated fatty acids in the management of gastrointestinal cancer. *Clinical Nutrition*. vol. 36, no. 1, pp. 65-78. 2017. <https://doi.org/10.1016/j.clnu.2016.01.007>
- [14] P.C. Haycock, M.C. Borges, K. Burrows, *et al.*, "The association between genetically elevated polyunsaturated fatty acids and risk of cancer". *eBiomedicine*. vol.91: 104510, 2023. <https://doi.org/10.1016/j.ebiom.2023.104510>
- [15] E.K. Aglago, I. Huybrechts, N. Murphy, *et al.*, "Consumption of Fish and Long-chain n-3 Polyunsaturated Fatty Acids Is Associated With Reduced Risk of Colorectal Cancer in a Large European Cohort". *Clinical Gastroenterology and Hepatology*. vol. 18, no. 3, pp. 654-666.e6. 2020. <https://doi.org/10.1016/j.cgh.2019.06.031>
- [16] H. Jiang, X. Shi, T.Y. Fan, *et al.*, "Dietary omega-3 polyunsaturated fatty acids and fish intake and risk of age-related macular degeneration" . *Clinical Nutrition*. vol.40, no. 12, pp: 5662-5673. 2021. [https://www.clinicalnutritionjournal.com/article/S0261-5614\(21\)00474-X/fulltext](https://www.clinicalnutritionjournal.com/article/S0261-5614(21)00474-X/fulltext)
- [17] M.A. Rincón-Cervera, NV. González-Barriga, R. Valenzuela, *et al.*, "Profile and distribution of fatty acids in edible parts of commonly consumed marine fishes in Chile", *Food Chemistry*, vol. 274, pp. 123-129, 2018. <https://doi.org/10.1016/j.foodchem.2018.08.113>
- [18] S. Sun, T. Ren, X. Li., *et al.*, "Polyunsaturated fatty acids synthesized by freshwater fish: A new insight to the roles of elovl2 and elovl5 in vivo", *Biochemical and Biophysical Research Communications*, vol. 532, pp. 414-419, 2020. <https://doi.org/10.1016/j.bbrc.2020.08.074>
- [19] M.B. Behyar, M. Hasanzadeh, F. Seidi, *et al.*, "Sensing of amino acids: Critical role of nanomaterials for the efficient biomedical analysis", *Microchemical Journal*, vol. 188, pp. 108452, 2023 <https://doi.org/10.1016/j.microc.2023.108452>
- [20] P. Shi, K. Liao, J. Xu, *et al.*, "Eicosapentaenoic acid mitigates palmitic acid-induced heat shock response, inflammation and repair processes in fish intestine", *Fish & Shellfish Immunology*, vol. 124, pp. 362-371, 2022. <https://doi.org/10.1016/j.fsi.2022.04.011>
- [21] J. Wen, L. Zeng, Y. Xu, *et al.*, Proximate composition, amino acid and fatty acid composition of fish maws, *Natural Product Research: Formerly*, vol. 30, no. 2, pp. 214-217, 2015. <https://doi.org/10.1080/14786419.2015.1040790>
- [22] F.O.S. Duarte, F.G. de Paula, C.S. Prado, *et al.*, "Better fatty acids profile in fillets of Nile Tilapia (*Oreochromis niloticus*) supplemented with fish oil", *Aquaculture*, Vol. 534, pp. 736241, 2021. <https://doi.org/10.1016/j.aquaculture.2020.736241>

- [23] K.L. Weaver, P. Ivester, J.A. Chilton, *et al.*, “The Content of Favorable and Unfavorable Polyunsaturated Fatty Acids Found in Commonly Eaten Fish”, *Journal of the American Dietetic Association*, vol. 108, No. 7, pp. 1178–1185, 2008. <https://doi.org/10.1016/j.jada.2008.04.023>
- [24] AOAC, “Official Methods of Analysis. 15th Edn., Association of Official Analytical Chemists (AOAC)”, Washington, DC., USA.1990. Reference Source
- [25] S.A. Cohen, “Amino acid analysis using pre-column derivatization with 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate”, *Protein Sequencing Protocols*, vol. 211, pp. 143-54, 2003. <https://doi.org/10.1385/1-59259-047-0:039>
- [26] M.A. Rajion, J.G. McLean, R.N. Cahill, “Essential fatty acid metabolism in the fetal and neonatal lamb”, *Australian Journal of Biological Sciences*, vol. 38, no. 1, pp. 33-40, 1985. [Essential fatty acids in the fetal and newborn lamb - PubMed \(nih.gov\)](https://pubmed.ncbi.nlm.nih.gov/1985/10.1016/0005-6273(85)91846-M)
- [27] T.L. Ulbricht, D.A. Southgate, “Coronary heart disease: Seven dietary factors”, *The Lancet*, Vol 338, no. 8773, pp. 985–992, 1991. [https://doi.org/10.1016/0140-6736\(91\)91846-M](https://doi.org/10.1016/0140-6736(91)91846-M)
- [28] D.B. Duncan, “Multiple ranges and multiple F tests”, *Biometrics*, vol. 11, pp. 1–42, 1955. <https://doi.org/10.2307/3001478>
- [29] R. Froese, “Cube law, condition factor and weight–length relationships: History, meta-analysis and recommendations”. *Journal of Applied Ichthyology*, vol. 22, no. 4, pp. 241-253, 2006. <https://doi.org/10.1111/j.1439-0426.2006.00805.x>
- [30] O.O. Famofofo, & W.O. Abdul, “Biometry, condition factors and length-weight relationships of sixteen fish species in Iwopin fresh-water ecotype of Lekki Lagoon, Ogun State, Southwest Nigeria”, *Heliyon*, vol. 6, no. 1, 2020. <https://doi.org/10.1016/j.heliyon.2019.e02957>
- [31] T.B. Bagenal, F.W. Tesch, “Age and Growth. In: Bagenal, T., Ed., Methods for Assessment of Fish Production in Fresh Waters”, *Blackwell Science Publications*, pp. 101-136, 1978.
- [32] E.O. Ahmed, M.E. Ali, A.A. Aziz, “Length-weight relationships and Condition factors of six fish species in Atbara River and Khashm el-girba Reservoir, Sudan”, *International Journal of Agriculture Sciences*, vol. 3, no.1, pp. 65–70, 2011. <http://dx.doi.org/10.9735/0975-3710.3.1.65-70>
- [33] N.C. Ujjania, M.P.S. Kohli, L.L. Sharma, “Length-weight relationship and condition factors of Indian major carps (C. catla, L. rohita and C. mrigala) in Mahi Bajaj Sagar, India”, *Research Journal of Biology*, vol. 2, no.1, pp. 30-36, 2012. <https://doi.org/10.21077/ijf.2017.64.special-issue.76263-27>
- [34] J.P. Bonjour, “Calcium and phosphate: A duet of ions playing for bone health”, *Journal of the American College of Nutrition*, vol. 30, no. 5, pp. 438S-48S, 2011. <https://doi.org/10.1080/07315724.2011.10719988>
- [35] B.P. Mohanty, T.V. Sankar, S. Ganguly, *et al.*, “Micronutrient composition of 35 food fishes from India and their significance in human nutrition”, *Biological Trace Element Research*, vol. 174, pp. 448-458, 2016. <https://doi.org/10.1007/s12011-016-0714-3>
- [36] F. Jabeen, A.A. Chudhry, “Chemical composition and fatty acid profiles of three freshwater species”, *Food Chemistry*, vol. 125, pp. 991-996, 2011. <https://doi.org/10.1016/j.foodchem.2010.09.103>

- [37] A.A. Nurnadia, A. Azrina, I. Amin, "Proximate composition and energetic value of selected marine fish and shellfish from the west coast of peninsular Malaysia". *International Food Research Journal*. Vol 18, pp. 137-148, 2011. <https://core.ac.uk/download/pdf/153807129.pdf>
- [38] S.C. Dyal, L. Balas, N.G. Bazan, *et al.*, "Polyunsaturated fatty acids and fatty acid-derived lipid mediators: Recent advances in the understanding of their biosynthesis, structures and functions", *Progress in Lipid Research*. vol. 86, pp. 101165, 2022. <https://doi.org/10.1016/j.plipres.2022.101165>
- [39] N. Kim, U.D.Sohn, V. Mangannan, *et al.*, "Leukotrienes in Acetylcholine-Induced Contraction of Esophageal Circular Smooth Muscle in Experimental", *Gastroenterology*, vol. 112, no.5, pp. 1548–1558, 1997. [https://doi.org/10.1016/S0016-5085\(97\)70036-2](https://doi.org/10.1016/S0016-5085(97)70036-2)
- [40] D. Swanson, R. Block, S.A.Mousa, "Omega-3 Fatty Acids EPA and DHA: Health Benefits Throughout Life". *Advances in Nutrition*, vol. 3, no.1, pp. 1-7,2012. <https://doi.org/10.3945/an.111.000893>
- [41] R.P. Mason, S.C.R. Sherratt, , L.H. Eckel, "Omega-3-fatty acids: Do they prevent cardiovascular disease?", *Best Practice & Research Clinical Endocrinology & Metabolism*, vol.37, no 3, 101681, 2023. <https://doi.org/10.1016/j.beem.2022.101681>.
- [42] J. M. Seddon, J. Cote, N. Davis, *et al.*, "Progression of age-related macular degeneration. Progression of age-related macular degeneration". *Arch Ophthalmol*. Vol. 121, no. 6, pp. 785-792, 2003. <https://doi.org/10.1001/archophth.121.6.785>
- [43] FAO/WHO: Fat and Oils in Human Nutrition: Report of a Joint Expert Consultation. FAO/WHO, Rome, Italy.
- [44] S.M. Pawar, S.R. Sonaware, "Fish muscle protein highest source of energy". *International Journal of Biodiversity and Conservation*. Vol. 5, no. 7, pp. 433-435, 2013. <https://doi.org/10.5897/IJBC12.043>
- [45] D. Tomé, C. Bos, "Lysine requirement through the human life cycle", *The Journal of Nutrition*, vol. 137, no.6, pp. 1642S–1645S, 2007. <https://doi.org/10.1093/jn/137.6.1642S>
- [46] M. Watford, "Glutamine and glutamate: Nonessential or essential amino acids?", *Animal Nutrition*, vol. 1, no. 3, pp. 119-122, 2015. <https://doi.org/10.1016/j.aninu.2015.08.008>
- [47] X. Li, S. Zheng, G. Wu, "Nutrition and metabolism of glutamate and glutamine in fish", *Amino Acids*. vol.52 , no.5, pp. 671-691, 2020. <https://doi.org/10.1007/s00726-020-02851-2>
- [48] M. Li, Y. Wu, L. Ye, " The Role of Amino Acids in Endothelial Biology and Function", *Cells*, vol.11, no.8, pp. 1372, 2022. <https://doi.org/10.3390/cells11081372>



Webmail
Univ. Bung Hatta

hafrizal syandri <syandri_1960@bunghatta.ac.id>

6639837: Revision requested

3 messages

International Journal of Food Science <ijfs@hindawi.com>

Mon, Nov 13, 2023 at 3:49 AM

Reply-To: International Journal of Food Science <deepanaa.rajadurai@hindawi.com>

To: Hafrijal Syandri <syandri_1960@bunghatta.ac.id>



Dear Dr. Hafrijal Syandri,

In order for your submission "Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security " to "International Journal of Food Science" to proceed further in the review process, you will need to revise your manuscript.

Reason & Details:

“

*Dear Authors,
I have received another report from the reviewer who suggested minor revisions before the acceptance of the manuscript.*

When you have finished revising, follow the link below to submit your revision:

MANUSCRIPT DETAILS

Kind regards,
Deepanaa Rajadurai
International Journal of Food Science

Reviewer Comments:

“

Reviewer 1 Comments to the Author

The idea to study the nutritional value of Local Freshwater Species is excellent and interesting. and deserves to be published in the International Journal of Food Science. There are some problems inredaction style and should be checked by an English speaker.

Abstract

- The objective should be described;

Keywords

- Abbreviations must be written in alphabetical order.

MM

- Animal material not Materials.

- TI : Stearic acid is a fatty acid that is neutral concerning cardiovascular disease. Normally you should not take it as a proatherogenic fatty acid in your calculations.

- Figure 1 : invisible, need to improve the resolution

This email was sent to syandri_1960@bunghatta.ac.id. You have received this email in regards to the account creation, submission, or peer review process of a paper submitted to a journal published by Hindawi Limited.

Hindawi Limited, 3rd Floor, Adam House, [1 Fitzroy Square, London, W1T 5HF, United Kingdom](#).

Hindawi respects your right to privacy. Please see our [privacy policy](#) for information on how we store, process, and safeguard your data.

[Unsubscribe](#)

hafrizal syandri <syandri_1960@bunghatta.ac.id>
To: International Journal of Food Science <deepanaa.rajadurai@hindawi.com>

Wed, Nov 15, 2023 at 4:04 PM

Dear
Deepanaa Rajadurai
International Journal of Food Science

Thank you for your email on 13 November 2023, we have revised manuscript No. 6639837 based on reviewer comments.

Best Regards
Hafrijal Syandri
[Quoted text hidden]

Phenom Emails <phenom.emails@hindawi.com>
Reply-To: Phenom Emails <phenom.emails@hindawi.com>
To: syandri_1960@bunghatta.ac.id

Wed, Nov 15, 2023 at 6:44 PM

Dear Dr. Syandri,

Thank you so much for your revised manuscript.

Please do not hesitate to get back in touch if you have any further questions.

Best Regards,

Deepanaa

Deepanaa Rajadurai
Editorial Assistant



Hindawi

[Hindawi.com](https://www.hindawi.com) | [Twitter](#) | [Facebook](#) | [LinkedIn](#) | [YouTube](#)

[Quoted text hidden]
, **hafrizal syandri** <syandri_1960@bunghatta.ac.id> wrote:
[Quoted text hidden]

[Quoted text hidden]

The contents of this email and any attachments are confidential and intended only for the person or entity to whom it is addressed. If you are not the intended recipient, any use, review, distribution, reproduction or any action taken in reliance upon this message is strictly prohibited. If you received this message in error, please immediately notify the sender and permanently delete all copies of the email and any attachments.

[Click here for translations of this disclaimer.](#)

Research Article

Analysis of Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security

Azrita Azrita¹, Hafrijal Syandri^{1*}, Hazlina Ahamad Zakeri², Harfiandri Damanhuri¹, Netti Aryani³

¹Faculty of Fisheries and Marine Science, Universitas Bung Hatta, 25131, Padang-West Sumatra Province, Indonesia

²Faculty of Science and Marine Environment, Universiti Malaysia Terengganu, Malaysia.

³Faculty of Fisheries and Marine Science, Universitas Riau, 28293, Pekanbaru-Riau Province, Indonesia

*Corresponding author: syandri_1960@bunghatta.ac.id

Abstract

Fish have become an irreplaceable dietary source of animal protein, especially among households with low socioeconomic status in rural and urban areas of Indonesia. This study aimed to analyse the proximate composition, minerals, fatty acids, and amino acids of three local Bagridae fish species in Indonesia. The standard AOAC method was employed to examine the proximate composition of the carcass, and the analysis of amino acids and fatty acids was conducted through HPLC and GC-MS, respectively. The mineral content was determined using AAS. The nutrient composition results of *Hemibagrus nemurus*, *Hemibagrus wyckii*, and *Mystus nigriceps* revealed that the protein content was 24.26%, 22.57%, and 21.39% (%dry weight), whereas the total lipid content was 6.64%, 7.47% and 7.75%, respectively. Regarding mineral contents, the calcium levels ranged from 1.49–1.66 mg/g, iron levels from 28.35–40.36 µg/g, and zinc levels from 24.03–54.46 µg/g. Among the fatty acids, palmitic acid was the most abundant in all three species, accounting for 25.59–30.70% of the total fatty acids. Additionally, significant amounts of C18:1 (oleic acid), C18:0 (stearic acid), and C20:4 (arachidonic acid) were also detected as primary fatty acids. The calculated atherogenic index values in the three species of Bagridae fish ranged from 0.73–0.99, while the thrombogenic index values varied between 0.54–0.75. The predominant amino acids found in the three species of Bagridae fish were glutamic acid with their concentrations ranging from 9.10–24.34%. These results indicate that consuming the meat of these three freshwater Bagridae fish species caught in the wild does not pose any health risks to consumers. They can be considered a safe and suitable food source with good nutritional quality.

Keywords: Amino acids, Fatty acids, Indonesia, Kampar Kanan River, Minerals

Commented [S1]: The abstract has been added with the study objectives and revised with yellow highlighting

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

1. Introduction

Fish serve as a high-quality protein source that is essential for satisfying dietary protein requirements, especially in areas where other protein sources are few or difficult to obtain [1,2]. Their accessibility can aid in ensuring a steady supply of food, particularly for communities whose main source of income is fishing or aquaculture [3]. Fish can contribute to a more varied selection of protein sources in communities, resulting in a diet that is more nutritious and balanced. Dietary fish can treat nutrient deficits and enhance general nutritional health. Fish are generally considered safe and offer a lower risk of contamination with harmful substances compared to some land-based animal products. A reliable indicator of a fish's quality, nutritional value, physiological status, and environment is its chemical makeup, which is found in fish flesh [4].

Hemibagrus nemurus, locally known as "baung", *Hemibagrus wyckii*, known as "geso" and *Mystus nigriceps*, known as "ingir-ingir," are three species of catfish found in the Kampar Kanan River, Indonesia [5,6]. Due to their high market value, these species have been identified as significant economic resources for rural communities [7]. Therefore, the analysis of the nutritional quality of fish meat in the study area is very important to determine whether it is beneficial to human health.

Knowing the composition of fatty acids and amino acids in fish meat is an essential factor that should not be ignored [8]. This information provides valuable insight into the nutritional quality of fish and the values of more nutritious and healthy foods [9]. In addition, a better understanding of the composition of fatty acids and amino acids can assist in selecting a suitable feed [10], developing a balanced diet [11], and maintaining optimal fish meat nutrition [12].

Omega-3 polyunsaturated fatty acids (PUFAs), particularly docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA) derived from fish oil, exhibit anti-inflammatory and anti-gastrointestinal cancer properties, making them crucial as immune-boosting nutrients [13,14]. Studies have demonstrated that ω -3 and ω -6 PUFAs exhibit beneficial effects in the management of cardiovascular diseases and cancers [15,16]. The fatty acid composition of PUFAs can differ among different types of fish, including both freshwater and marine species

[17, 18]. Amino acids (AAs), such as cysteine, arginine, tyrosine, glycine, proline, and serine, play a vital role in illness and stress situations and in the prevention of inflammation and repair processes in fish intestines [19, 20].

Furthermore, knowledge of the atherogenicity index (AI) and thrombogenicity index (TI) of fatty acids in the three local freshwater fish species of Bagridae can contribute to improving dietary guidelines. The AI and TI can be used to develop recommendations aimed at improving heart health [21,22]. These recommendations may involve selecting fish with favourable fatty acid profiles, allowing individuals to choose the most suitable fish species based on their specific needs [17, 23]. Therefore, this study aims to examine the nutritional composition profile, mineral content, amino acids, and fatty acids, and AI and TI of three freshwater fish species, namely, *H. nemurus*, *H. wyckii*, and *M. nigriceps*. These three fish species are generally consumed by people who live around the Kampar Kanan River in Kampar Regency, Riau Province, Indonesia.

2. Material and methods

2.1. Animal material

In this study, a total of 27 Bagridae fish belonging to three distinct species (*H. nemurus*, *H. wyckii*, and *M. nigriceps*) were examined. The specimens were obtained from local fishers operating in the upper stretches of the Kampar Kanan River in Kampar Regency, Riau Province (Figure 1). Sampling took place between June and August 2022 and was verified by the Fisheries Department, Faculty of Fisheries and Marine Science, Universitas Bung Hatta, Indonesia. The samples were placed in plastic bags and transported to the laboratory in an insulated icebox. The samples were then grouped based on their respective sampling locations.

2.2. Biometry measurement

Upon reaching the laboratory within 10 hours, the fish samples were subjected to gutting and washing. Each specimen was weighed (TW) and measured to determine the standard length (SL) as well as the depth or maximum height (H). TW was measured using balance scale (OHAUS Model CT 6000 USA) with a precision of 0.01 g. The length measurement was taken from the tip of the mouth to the end of the upper lobe of the caudal fin, representing the total body length, using a metre ruler with an accuracy of 1 mm. The height measurement involved a vertical assessment of the body's maximum height measured using a

Commented [S2]: The author has been revised

Formatted: Highlight

Digital Sekhmet Sigma Vernier calliper with an accuracy of 1 mm. The condition factor (CF) was calculated using the formula $CF = (TW/SL^3) \times 100$.

2.3. Proximate, amino acid, and mineral content analysis

The proximate carcass composition was assessed following the AOAC standard methods [24]. The samples were dried to a constant weight at 105 °C. The crude protein content was analysed using the standard Kjeldahl method, which involves multiplying the nitrogen content by a factor of 6.25. The Soxhlet method with ether extraction was employed to analyse crude lipids. The ash content was obtained by incinerating the samples at 550 °C for 16 h. The amino acid composition was determined using a high-performance liquid chromatography (HPLC) system, which consisted of a Waters® 1525 binary HPLC pump, Waters® 717 autosamplers, and Waters® 2475 multi- λ fluorescence detector (with excitation at 250 nm and emission at 395 nm). The samples were hydrolysed in triplicate with 6 N hydrochloric acid for 24 h at 11 °C [25].

For mineral data composition (Na, Mg, Ca, K, P, Fe, Cu, Mn, and Zn), the ashed feed sample and carcass were dissolved in 1 ml of hydrochloric acid (35% v/v Suprapur®, Merck) and then filtered with cellulose filter paper (Watchman No 1, International Ltd; Maidstone, UK) and diluted to an appropriate concentration for each elemental mineral. P levels were analysed with a Perkin-Elmer AA spectrophotometer mod 3110 (Norwalk, CT, USA).

2.4. Fatty acid analysis

The fish meat was examined utilizing the gas chromatography–mass spectrometry (GC–MS) method. The method of Folch et al. (1957) modified by Rajion et al. [26] was employed to perform the total fat extraction. This involved using a solvent system consisting of chloroform and methanol at a 2:1 (v/v) ratio. The process of transmethylation was conducted using a solution of 14% methanolic boron trifluoride. The fatty acid composition of the meat was then analysed at the SIG Laboratory, Accredited Testing Laboratory-LP-184-IDN.

The nutritional quality of lipids (AI and TI) was calculated using the following equations [27]:

$$AI = \frac{[(C12:0 + 4 \times C14:0 + C16:0)]}{\sum MUFA + \sum n-6 + \sum n-3}$$

$$TI = \frac{[C14:0 + C16:0 + C18:0]}{[(0.5 \times \sum MUFA) + (0.5 \times \sum n-6) + (3 \times \sum n-3) + (\sum n-3 / \sum n-6)]}$$

where:

AI = Atherogenic index

TI = Thrombogenic index

C12:0 = lauric acid

C14:0 = myristic acid

C16:0 = palmitic acid

C18:0 = stearic acid

\sum MUFA = sum of the concentrations of all monosaturated fatty acids

\sum n-6 = sum of the concentrations of n-6 polyunsaturated fatty acids

\sum n-3 = sum of the concentrations of n-3 polyunsaturated fatty acids

2.5. Data analysis

Data analysis was performed using the SPSS 16.0 software package (SPSS; Chicago, IL). The homogeneity of variances was assessed using Levene's test. One-way ANOVA was carried out to determine the parameters of proximate and amino acid composition, including the composition of fatty acids and nutritional quality of lipids (AI and TI) for each species, followed by the post hoc Duncan's multiple range test [28]. The results are reported as the mean values \pm standard errors for each parameter.

Commented [S3]: We appreciate the reviewer's contribution regarding the perspective that C18:0 should not be considered a pro-atherogenic fatty acid. However, to ensure alignment with the Thrombogenic Index analysis, we would like to request further clarification as to whether the authors expected to remove C18:0 from our data

Formatted: Highlight

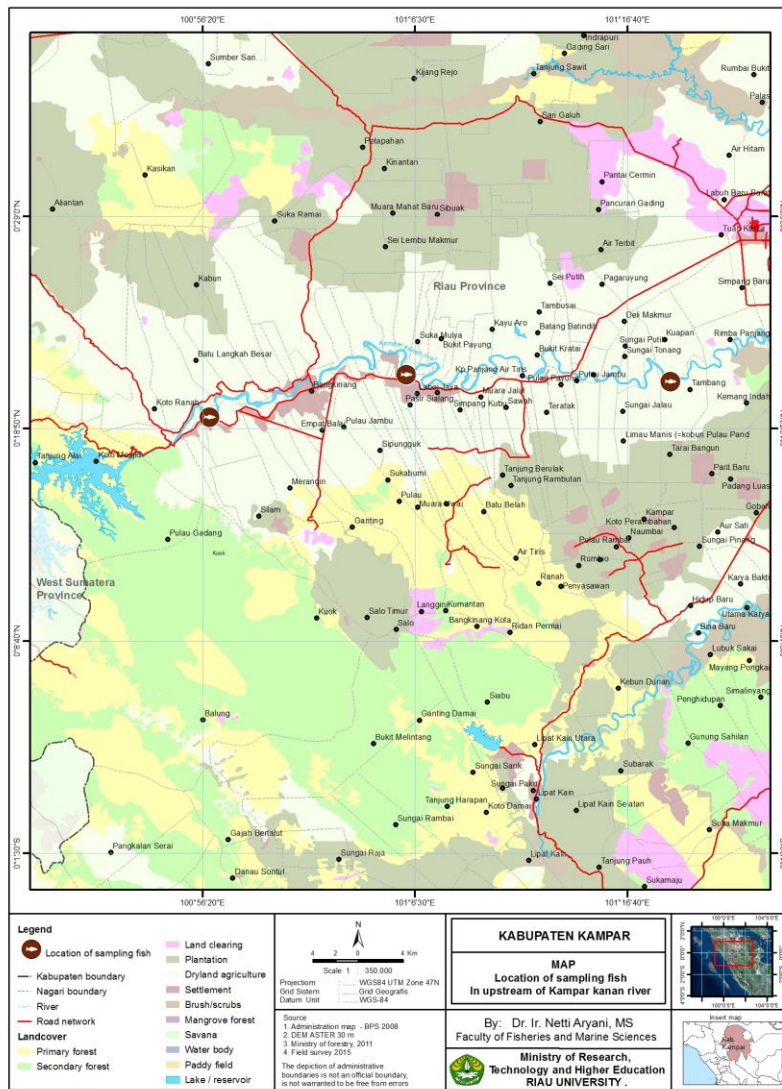


Figure 1. Map of Kampar Regency, Riau Province, and sampling locations of the three species of Bagridae fish.

Formatted: Highlight

Commented [S4]: Figure 1 has been revised

3. Results

Table 1 presents the average wet weight, standard length, height, and the results of meat nutritional composition analysis for three indigenous Indonesian freshwater Bagridae species. The moisture content (% wet weight) ranged from 82.40– 85.39%. Among the species examined, *M. nigriceps* exhibited the lowest protein content at 21.39%, while *H. nemurus* displayed the highest value at 24.26%. However, *M. nigriceps* had a higher mineral content, including of iron and zinc.

Table 2 summarizes the fatty acid composition (%total fatty acid) for the three Bagridae fish species. C16:0 was the predominant fatty acid in the three species of Bagridae fish, with percentages ranging from 25.09–30.71%. The additional prominent fatty acids found were C18:1, C18:0, and C22:6. *H. nemurus* exhibited a ω -6/ ω -3 ratio below 1 (0.82), while *H. wyckii* and *M. nigriceps* had ratios of 1.11 and 1.02, respectively. The obtained PUFA/saturated fatty acid (SFA) (P/S) ratio varied between 0.61 and 0.76. The AI values varied between 0.76 and 0.99 and the TI between 0.54 and 0.75 (**Table 3**).

Table 4 presents the amino acid composition (% of total protein) of the three Bagridae species. Glutamic acid was the dominant amino acid, ranging from 19.29–24.34%, followed by aspartic acid, which ranged from 9.21–11.27%. The lysine content was consistent, ranging from 9.67–9.87%. In *H. nemurus*, the levels of various amino acids ranged from 0.86–24.35%. Similarly, in *H. wyckii*, the amino acid levels ranged from 0.76–21.58%, while in *M. nigriceps*, the levels ranged from 0.74–19.29%.

Table 1. Results of biometric, proximate and mineral composition of three species of Bagridae fish

	Species		
	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
<i>Biometric measurements</i>			
Total weight (g)	389.99 ± 24.96 ^a	1,390.33 ± 168.29 ^b	17.57 ± 1.53 ^c
Standard length (cm)	28.16 ± 0.53 ^a	44.14 ± 1.98 ^b	10.29 ± 0.15 ^c
Height (cm)	8.44 ± 0.16 ^a	8.82 ± 0.39 ^b	2.57 ± 0.03 ^c
Condition factor	1.63 ± 0.08	1.58 ± 0.07	1.56 ± 0.08
<i>Proximate composition</i>			
Crude protein (% DW)	24.26 ± 0.87 ^a	22.57 ± 0.37 ^b	21.39 ± 0.15 ^c
Crude fat (% DW)	6.64 ± 0.03 ^a	7.47 ± 0.02 ^b	7.75 ± 0.40 ^c
Crude ash (% DW)	1.94 ± 0.02 ^a	2.30 ± 0.09 ^b	1.59 ± 0.02 ^c
Moisture (% WW)	82.40 ± 2.51	85.39 ± 2.36	83.75 ± 2.21

<i>Mineral composition</i>			
Macrominerals (mg/g)			
Sodium	0.99 ± 0.00 ^a	1.61 ± 0.01 ^b	1.61 ± 0.02 ^c
Magnesium	1.12 ± 0.01 ^a	0.57 ± 0.02 ^b	1.15 ± 0.01 ^c
Calcium	1.66 ± 0.00 ^a	1.55 ± 0.02 ^b	1.49 ± 0.04 ^c
Potassium	0.71 ± 0.00 ^a	0.55 ± 0.02 ^b	0.43 ± 0.01 ^c
Phosphorous	7.03 ± 0.03 ^a	2.74 ± 0.02 ^b	6.45 ± 0.07 ^c
Microminerals (µg/g)			
Iron	28.30 ± 0.11 ^a	28.73 ± 0.08 ^b	40.36 ± 0.55 ^c
Copper	8.93 ± 0.03 ^a	7.46 ± 0.09 ^b	6.21 ± 0.32 ^c
Manganese	1.64 ± 0.02 ^a	2.64 ± 0.02 ^b	2.84 ± 0.02 ^c
Zinc	24.03 ± 0.45 ^a	24.61 ± 0.19 ^b	54.46 ± 0.17 ^c

Values are mean % ± SE; *Values in the same row followed by different letters are significantly different (p<0.05).

Table 2. Fatty acid composition of three species of Bagridae fish

Fatty acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
C12:0 (lauric acid)	1.97 ± 0.05 ^a	3.94 ± 0.01 ^b	4.27 ± 0.01 ^c
C14:0 (Meristic acid)	2.38 ± 0.03 ^a	2.72 ± 0.08 ^b	2.89 ± 0.01 ^c
C16:0 (Palmitic acid)	27.23 ± 0.06 ^a	30.70 ± 0.06 ^b	25.59 ± 0.01 ^c
C18:0 (Stearic acid)	16.17 ± 0.02 ^a	13.54 ± 0.02 ^b	10.64 ± 0.01 ^c
C20:0 (Arachidic acid)	0.23 ± 0.01 ^a	0.14 ± 0.01 ^b	0.37 ± 0.01 ^c
C16:1 (Palmitoleic acid)	1.78 ± 0.00 ^a	1.27 ± 0.04 ^b	4.93 ± 0.01 ^c
C18:1 (Oleic acid)	16.86 ± 0.01 ^a	16.18 ± 0.01 ^b	16.97 ± 0.01 ^c
C18:2 (Linoleic acid)	3.24 ± 0.02 ^a	3.84 ± 0.33 ^b	4.55 ± 0.001 ^c
C18:3 (Linolenic acid)	1.12 ± 0.01 ^a	0.82 ± 0.01 ^b	1.57 ± 0.000 ^c
C20:4 (Arachidonic acid)	12.13 ± 0.01 ^a	12.49 ± 0.09 ^b	12.41 ± 0.02 ^c
C20:5 (Eicosapentaenoic acid; EPA)	2.06 ± 0.08 ^a	2.34 ± 0.04 ^b	1.35 ± 0.01 ^c
C22:6 (Docosahexaenoic acid; DHA)	15.65 ± 0.01 ^a	11.56 ± 0.04 ^b	13.64 ± 0.02 ^c

Values are mean % ± SE; *Values in the same row with a different superscript are significantly different (p<0.05).

Table 3. Fatty acid ω-6/ω-3 and polyunsaturated/saturated fatty acid ratio of three species of Bagridae fish

	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
ω-6/ω-3 ratio	0.82 ± 0.00 ^a	1.11 ± 0.02 ^b	1.02 ± 0.01 ^c

P/S ratio	0.69 ± 0.01 ^a	0.61 ± 0.06 ^b	0.76 ± 0.00 ^c
AI	0.87 ± 0.01 ^a	0.99 ± 0.06 ^b	0.73 ± 0.00 ^c
TI	0.63 ± 0.01 ^a	0.75 ± 0.03 ^b	0.54 ± 0.00 ^c

P/S, polyunsaturated/saturated fatty acid; AI, atherogenic index; TI, thrombogenic index
 Values are mean % ± SE; *Values in the same row followed by different letters are significantly different (p<0.05).

Table 4. Amino acid composition of three species of Bagridae fish

Amino acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
Aspartic acid	10.39 ± 0.01 ^a	9.61 ± 0.04 ^b	11.28 ± 0.02 ^c
Glutamic acid	24.34 ± 0.00 ^a	21.58 ± 0.41 ^b	19.29 ± 0.03 ^c
Serine	3.74 ± 0.00 ^a	5.40 ± 0.03 ^b	7.50 ± 0.06 ^c
Glycine	4.02 ± 0.00 ^a	5.77 ± 0.04 ^b	4.69 ± 0.02 ^c
Histidine	2.39 ± 0.00 ^a	2.90 ± 0.03 ^b	2.21 ± 0.02 ^c
Arginine	6.17 ± 0.01 ^a	8.27 ± 0.03 ^b	6.22 ± 0.03 ^c
Threonine	4.49 ± 0.01 ^a	4.19 ± 0.03 ^b	4.39 ± 0.04 ^c
Alanine	7.03 ± 0.00 ^a	6.04 ± 0.02 ^b	7.89 ± 0.01 ^c
Proline	2.40 ± 0.00 ^a	2.71 ± 0.01 ^b	2.21 ± 0.02 ^c
Tyrosine	3.91 ± 0.01 ^a	3.29 ± 0.06 ^b	3.82 ± 0.04 ^c
Valine	3.73 ± 0.00 ^a	4.02 ± 0.01 ^b	3.85 ± 0.04 ^c
Methionine	3.42 ± 0.00 ^a	1.96 ± 0.02 ^b	1.27 ± 0.03 ^c
Cystine	0.87 ± 0.00 ^a	0.76 ± 0.02 ^b	0.75 ± 0.03 ^c
Isoleucine	3.82 ± 0.001 ^a	3.96 ± 0.003 ^b	3.66 ± 0.002 ^c
Leucine	6.15 ± 0.000 ^a	6.69 ± 0.002 ^b	6.28 ± 0.004 ^c
Phenylalanine	5.13 ± 0.000 ^a	6.03 ± 0.005 ^b	5.18 ± 0.027 ^c
Lysine	9.61 ± 0.001 ^a	9.82 ± 0.026 ^b	9.87 ± 0.003 ^c

Values are mean % ± SE; *Values in the same row followed by different letters are significantly different (p<0.05).

4. Discussion

The results of this study provide important new information about the nutritional makeup of the studied species of Bagridae fish. The biometric measures of the fish yielded important information about their physical traits and health. Researchers are able to determine the average weight of fish at various lengths by examining the link between length and weight for a particular species. This knowledge is essential for understanding the trends in fish population growth, identifying potential shifts in growth rates over time, and assessing the general health and welfare of fish [29]. A variety of factors, including sex, age, maturity level, size, stomach fullness level, sampling strategies, sample sizes, and environmental circumstances, have an impact on fish health and the parameters that determine length–weight connections in fish [30]. However, none of these factors were considered in our study. In general, according to the theory put forth by [Bagenal and Tesch \[31\]](#), heavier fish of a given length are in better physiological condition. The condition factor is also used in fisheries science to compare the health, fatness, and condition of fish [32]. A condition factor of > 1 is favourable, indicating a good feed nutrition level and suitable living habitat [33]. Thus, based on the findings in this study, the Kampar Kanan River provides a suitable habitat for the fish, ensuring their accessibility and availability for community consumption.

The fish samples exhibited good nutritional profiles, with high amounts of proteins and lipids as well as moisture content contributing to their overall nutritional value, according to the results of proximate analysis and mineral composition (see Table 1). These results imply that the fish species studied provide a possible source of nutrients to solve issues with food security. The study of the mineral composition revealed the existence of critical minerals such as phosphorus (P), calcium (Ca), magnesium (Mg), iron (Fe), and zinc (Zn) in the fish samples in addition to proteins and lipids. All three Bagridae fish significantly differed from each other in terms of their crude ash content and mineral profile (see Table 1). However, in general, P and Ca are the two highest minerals found in all fish. Both of these macro minerals are good for human bone and tooth health, as key components of hydroxyapatite, the mineral matrix that provides strength and structure to bones and teeth [34].

In the current study, the meat of three species of Bagridae fish from the Kampar Kanan River in Indonesia was found to contain an abundance of microminerals such as Fe and Zn. *M. nigriceps* has been reported to exhibit higher mineral content, including of Fe and Zn. This species is categorized as a small indigenous fish (SIF) with a maximum standard length

of approximately 10.29 cm. According to [35], SIFs are generally known to be rich in Ca, Fe, and Mn.

In this study, all three Bagridae fish had a total fat content of approximately 7–8% by weight, with a higher water content ranging from 82.40–85.39% (Table 1). These values are similar to those found in wild and cultured sea bass *Dicentrarchus labrax* [36]. Additionally, the fish species can be grouped into different categories, including high-fat fish (>8%), medium-fat fish (4–8%), low-fat fish (2–4%), and lean fish (<2%) [37]. According to the classification established by [37], the three indigenous Bagridae fish species are characterized as having a moderate fat content level, which is reflected by the high levels of DHA and EPA in the meat of *H. nemurus*, *H. wyckii*, and *M. nigriceps* (Table 2). Therefore, we recommend that consuming the meat of these three fish species can indirectly boost food security in rural and urban communities.

All three Bagridae fish species analysed in the study contained arachidonic acid (C20:4), at 12.13% for *H. nemurus*, 12.49% for *H. wyckii*, and 12.41% for *M. nigriceps*. Arachidonic acid serves as a precursor for synthesizing prostaglandins and leukotrienes [38]. Leukotrienes are crucial in allergic responses, inducing smooth muscle contraction and increasing vascular permeability [39]. Additionally, all three species also contained EPA ranging from 1.35–2.34% and DHA ranging from 11.56–15.65%. The increased consumption of omega-3 fatty acids, such as EPA and DHA, has been linked to a decreased risk of pain development in older individuals and to the prevention of Alzheimer's disease [40,41].

As determined by the current study, the ω -6/ ω -3 ratios of the three Bagridae fish species ranged from 1:0.8 to 1:0.97. It was found that all three fish species had ω -6/ ω -3 ratios within the recommended range for a typical Indonesian diet. In contrast, a typical Western diet is characterized by a high ω -6/ ω -3 ratio of 10:1 to 20:1, while a Mediterranean diet has a ratio of 4:1 to 5:1 [42]. Additionally, the P/S ratio analysis showed that the three Bagridae fish species are sources of PUFAs that meet the requirements for food safety.

The Food and Agriculture Organization (FAO) and the World Health Organization (WHO) have recommended AI and TI values ranging from 0.4 to 0.5 for promoting human health [43]. In the current study, the observed AI values in the three freshwater species of Bagridae ranged from 0.73 to 0.99, while the TI values varied between 0.54 and 0.75. This finding is associated with a notable disparity in the values of SFAs among the three Bagridae

fish species. Based on these findings, we confirm that consuming the meat of the three freshwater species of Bagridae fish caught in the wild does not endanger consumer health and can serve as a suitable source of food safety and nutritional quality.

Fish with a high crude protein content, as shown by the three studied Bagridae fish (Table 1), are regarded as being nutritionally useful for human health. The protein and amino acid contents can directly indicate the nutritional quality of meat [44]. As shown in Table 4, *H. wyckii* had a higher content of essential amino acids (EAAs) than *H. nemurus* and *M. nigriceps*. Among the EAAs, lysine (Lys) was found to be present in the highest amount in all three species. Lys is necessary for tissue growth and repair, reduced stress-induced anxiety, and the conversion of fatty acids into energy [45-47].

More than 60% of the amino acid composition in all three Brigade fish studied comes from nonessential amino acids (NEAAs), particularly glutamic acid (Table 4). Glutamic acid and its amine, glutamine, are highly abundant amino acids found in fish in the free and protein-bound forms [48]. Other amino acids found at abundant levels in the three fish also play important physiological and biochemical functions. The NEAA arginine, for example, is involved in protein synthesis, creatine formation, and various other biochemical processes within the body. The EAAs leucine, phenylalanine, and methionine play important roles in the synthesis of nitric oxide, a molecule that can expand blood vessels [48].

4. Conclusions

The present study provides insightful information about the fatty acid and amino acid compositions, biometric measurements, proximate analysis, and mineral composition of three local freshwater species of Indonesian Bagridae fish. The findings show their potential as a nutrient-dense, sustainable food source that can support efforts to increase food security. These fish species might be incorporated into the local diet to increase nutrient intake and alleviate nutritional inadequacies, which would ultimately improve Indonesia's overall food security status.

Data Availability

The data utilized in this research has not been previously released or published in any form. The datasets employed and/or analysed during the present study can be obtained by contacting the corresponding author.

Conflicts of Interest

The authors confirm that they do not have any conflicting interests.

Acknowledgements

The authors acknowledge and express their gratitude for the financial support received from the Ministry of Education, Culture, Research, and Technology of the Republic of Indonesia, specifically through grant no. 076/E5/PG. 02.00. PT/2022, which funded the research. The authors affirm that there is no conflict of interest associated with the utilization of these research funds. Furthermore, the authors extend their appreciation to the students and fishers involved in the research conducted in the upper reaches of the Kampar Kanan River, Indonesia.

References

- [1] A.G. Hasselberg, I.Aakre, J. Scholtens *et al.*, “Fish for food and nutrition security in Ghana: Challenges and opportunities,” *Global Food Security*. vol 26: 100380, 2020 <https://doi.org/10.1016/j.gfs.2020.100380>
- [2] N.Tran, U.P, Rodriguez, C.Y.Chan., *et al.*, “Future scenarios of fish supply and demand for food and nutrition security in Bangladesh: An analysis with the AsiaFish model”, *Aquaculture*. Vol.568: 739288, 2023. <https://doi.org/10.1016/j.aquaculture.2023.739288>
- [3] C.Z. Levkoe, K. Lowitt, C. Nelson. “Fish as food: Exploring a food sovereignty approach to small-scale fisheries”. *Marine Policy*. vol 85: pp.65-70, 2017. <https://doi.org/10.1016/j.marpol.2017.08.018>
- [4] M. Ravichandran, N.S. Hettiarachchy, V. Ganesh *et al.*, “Enhancement of antimicrobial activities of naturally occurring phenolic compounds by nanoscale delivery against listeria monocytogenes, escherichia coli O157:H7 and salmonella typhimurium in broth and chicken meat system”. *Journal of Food Safety*. vol 31, no. 4, pp. 462-471,2011. <https://doi.org/10.1111/j.1745-4565.2011.00322.x>
- [5] N. Aryani, I. Suharman, Azrita, *et al.*, “Diversity and distribution of fish fauna of upstream and downstream areas at Koto Panjang Reservoir, Riau Province, Indonesia”. *F1000 Research*. vol 8, pp. 1435. 2020. <https://f1000research.com/articles/8-1435/v2>
- [6] Syafraldi, Dahelmi, D.I. Roesma, *et al.*, “Length-weight relationships and condition factor of Two-Spot Catfish (*Mystus nigriceps* [Valenciennes, 1840]) Pisces, Bagridae], from Kampar Kanan River and Kampar Kiri River in Indonesia”. *Pakistan Journal of Biological Science*, vol. 23, no. 12, pp. 1636-1642, 2020. <https://scialert.net/fulltext/?doi=pjbs.2020.1636.1642>
- [7] N. Aryani, “Native species in Kampar Kanan River, Riau Province Indonesia. *International Journal of Fisheries and Aquatic Studies*”, vol.2, no.5, pp. 213-217. 2015. <https://www.fisheriesjournal.com/archives/2015/vol2issue5/PartD/2-5-63.pdf>
- [8] Azrita, H. Syandri, N. Aryani, *et al.*, “The utilization of new products formulated from water coconut, palm sap sugar, and fungus to increase nutritional feed quality, feed

- efficiency, growth, and carcass of gurami sago (*Osphronemus goramy* Lacepède, 1801) juvenile”. *F1000 Research*. vol. 10. :1121, pp. 1-16, 2021. <https://f1000research.com/articles/10-1121/v1>
- [9] C. Bene, R. Arthur, H. Norbury *et al.*, “Contribution of Fisheries and Aquaculture to Food Security and Poverty Reduction: Assessing the Current Evidence”. *World Development*, 79: 177–196. 2016. <http://dx.doi.org/10.1016/j.worlddev.2015.11.007>
- [10] N. Aryani, I. Suharman, S. Hasibuan, *et al.*, “Fatty acid composition on diet and carcasses, growth, body indices and profile serum of Asian redbtail catfish (*Hemibagrus nemurus*) fed a diet containing different levels of EPA and DHA”. *F1000Research*. 11:1409.pp.1-23, 2023. <https://doi.org/10.12688/f1000research.126487.2>
- [11] E.N. Kingsley, O.O. Cyril, O.I. Patience, “Potential contribution of selected wild fish species to the minerals intake of Pregnant and Lactating Women, Children and Adults in Rural Riverine Communities of Edo State: Insights and Outcomes”. *Measurement: Food*, 8, 100063 : 1-7, 2022. <https://doi.org/10.1016/j.meafoc.2022.100063>
- [12] A.A. Sausa, L. Nora, D.L.A. Lopes, *et al.*, Vegetable choline in feed for Nile tilapia (*Oreochromis niloticus*) raised in a biofloc technology system (BFT): Biofloc composition, chemical composition, and fatty acid profiles in meat“. *Aquaculture*. vol. 545. 737174, 2021. <https://doi.org/10.1016/j.aquaculture.2021.737174>
- [13] A.M. Eltweri, A.L. Thomas, M. Metcalfe, *et al.*, Potential applications of fish oils rich in omega-3 polyunsaturated fatty acids in the management of gastrointestinal cancer. *Clinical Nutrition*. vol. 36, no. 1, pp. 65-78. 2017. <https://doi.org/10.1016/j.clnu.2016.01.007>
- [14] P.C. Haycock, M.C. Borges, K. Burrows, *et al.*, “The association between genetically elevated polyunsaturated fatty acids and risk of cancer”. *eBiomedicine*. vol.91: 104510, 2023. <https://doi.org/10.1016/j.ebiom.2023.104510>
- [15] E.K. Aglago, I. Huybrechts, N. Murphy, *et al.*, “Consumption of Fish and Long-chain n-3 Polyunsaturated Fatty Acids Is Associated With Reduced Risk of Colorectal Cancer in a Large European Cohort”. *Clinical Gastroenterology and Hepatology*. vol. 18, no. 3, pp. 654-666.e6. 2020. <https://doi.org/10.1016/j.cgh.2019.06.031>
- [16] H. Jiang, X. Shi, T.Y. Fan, *et al.*, “Dietary omega-3 polyunsaturated fatty acids and fish intake and risk of age-related macular degeneration” . *Clinical Nutrition*. vol.40, no. 12, pp: 5662-5673. 2021. [https://www.clinicalnutritionjournal.com/article/S0261-5614\(21\)00474-X/fulltext](https://www.clinicalnutritionjournal.com/article/S0261-5614(21)00474-X/fulltext)
- [17] M.A. Rincón-Cervera, NV. González-Barriga, R. Valenzuela, *et al.*, “Profile and distribution of fatty acids in edible parts of commonly consumed marine fishes in Chile”, *Food Chemistry*, vol. 274, pp. 123-129, 2018. <https://doi.org/10.1016/j.foodchem.2018.08.113>
- [18] S. Sun, T. Ren, X. Li., *et al.*, “Polyunsaturated fatty acids synthesized by freshwater fish: A new insight to the roles of elovl2 and elovl5 in vivo”, *Biochemical and Biophysical Research Communications*, vol. 532, pp. 414-419, 2020. <https://doi.org/10.1016/j.bbrc.2020.08.074>
- [19] M.B. Behyar, M. Hasanzadeh, F. Seidi, *et al.*, “Sensing of amino acids: Critical role of nanomaterials for the efficient biomedical analysis”, *Microchemical Journal*, vol. 188, pp. 108452, 2023 <https://doi.org/10.1016/j.microc.2023.108452>

- [20] P. Shi, K. Liao, J. Xu, *et al.*, “Eicosapentaenoic acid mitigates palmitic acid-induced heat shock response, inflammation and repair processes in fish intestine”, *Fish & Shellfish Immunology*, vol. 124, pp. 362-371, 2022. <https://doi.org/10.1016/j.fsi.2022.04.011>
- [21] J. Wen, L. Zeng, Y. Xu, *et al.*, Proximate composition, amino acid and fatty acid composition of fish maws, *Natural Product Research: Formerly*, vol. 30, no. 2, pp. 214-217, 2015. <https://doi.org/10.1080/14786419.2015.1040790>
- [22] F.O.S. Duarte, F.G. de Paula, C.S. Prado, *et al.*, “Better fatty acids profile in fillets of Nile Tilapia (*Oreochromis niloticus*) supplemented with fish oil”, *Aquaculture*, Vol. 534, pp. 736241, 2021. <https://doi.org/10.1016/j.aquaculture.2020.736241>
- [23] K.L. Weaver, P. Ivester, J.A. Chilton, *et al.*, “The Content of Favorable and Unfavorable Polyunsaturated Fatty Acids Found in Commonly Eaten Fish”, *Journal of the American Dietetic Association*, vol. 108, No. 7, pp. 1178–1185, 2008. <https://doi.org/10.1016/j.jada.2008.04.023>
- [24] AOAC, “Official Methods of Analysis. 15th Edn., Association of Official Analytical Chemists (AOAC)”, Washington, DC., USA.1990. Reference Source
- [25] S.A. Cohen, “Amino acid analysis using pre-column derivatization with 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate”, *Protein Sequencing Protocols*, vol. 211, pp. 143-54, 2003. <https://doi.org/10.1385/1-59259-047-0:039>
- [26] M.A. Rajion, J.G. McLean, R.N. Cahill, “Essential fatty acid metabolism in the fetal and neonatal lamb”, *Australian Journal of Biological Sciences*, vol. 38, no. 1, pp. 33-40, 1985. [Essential fatty acids in the fetal and newborn lamb - PubMed \(nih.gov\)](https://pubmed.ncbi.nlm.nih.gov/2711111/)
- [27] T.L. Ulbricht, D.A. Southgate, “Coronary heart disease: Seven dietary factors”, *The Lancet*, Vol 338, no. 8773, pp. 985–992, 1991. [https://doi.org/10.1016/0140-6736\(91\)91846-M](https://doi.org/10.1016/0140-6736(91)91846-M)
- [28] D.B. Duncan, “Multiple ranges and multiple F tests”, *Biometrics*, vol. 11, pp. 1–42, 1955. <https://doi.org/10.2307/3001478>
- [29] R. Froese, “Cube law, condition factor and weight–length relationships: History, meta-analysis and recommendations”. *Journal of Applied Ichthyology*, vol. 22, no. 4, pp. 241-253, 2006. <https://doi.org/10.1111/j.1439-0426.2006.00805.x>
- [30] O.O. Famofofo, & W.O. Abdul, “Biometry, condition factors and length-weight relationships of sixteen fish species in Iwopin fresh-water ecotype of Lekki Lagoon, Ogun State, Southwest Nigeria”, *Heliyon*, vol. 6, no. 1, 2020. <https://doi.org/10.1016/j.heliyon.2019.e02957>
- [31] T.B. Bagenal, F.W. Tesch, “Age and Growth. In: Bagenal, T., Ed., Methods for Assessment of Fish Production in Fresh Waters”, *Blackwell Science Publications*, pp. 101-136, 1978.
- [32] E.O. Ahmed, M.E. Ali, A.A. Aziz, “Length-weight relationships and Condition factors of six fish species in Atbara River and Khashm el-girba Reservoir, Sudan”, *International Journal of Agriculture Sciences*, vol. 3, no.1, pp. 65–70, 2011. <http://dx.doi.org/10.9735/0975-3710.3.1.65-70>
- [33] N.C. Ujjania, M.P.S. Kohli, L.L. Sharma, “Length-weight relationship and condition factors of Indian major carps (*C. catla*, *L. rohita* and *C. mrigala*) in Mahi Bajaj Sagar, India”, *Research Journal of Biology*, vol. 2, no.1, pp. 30-36, 2012. <https://doi.org/10.21077/ijf.2017.64.special-issue.76263-27>

- [34] J.P. Bonjour, "Calcium and phosphate: A duet of ions playing for bone health", *Journal of the American College of Nutrition*, vol. 30, no. 5, pp. 438S-48S, 2011. <https://doi.org/10.1080/07315724.2011.10719988>
- [35] B.P. Mohanty, T.V. Sankar, S. Ganguly, *et al.*, "Micronutrient composition of 35 food fishes from India and their significance in human nutrition", *Biological Trace Element Research*, vol. 174, pp. 448-458, 2016. <https://doi.org/10.1007/s12011-016-0714-3>
- [36] F. Jabeen, A.A. Chudhry, "Chemical composition and fatty acid profiles of three freshwater species", *Food Chemistry*, vol. 125, pp. 991-996, 2011. <https://doi.org/10.1016/j.foodchem.2010.09.103>
- [37] A.A. Nurnadia, A. Azrina, I. Amin, "Proximate composition and energetic value of selected marine fish and shellfish from the west coast of peninsular Malaysia". *International Food Research Journal*. Vol 18, pp. 137-148, 2011. <https://core.ac.uk/download/pdf/153807129.pdf>
- [38] S.C. Dyall, L. Balas, N.G. Bazan, *et al.*, "Polyunsaturated fatty acids and fatty acid-derived lipid mediators: Recent advances in the understanding of their biosynthesis, structures and functions", *Progress in Lipid Research*. vol. 86, pp. 101165, 2022. <https://doi.org/10.1016/j.plipres.2022.101165>
- [39] N. Kim, U.D.Sohn, V. Mangannan, *et al.*, "Leukotrienes in Acetylcholine-Induced Contraction of Esophageal Circular Smooth Muscle in Experimental", *Gastroenterology*, vol. 112, no.5, pp. 1548-1558, 1997. [https://doi.org/10.1016/S0016-5085\(97\)70036-2](https://doi.org/10.1016/S0016-5085(97)70036-2)
- [40] D. Swanson, R. Block, S.A.Mousa, "Omega-3 Fatty Acids EPA and DHA: Health Benefits Throughout Life". *Advances in Nutrition*, vol. 3, no.1, pp. 1-7,2012. <https://doi.org/10.3945/an.111.000893>
- [41] R.P. Mason, S.C.R. Sherratt, L.H. Eckel, "Omega-3-fatty acids: Do they prevent cardiovascular disease?", *Best Practice & Research Clinical Endocrinology & Metabolism*, vol.37, no 3, 101681, 2023. <https://doi.org/10.1016/j.beem.2022.101681>.
- [42] J. M. Seddon, J. Cote, N. Davis, *et al.*, "Progression of age-related macular degeneration. Progression of age-related macular degeneration". *Arch Ophthalmol*. Vol. 121, no. 6, pp. 785-792, 2003. <https://doi.org/10.1001/archophth.121.6.785>
- [43] FAO/WHO: Fat and Oils in Human Nutrition: Report of a Joint Expert Consultation. FAO/WHO, Rome, Italy.
- [44] S.M. Pawar, S.R. Sonaware, "Fish muscle protein highest source of energy". *International Journal of Biodiversity and Conservation*. Vol. 5, no. 7, pp. 433-435, 2013. <https://doi.org/10.5897/IJBC12.043>
- [45] D. Tomé, C. Bos, "Lysine requirement through the human life cycle", *The Journal of Nutrition*, vol. 137, no.6, pp. 1642S-1645S, 2007. <https://doi.org/10.1093/jn/137.6.1642S>
- [46] M. Watford, "Glutamine and glutamate: Nonessential or essential amino acids?", *Animal Nutrition*, vol. 1, no. 3, pp. 119-122, 2015. <https://doi.org/10.1016/j.aninu.2015.08.008>
- [47] X. Li, S. Zheng, G. Wu, "Nutrition and metabolism of glutamate and glutamine in fish", *Amino Acids*. vol.52, no.5, pp. 671-691, 2020. <https://doi.org/10.1007/s00726-020-02851-2>

- [48] M. Li, Y. Wu, L. Ye, “ The Role of Amino Acids in Endothelial Biology and Function”, *Cells*, vol.11, no.8, pp. 1372, 2022. <https://doi.org/10.3390/cells11081372>



Webmail
Univ. Bung Hatta

hafrizal syandri <syandri_1960@bunghatta.ac.id>

6639837: Revision requested

4 messages

International Journal of Food Science <ijfs@hindawi.com>
Reply-To: International Journal of Food Science <drajadurai@wiley.com>
To: Hafrijal Syandri <syandri_1960@bunghatta.ac.id>

Fri, Dec 1, 2023 at 6:35 PM

WILEY

Dear Dr. Hafrijal Syandri,

In order for your submission "Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security " to "International Journal of Food Science" to proceed further in the review process, you will need to revise your manuscript.

Reason & Details:

“

The reviewer suggests a few remarks before the acceptance of the manuscript.

When you have finished revising, follow the link below to submit your revision:

MANUSCRIPT DETAILS

Kind regards,
Deepanaa Rajadurai
International Journal of Food Science

Reviewer Comments:

“

Reviewer 1 Comments to the Author

The article is very much improved. The author has made a considerable effort but a few remarks are mentioned below.

animal material not animal materials

TI : I know that this is the formula proposed by Ulbricht, D.A. Southgate. I wanted to warn you that C18:0 should not be considered a pro-atherogenic fatty acid.

The principle of lipid health indices is to make the relationship between protherogens and antiatherogens.

This email was sent to syandri_1960@bunghatta.ac.id by John Wiley & Sons, Inc.

111 River Street, Hoboken, NJ 07030 USA. 877-762-2974.

For more information, please see our [privacy policy](#).

hafrizal syandri <syandri_1960@bunghatta.ac.id>
To: International Journal of Food Science <drajadurai@wiley.com>

Sat, Dec 9, 2023 at 4:40 AM

Dear

Deepanaa Rajadurai
International Journal of Food Science

Thank you for your email on December 1, 2023. The author has revised manuscript No. 6639837. We appreciate the reviewer's contribution that C18:0 should not be considered a pro-atherogenic fatty acid. However, to ensure alignment with the Thrombogenic Index analysis, we request further clarification on whether the reviewer expected to remove C18:0 from our data (green highlight in the manuscript). We are delighted to get this information.

Best regards

Hafrijal Syandri

[Quoted text hidden]

Deepanaa Rajadurai <deepanaa.rajadurai@hindawi.com>
Reply-To: Deepanaa Rajadurai <deepanaa.rajadurai@hindawi.com>
To: syandri_1960@bunghatta.ac.id

Sat, Dec 9, 2023 at 12:06 PM

Dear Dr. Syandri,

Thank you for your email regarding your manuscript.

Please be informed I have contacted the reviewer regarding your concern. Once I receive a response, I will get back to you shortly.

Please do not hesitate to get back in touch, if you have any further questions.

Best Regards,

Deepanaa

Deepanaa Rajadurai
Editorial Assistant



Hindawi

Hindawi.com | [Twitter](#) | [Facebook](#) | [LinkedIn](#) | [YouTube](#)

[Quoted text hidden]

, hafrizal syandri <syandri_1960@bunghatta.ac.id> wrote:

[Quoted text hidden]

The contents of this email and any attachments are confidential and intended only for the person or entity to whom it is addressed. If you are not the intended recipient, any use, review, distribution, reproduction or any action taken in reliance upon this message is strictly prohibited. If you received this message in error, please immediately notify the sender and permanently delete all copies of the email and any attachments.

[Click here for translations of this disclaimer.](#)

Deepanaa Rajadurai <deepanaa.rajadurai@hindawi.com>
Reply-To: Deepanaa Rajadurai <deepanaa.rajadurai@hindawi.com>
To: syandri_1960@bunghatta.ac.id

Mon, Dec 11, 2023 at 7:47 PM

Dear Dr. Syandri,

Hope you are good and well.

Please be informed that the reviewer has conveyed as follows "Regarding my remark about C18:0, it is preferable to recalculate the TI without using stearic fatty acid (C18:0) as a proatherogenic fatty acid".

Please do not hesitate to get back in touch, if you have any further questions.

Best Regards,

Deepanaa

Deepanaa Rajadurai
Editorial Assistant



Hindawi

[Hindawi.com](https://www.hindawi.com) | [Twitter](#) | [Facebook](#) | [LinkedIn](#) | [YouTube](#)

On Sat, 9 Dec at 5:06 AM , Deepanaa Rajadurai <deepanaa.rajadurai@hindawi.com> wrote:
Dear Dr. Syandri,

Thank you for your email regarding your manuscript.

Please be informed I have contacted the reviewer regarding your concern. Once I receive a response, I will get back to you shortly.

Please do not hesitate to get back in touch, if you have any further questions.

Best Regards,

Deepanaa

Deepanaa Rajadurai
Editorial Assistant



Hindawi

[Hindawi.com](#) | [Twitter](#) | [Facebook](#) | [LinkedIn](#) | [YouTube](#)

On Fri, 8 Dec at 9:40 PM , hafrizal syandri <syandri_1960@bunghatta.ac.id> wrote:

[Quoted text hidden]

The contents of this email and any attachments are confidential and intended only for the person or entity to whom it is addressed. If you are not the intended recipient, any use, review, distribution, reproduction or any action taken in reliance upon this message is strictly prohibited. If you received this message in error, please immediately notify the sender and permanently delete all copies of the email and any attachments.

[Click here for translations of this disclaimer.](#)

Research Article

Analysis of Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security

Azrita Azrita¹, Hafrijal Syandri^{1*}, Hazlina Ahamad Zakeri², Harfiandri Damanhuri¹, Netti Aryani³

¹Faculty of Fisheries and Marine Science, Universitas Bung Hatta, 25131, Padang-West Sumatra Province, Indonesia

²Faculty of Science and Marine Environment, Universiti Malaysia Terengganu, Malaysia.

³Faculty of Fisheries and Marine Science, Universitas Riau, 28293, Pekanbaru-Riau Province, Indonesia

*Corresponding author: syandri_1960@bunghatta.ac.id

Abstract

Fish have become an irreplaceable dietary source of animal protein, especially among households with low socioeconomic status in rural and urban areas of Indonesia. This study aimed to analyse the proximate composition, minerals, fatty acids, and amino acids of three local Bagridae fish species in Indonesia. The standard AOAC method was employed to examine the proximate composition of the carcass, and the analysis of amino acids and fatty acids was conducted through HPLC and GC-MS, respectively. The mineral content was determined using AAS. The nutrient composition results of *Hemibagrus nemurus*, *Hemibagrus wyckii*, and *Mystus nigriceps* revealed that the protein content was 24.26%, 22.57%, and 21.39% (%dry weight), whereas the total lipid content was 6.64%, 7.47% and 7.75%, respectively. Regarding mineral contents, the calcium levels ranged from 1.49–1.66 mg/g, iron levels from 28.35–40.36 µg/g, and zinc levels from 24.03–54.46 µg/g. Among the fatty acids, palmitic acid was the most abundant in all three species, accounting for 25.59–30.70% of the total fatty acids. Additionally, significant amounts of C18:1 (oleic acid), C18:0 (stearic acid), and C20:4 (arachidonic acid) were also detected as primary fatty acids. The calculated atherogenic index values in the three species of Bagridae fish ranged from 0.73–0.99, while the thrombogenic index values varied between 0.54–0.75. The predominant amino acids found in the three species of Bagridae fish were glutamic acid with their concentrations ranging from 9.10–24.34%. These results indicate that consuming the meat of these three freshwater Bagridae fish species caught in the wild does not pose any health risks to consumers. They can be considered a safe and suitable food source with good nutritional quality.

Keywords: Amino acids, Fatty acids, Indonesia, Kampar Kanan River, Minerals

Commented [S1]: The abstract has been added with the study objectives and revised with yellow highlighting

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

1. Introduction

Fish serve as a high-quality protein source that is essential for satisfying dietary protein requirements, especially in areas where other protein sources are few or difficult to obtain [1,2]. Their accessibility can aid in ensuring a steady supply of food, particularly for communities whose main source of income is fishing or aquaculture [3]. Fish can contribute to a more varied selection of protein sources in communities, resulting in a diet that is more nutritious and balanced. Dietary fish can treat nutrient deficits and enhance general nutritional health. Fish are generally considered safe and offer a lower risk of contamination with harmful substances compared to some land-based animal products. A reliable indicator of a fish's quality, nutritional value, physiological status, and environment is its chemical makeup, which is found in fish flesh [4].

Hemibagrus nemurus, locally known as "baung", *Hemibagrus wyckii*, known as "geso" and *Mystus nigriceps*, known as "ingir-ingir," are three species of catfish found in the Kanan River, Indonesia [5,6]. Due to their high market value, these species have been identified as significant economic resources for rural communities [7]. Therefore, the analysis of the nutritional quality of fish meat in the study area is very important to determine whether it is beneficial to human health.

Knowing the composition of fatty acids and amino acids in fish meat is an essential factor that should not be ignored [8]. This information provides valuable insight into the nutritional quality of fish and the values of more nutritious and healthy foods [9]. In addition, a better understanding of the composition of fatty acids and amino acids can assist in selecting a suitable feed [10], developing a balanced diet [11], and maintaining optimal fish meat nutrition [12].

Omega-3 polyunsaturated fatty acids (PUFAs), particularly docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA) derived from fish oil, exhibit anti-inflammatory and anti-gastrointestinal cancer properties, making them crucial as immune-boosting nutrients [13,14]. Studies have demonstrated that ω -3 and ω -6 PUFAs exhibit beneficial effects in the management of cardiovascular diseases and cancers [15,16]. The fatty acid composition of PUFAs can differ among different types of fish, including both freshwater and marine species

[17, 18]. Amino acids (AAs), such as cysteine, arginine, tyrosine, glycine, proline, and serine, play a vital role in illness and stress situations and in the prevention of inflammation and repair processes in fish intestines [19, 20].

Furthermore, knowledge of the atherogenicity index (AI) and thrombogenicity index (TI) of fatty acids in the three local freshwater fish species of Bagridae can contribute to improving dietary guidelines. The AI and TI can be used to develop recommendations aimed at improving heart health [21,22]. These recommendations may involve selecting fish with favourable fatty acid profiles, allowing individuals to choose the most suitable fish species based on their specific needs [17, 23]. Therefore, this study aims to examine the nutritional composition profile, mineral content, amino acids, and fatty acids, and AI and TI of three freshwater fish species, namely, *H. nemurus*, *H. wyckii*, and *M. nigriceps*. These three fish species are generally consumed by people who live around the Kampar Kanan River in Kampar Regency, Riau Province, Indonesia.

2. Material and methods

2.1. Animal material

In this study, a total of 27 Bagridae fish belonging to three distinct species (*H. nemurus*, *H. wyckii*, and *M. nigriceps*) were examined. The specimens were obtained from local fishers operating in the upper stretches of the Kampar Kanan River in Kampar Regency, Riau Province (Figure 1). Sampling took place between June and August 2022 and was verified by the Fisheries Department, Faculty of Fisheries and Marine Science, Universitas Bung Hatta, Indonesia. The samples were placed in plastic bags and transported to the laboratory in an insulated icebox. The samples were then grouped based on their respective sampling locations.

2.2. Biometry measurement

Upon reaching the laboratory within 10 hours, the fish samples were subjected to gutting and washing. Each specimen was weighed (TW) and measured to determine the standard length (SL) as well as the depth or maximum height (H). TW was measured using balance scale (OHAUS Model CT 6000 USA) with a precision of 0.01 g. The length measurement was taken from the tip of the mouth to the end of the upper lobe of the caudal fin, representing the total body length, using a metre ruler with an accuracy of 1 mm. The height measurement involved a vertical assessment of the body's maximum height measured using a Digital Sekhmet Sigma

Commented [S2]: The author has been revised

Formatted: Highlight

Vernier calliper with an accuracy of 1 mm. The condition factor (CF) was calculated using the formula $CF = (TW/SL^3) \times 100$.

2.3. Proximate, amino acid, and mineral content analysis

The proximate carcass composition was assessed following the AOAC standard methods [24]. The samples were dried to a constant weight at 105 °C. The crude protein content was analysed using the standard Kjeldahl method, which involves multiplying the nitrogen content by a factor of 6.25. The Soxhlet method with ether extraction was employed to analyse crude lipids. The ash content was obtained by incinerating the samples at 550 °C for 16 h. The amino acid composition was determined using a high-performance liquid chromatography (HPLC) system, which consisted of a Waters® 1525 binary HPLC pump, Waters® 717 autosamplers, and Waters® 2475 multi-λ fluorescence detector (with excitation at 250 nm and emission at 395 nm). The samples were hydrolysed in triplicate with 6 N hydrochloric acid for 24 h at 11 °C [25].

For mineral data composition (Na, Mg, Ca, K, P, Fe, Cu, Mn, and Zn), the ashed feed sample and carcass were dissolved in 1 ml of hydrochloric acid (35% v/v Suprapur®, Merck) and then filtered with cellulose filter paper (Watchman No 1, International Ltd; Maidstone, UK) and diluted to an appropriate concentration for each elemental mineral. P levels were analysed with a Perkin-Elmer AA spectrophotometer mod 3110 (Norwalk, CT, USA).

2.4. Fatty acid analysis

The fish meat was examined utilizing the gas chromatography–mass spectrometry (GC–MS) method. The method of Folch et al. (1957) modified by Rajion et al. [26] was employed to perform the total fat extraction. This involved using a solvent system consisting of chloroform and methanol at a 2:1 (v/v) ratio. The process of transmethylation was conducted using a solution of 14% methanolic boron trifluoride. The fatty acid composition of the meat was then analysed at the SIG Laboratory, Accredited Testing Laboratory-LP-184-IDN.

The nutritional quality of lipids (AI and TI) was calculated using the following equations [27]:

$$AI = \frac{[(C12:0 + 4 \times C14:0 + C16:0)]}{\sum MUFA + \sum n-6 + \sum n-3}$$

$$TI = \frac{[(C14:0 + C16:0 + C18:0)]}{[(0.5 \times \sum MUFA) + (0.5 \times \sum n-6) + (3 \times \sum n-3) + (\sum n-3 / \sum n-6)]}$$

where:

AI = Atherogenic index

TI = Thrombogenic index

C12:0 = lauric acid

C14:0 = myristic acid

C16:0 = palmitic acid

C18:0 = stearic acid

Σ MUFA = sum of the concentrations of all monosaturated fatty acids

Σ n-6 = sum of the concentrations of n-6 polyunsaturated fatty acids

Σ n-3 = sum of the concentrations of n-3 polyunsaturated fatty acids

2.5. Data analysis

Data analysis was performed using the SPSS 16.0 software package (SPSS; Chicago, IL). The homogeneity of variances was assessed using Levene's test. One-way ANOVA was carried out to determine the parameters of proximate and amino acid composition, including the composition of fatty acids and nutritional quality of lipids (AI and TI) for each species, followed by the post hoc Duncan's multiple range test [28]. The results are reported as the mean values \pm standard errors for each parameter.

Commented [S3]: We appreciate the reviewer's contribution regarding the perspective that C18:0 should not be considered a pro-atherogenic fatty acid. However, to ensure alignment with the Thrombogenic Index analysis, we would like to request further clarification as to whether the authors expected to remove C18:0 from our data

Formatted: Highlight

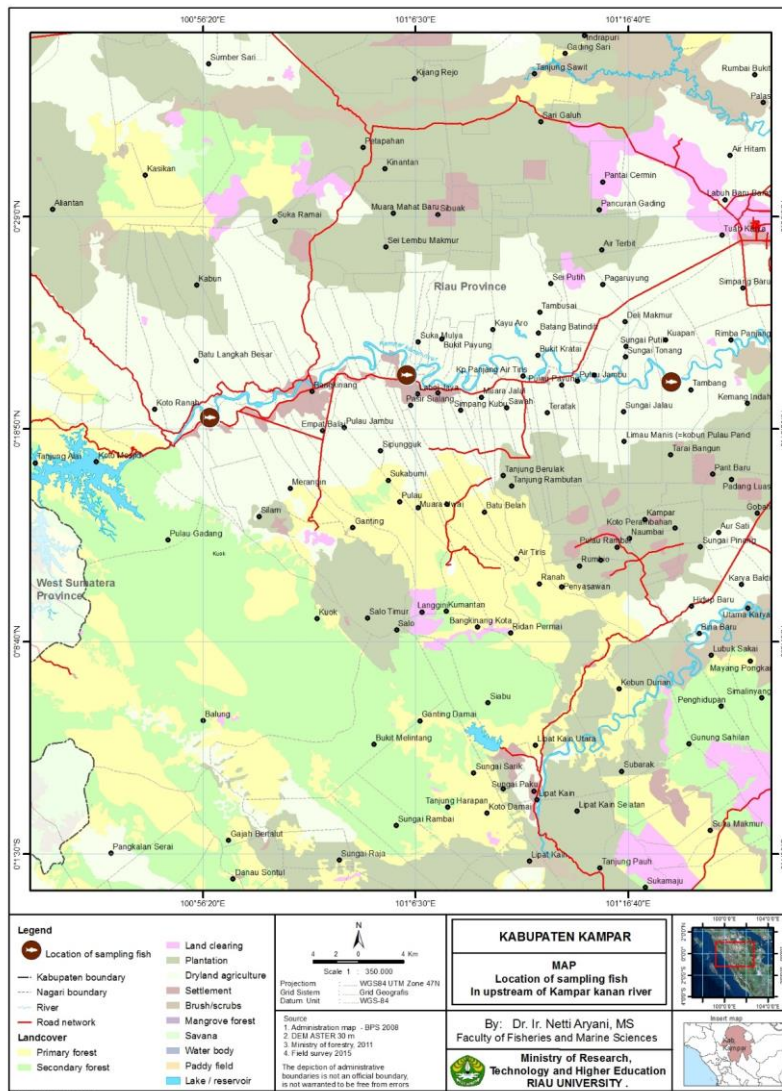


Figure 1. Map of Kampar Regency, Riau Province, and sampling locations of the three species of Bagridae fish.

Formatted: Highlight

Commented [S4]: Figure 1 has been revised

3. Results

Table 1 presents the average wet weight, standard length, height, and the results of meat nutritional composition analysis for three indigenous Indonesian freshwater Bagridae species. The moisture content (% wet weight) ranged from 82.40–85.39%. Among the species examined, *M. nigriceps* exhibited the lowest protein content at 21.39%, while *H. nemurus* displayed the highest value at 24.26%. However, *M. nigriceps* had a higher mineral content, including of iron and zinc.

Table 2 summarizes the fatty acid composition (%total fatty acid) for the three Bagridae fish species. C16:0 was the predominant fatty acid in the three species of Bagridae fish, with percentages ranging from 25.09–30.71%. The additional prominent fatty acids found were C18:1, C18:0, and C22:6. *H. nemurus* exhibited a ω -6/ ω -3 ratio below 1 (0.82), while *H. wyckii* and *M. nigriceps* had ratios of 1.11 and 1.02, respectively. The obtained PUFA/saturated fatty acid (SFA) (P/S) ratio varied between 0.61 and 0.76. The AI values varied between 0.76 and 0.99 and the TI between 0.54 and 0.75 (**Table 3**).

Table 4 presents the amino acid composition (% of total protein) of the three Bagridae species. Glutamic acid was the dominant amino acid, ranging from 19.29–24.34%, followed by aspartic acid, which ranged from 9.21–11.27%. The lysine content was consistent, ranging from 9.67–9.87%. In *H. nemurus*, the levels of various amino acids ranged from 0.86–24.35%. Similarly, in *H. wyckii*, the amino acid levels ranged from 0.76–21.58%, while in *M. nigriceps*, the levels ranged from 0.74–19.29%.

Table 1. Results of biometric, proximate and mineral composition of three species of Bagridae fish

	Species		
	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
<i>Biometric measurements</i>			
Total weight (g)	389.99 ± 24.96 ^a	1,390.33 ± 168.29 ^b	17.57 ± 1.53 ^c
Standard length (cm)	28.16 ± 0.53 ^a	44.14 ± 1.98 ^b	10.29 ± 0.15 ^c
Height (cm)	8.44 ± 0.16 ^a	8.82 ± 0.39 ^b	2.57 ± 0.03 ^c
Condition factor	1.63 ± 0.08	1.58 ± 0.07	1.56 ± 0.08
<i>Proximate composition</i>			
Crude protein (% DW)	24.26 ± 0.87 ^a	22.57 ± 0.37 ^b	21.39 ± 0.15 ^c
Crude fat (% DW)	6.64 ± 0.03 ^a	7.47 ± 0.02 ^b	7.75 ± 0.40 ^c
Crude ash (% DW)	1.94 ± 0.02 ^a	2.30 ± 0.09 ^b	1.59 ± 0.02 ^c

Moisture (% WW)	82.40 ± 2.51	85.39 ± 2.36	83.75 ± 2.21
<i>Mineral composition</i>			
<i>Macrominerals (mg/g)</i>			
Sodium	0.99 ± 0.00 ^a	1.61 ± 0.01 ^b	1.61 ± 0.02 ^c
Magnesium	1.12 ± 0.01 ^a	0.57 ± 0.02 ^b	1.15 ± 0.01 ^c
Calcium	1.66 ± 0.00 ^a	1.55 ± 0.02 ^b	1.49 ± 0.04 ^c
Potassium	0.71 ± 0.00 ^a	0.55 ± 0.02 ^b	0.43 ± 0.01 ^c
Phosphorous	7.03 ± 0.03 ^a	2.74 ± 0.02 ^b	6.45 ± 0.07 ^c
<i>Microminerals (µg/g)</i>			
Iron	28.30 ± 0.11 ^a	28.73 ± 0.08 ^b	40.36 ± 0.55 ^c
Copper	8.93 ± 0.03 ^a	7.46 ± 0.09 ^b	6.21 ± 0.32 ^c
Manganese	1.64 ± 0.02 ^a	2.64 ± 0.02 ^b	2.84 ± 0.02 ^c
Zinc	24.03 ± 0.45 ^a	24.61 ± 0.19 ^b	54.46 ± 0.17 ^c

Values are mean % ± SE; *Values in the same row followed by different letters are significantly different (p<0.05).

Table 2. Fatty acid composition of three species of Bagridae fish

Fatty acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
C12:0 (lauric acid)	1.97 ± 0.05 ^a	3.94 ± 0.01 ^b	4.27 ± 0.01 ^c
C14:0 (Meristic acid)	2.38 ± 0.03 ^a	2.72 ± 0.08 ^b	2.89 ± 0.01 ^c
C16:0 (Palmitic acid)	27.23 ± 0.06 ^a	30.70 ± 0.06 ^b	25.59 ± 0.01 ^c
C18:0 (Stearic acid)	16.17 ± 0.02 ^a	13.54 ± 0.02 ^b	10.64 ± 0.01 ^c
C20:0 (Arachidic acid)	0.23 ± 0.01 ^a	0.14 ± 0.01 ^b	0.37 ± 0.01 ^c
C16:1 (Palmitoleic acid)	1.78 ± 0.00 ^a	1.27 ± 0.04 ^b	4.93 ± 0.01 ^c
C18:1 (Oleic acid)	16.86 ± 0.01 ^a	16.18 ± 0.01 ^b	16.97 ± 0.01 ^c
C18:2 (Linoleic acid)	3.24 ± 0.02 ^a	3.84 ± 0.33 ^b	4.55 ± 0.001 ^c
C18:3 (Linolenic acid)	1.12 ± 0.01 ^a	0.82 ± 0.01 ^b	1.57 ± 0.00 ^c
C20:4 (Arachidonic acid)	12.13 ± 0.01 ^a	12.49 ± 0.09 ^b	12.41 ± 0.02 ^c
C20:5 (Eicosapentaenoic acid; EPA)	2.06 ± 0.08 ^a	2.34 ± 0.04 ^b	1.35 ± 0.01 ^c
C22:6 (Docosahexaenoic acid; DHA)	15.65 ± 0.01 ^a	11.56 ± 0.04 ^b	13.64 ± 0.02 ^c

Values are mean % ± SE; *Values in the same row with a different superscript are significantly different (p<0.05).

Table 3. Fatty acid ω-6/ω-3 and polyunsaturated/saturated fatty acid ratio of three species of Bagridae fish

	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
--	---------------------------	--------------------------	-------------------------

ω -6/ ω -3 ratio	0.82 ± 0.00^a	1.11 ± 0.02^b	1.02 ± 0.01^c
P/S ratio	0.69 ± 0.01^a	0.61 ± 0.06^b	0.76 ± 0.00^c
AI	0.87 ± 0.01^a	0.99 ± 0.06^b	0.73 ± 0.00^c
TI	0.63 ± 0.01^a	0.75 ± 0.03^b	0.54 ± 0.00^c

P/S, polyunsaturated/saturated fatty acid; AI, atherogenic index; TI, thrombogenic index
 Values are mean % \pm SE; *Values in the same row followed by different letters are significantly different ($p < 0.05$).

Table 4. Amino acid composition of three species of Bagridae fish

Amino acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
Aspartic acid	10.39 ± 0.01^a	9.61 ± 0.04^b	11.28 ± 0.02^c
Glutamic acid	24.34 ± 0.00^a	21.58 ± 0.41^b	19.29 ± 0.03^c
Serine	3.74 ± 0.00^a	5.40 ± 0.03^b	7.50 ± 0.06^c
Glycine	4.02 ± 0.00^a	5.77 ± 0.04^b	4.69 ± 0.02^c
Histidine	2.39 ± 0.00^a	2.90 ± 0.03^b	2.21 ± 0.02^c
Arginine	6.17 ± 0.01^a	8.27 ± 0.03^b	6.22 ± 0.03^c
Threonine	4.49 ± 0.01^a	4.19 ± 0.03^b	4.39 ± 0.04^c
Alanine	7.03 ± 0.00^a	6.04 ± 0.02^b	7.89 ± 0.01^c
Proline	2.40 ± 0.00^a	2.71 ± 0.01^b	2.21 ± 0.02^c
Tyrosine	3.91 ± 0.01^a	3.29 ± 0.06^b	3.82 ± 0.04^c
Valine	3.73 ± 0.00^a	4.02 ± 0.01^b	3.85 ± 0.04^c
Methionine	3.42 ± 0.00^a	1.96 ± 0.02^b	1.27 ± 0.03^c
Cystine	0.87 ± 0.00^a	0.76 ± 0.02^b	0.75 ± 0.03^c
Isoleucine	3.82 ± 0.001^a	3.96 ± 0.003^b	3.66 ± 0.002^c
Leucine	6.15 ± 0.000^a	6.69 ± 0.002^b	6.28 ± 0.004^c
Phenylalanine	5.13 ± 0.000^a	6.03 ± 0.005^b	5.18 ± 0.027^c
Lysine	9.61 ± 0.001^a	9.82 ± 0.026^b	9.87 ± 0.003^c

Values are mean % \pm SE; *Values in the same row followed by different letters are significantly different ($p < 0.05$).

4. Discussion

The results of this study provide important new information about the nutritional makeup of the studied species of Bagridae fish. The biometric measures of the fish yielded important information about their physical traits and health. Researchers are able to determine the average weight of fish at various lengths by examining the link between length and weight for a particular species. This knowledge is essential for understanding the trends in fish population growth, identifying potential shifts in growth rates over time, and assessing the general health and welfare of fish [29]. A variety of factors, including sex, age, maturity level, size, stomach fullness level, sampling strategies, sample sizes, and environmental circumstances, have an impact on fish health and the parameters that determine length–weight connections in fish [30]. However, none of these factors were considered in our study. In general, according to the theory put forth by Bagenal and Tesch [31], heavier fish of a given length are in better physiological condition. The condition factor is also used in fisheries science to compare the health, fatness, and condition of fish [32]. A condition factor of > 1 is favourable, indicating a good feed nutrition level and suitable living habitat [33]. Thus, based on the findings in this study, the Kampar Kanan River provides a suitable habitat for the fish, ensuring their accessibility and availability for community consumption.

The fish samples exhibited good nutritional profiles, with high amounts of proteins and lipids as well as moisture content contributing to their overall nutritional value, according to the results of proximate analysis and mineral composition (see Table 1). These results imply that the fish species studied provide a possible source of nutrients to solve issues with food security. The study of the mineral composition revealed the existence of critical minerals such as phosphorus (P), calcium (Ca), magnesium (Mg), iron (Fe), and zinc (Zn) in the fish samples in addition to proteins and lipids. All three Bagridae fish significantly differed from each other in terms of their crude ash content and mineral profile (see Table 1). However, in general, P and Ca are the two highest minerals found in all fish. Both of these macro minerals are good for human bone and tooth health, as key components of hydroxyapatite, the mineral matrix that provides strength and structure to bones and teeth [34].

In the current study, the meat of three species of Bagridae fish from the Kampar Kanan River in Indonesia was found to contain an abundance of microminerals such as Fe and Zn. *M. nigriceps* has been reported to exhibit higher mineral content, including of Fe and Zn. This

species is categorized as a small indigenous fish (SIF) with a maximum standard length of approximately 10.29 cm. According to [35], SIFs are generally known to be rich in Ca, Fe, and Mn.

In this study, all three Bagridae fish had a total fat content of approximately 7–8% by weight, with a higher water content ranging from 82.40–85.39% (Table 1). These values are similar to those found in wild and cultured sea bass *Dicentrarchus labrax* [36]. Additionally, the fish species can be grouped into different categories, including high-fat fish (>8%), medium-fat fish (4–8%), low-fat fish (2–4%), and lean fish (<2%) [37]. According to the classification established by [37], the three indigenous Bagridae fish species are characterized as having a moderate fat content level, which is reflected by the high levels of DHA and EPA in the meat of *H. nemurus*, *H. wyckii*, and *M. nigriceps* (Table 2). Therefore, we recommend that consuming the meat of these three fish species can indirectly boost food security in rural and urban communities.

All three Bagridae fish species analysed in the study contained arachidonic acid (C20:4), at 12.13% for *H. nemurus*, 12.49% for *H. wyckii*, and 12.41% for *M. nigriceps*. Arachidonic acid serves as a precursor for synthesizing prostaglandins and leukotrienes [38]. Leukotrienes are crucial in allergic responses, inducing smooth muscle contraction and increasing vascular permeability [39]. Additionally, all three species also contained EPA ranging from 1.35–2.34% and DHA ranging from 11.56–15.65%. The increased consumption of omega-3 fatty acids, such as EPA and DHA, has been linked to a decreased risk of pain development in older individuals and to the prevention of Alzheimer's disease [40,41].

As determined by the current study, the ω -6/ ω -3 ratios of the three Bagridae fish species ranged from 1:0.8 to 1:0.97. It was found that all three fish species had ω -6/ ω -3 ratios within the recommended range for a typical Indonesian diet. In contrast, a typical Western diet is characterized by a high ω -6/ ω -3 ratio of 10:1 to 20:1, while a Mediterranean diet has a ratio of 4:1 to 5:1 [42]. Additionally, the P/S ratio analysis showed that the three Bagridae fish species are sources of PUFAs that meet the requirements for food safety.

The Food and Agriculture Organization (FAO) and the World Health Organization (WHO) have recommended AI and TI values ranging from 0.4 to 0.5 for promoting human health [43]. In the current study, the observed AI values in the three freshwater species of Bagridae ranged from 0.73 to 0.99, while the TI values varied between 0.54 and 0.75. This finding is associated

with a notable disparity in the values of SFAs among the three Bagridae fish species. Based on these findings, we confirm that consuming the meat of the three freshwater species of Bagridae fish caught in the wild does not endanger consumer health and can serve as a suitable source of food safety and nutritional quality.

Fish with a high crude protein content, as shown by the three studied Bagridae fish (Table 1), are regarded as being nutritionally useful for human health. The protein and amino acid contents can directly indicate the nutritional quality of meat [44]. As shown in Table 4, *H. wyckii* had a higher content of essential amino acids (EAAs) than *H. nemurus* and *M. nigriceps*. Among the EAAs, lysine (Lys) was found to be present in the highest amount in all three species. Lys is necessary for tissue growth and repair, reduced stress-induced anxiety, and the conversion of fatty acids into energy [45-47].

More than 60% of the amino acid composition in all three Brigade fish studied comes from nonessential amino acids (NEAAs), particularly glutamic acid (Table 4). Glutamic acid and its amine, glutamine, are highly abundant amino acids found in fish in the free and protein-bound forms [48]. Other amino acids found at abundant levels in the three fish also play important physiological and biochemical functions. The NEAA arginine, for example, is involved in protein synthesis, creatine formation, and various other biochemical processes within the body. The EAAs leucine, phenylalanine, and methionine play important roles in the synthesis of nitric oxide, a molecule that can expand blood vessels [48].

4. Conclusions

The present study provides insightful information about the fatty acid and amino acid compositions, biometric measurements, proximate analysis, and mineral composition of three local freshwater species of Indonesian Bagridae fish. The findings show their potential as a nutrient-dense, sustainable food source that can support efforts to increase food security. These fish species might be incorporated into the local diet to increase nutrient intake and alleviate nutritional inadequacies, which would ultimately improve Indonesia's overall food security status.

Data Availability

The data utilized in this research has not been previously released or published in any form. The datasets employed and/or analysed during the present study can be obtained by contacting the corresponding author.

Conflicts of Interest

The authors confirm that they do not have any conflicting interests.

Acknowledgements

The authors acknowledge and express their gratitude for the financial support received from the Ministry of Education, Culture, Research, and Technology of the Republic of Indonesia, specifically through grant no. 076/E5/PG. 02.00. PT/2022, which funded the research. The authors affirm that there is no conflict of interest associated with the utilization of these research funds. Furthermore, the authors extend their appreciation to the students and fishers involved in the research conducted in the upper reaches of the Kampar Kanan River, Indonesia.

References

- [1] A.G. Hasselberg, I.Aakre, J. Scholtens *et al.*, “Fish for food and nutrition security in Ghana: Challenges and opportunities,” *Global Food Security*. vol 26: 100380, 2020 <https://doi.org/10.1016/j.gfs.2020.100380>
- [2] N.Tran, U.P, Rodriguez, C.Y.Chan., *et al.*, “Future cenarios of fish supply and demand for food and nutrition security in Bangladesh: An analysis with the AsiaFish model”, *Aquaculture*. Vol.568: 739288, 2023. <https://doi.org/10.1016/j.aquaculture.2023.739288>
- [3] C.Z. Levkoe, K. Lowitt, C. Nelson. “Fish as food: Exploring a food sovereignty approach to small-scale fisheries”. *Marine Policy*. vol 85: pp.65-70, 2017. <https://doi.org/10.1016/j.marpol.2017.08.018>
- [4] M. Ravichandran, N.S. Hettiarachchy, V. Ganesh *et al.*, “Enhancement of antimicrobial activities of naturally occurring phenolic compounds by nanoscale delivery against listeria monocytogenes, escherichia coli O157:H7 and salmonella typhimurium in broth and chicken meat system”. *Journal of Food Safety*. vol 31, no. 4, pp. 462-471,2011. <https://doi.org/10.1111/j.1745-4565.2011.00322.x>
- [5] N. Aryani, I. Suharman, Azrita, *et al.*, “Diversity and distribution of fish fauna of upstream and downstream areas at Koto Panjang Reservoir, Riau Province, Indonesia”. *F1000 Research*. vol 8, pp. 1435. 2020. <https://f1000research.com/articles/8-1435/v2>
- [6] Syafrialdi, Dahelmi, D.I. Roesma, *et al.*, “Length-weight relationships and condition factor of Two-Spot Catfish (*Mystus nigriceps* [Valenciennes, 1840]) Pisces, Bagridae], from Kampar Kanan River and Kampar Kiri River in Indonesia”. *Pakistan Journal of Biological Science*, vol. 23, no. 12, pp. 1636-1642, 2020. <https://scialert.net/fulltext/?doi=pjbs.2020.1636.1642>

- [7] N. Aryani, "Native species in Kampar Kanan River, Riau Province Indonesia. *International Journal of Fisheries and Aquatic Studies*", vol.2, no.5, pp. 213-217. 2015. <https://www.fisheriesjournal.com/archives/2015/vol2issue5/PartD/2-5-63.pdf>
- [8] Azrita, H. Syandri, N. Aryani, *et al.*, "The utilization of new products formulated from water coconut, palm sap sugar, and fungus to increase nutritional feed quality, feed efficiency, growth, and carcass of gurami sago (*Osphronemus goramy* Lacepède, 1801) juvenile". *F1000 Research*. vol. 10. :1121, pp. 1-16, 2021. <https://f1000research.com/articles/10-1121/v1>
- [9] C. Bene, R. Arthur, H. Norbury *et al.*, "Contribution of Fisheries and Aquaculture to Food Security and Poverty Reduction: Assessing the Current Evidence". *World Development*, 79: 177–196. 2016. <http://dx.doi.org/10.1016/j.worlddev.2015.11.007>
- [10] N. Aryani, I. Suharman, S. Hasibuan, *et al.*, "Fatty acid composition on diet and carcasses, growth, body indices and profile serum of Asian redbtail catfish (*Hemibagrus nemurus*) fed a diet containing different levels of EPA and DHA". *F1000Research*. 11:1409.pp.1-23, 2023. <https://doi.org/10.12688/f1000research.126487.2>
- [11] E.N. Kingsley, O.O. Cyril, O.I. Patience, "Potential contribution of selected wild fish species to the minerals intake of Pregnant and Lactating Women, Children and Adults in Rural Riverine Communities of Edo State: Insights and Outcomes". *Measurement: Food*, 8, 100063 : 1-7, 2022. <https://doi.org/10.1016/j.meafoo.2022.100063>
- [12] A.A. Sausa, L. Nora, D.L.A. Lopes, *et al.*, "Vegetable choline in feed for Nile tilapia (*Oreochromis niloticus*) raised in a biofloc technology system (BFT): Biofloc composition, chemical composition, and fatty acid profiles in meat". *Aquaculture*. vol. 545. 737174, 2021. <https://doi.org/10.1016/j.aquaculture.2021.737174>
- [13] A.M. Eltweri, A.L. Thomas, M. Metcalfe, *et al.*, "Potential applications of fish oils rich in omega-3 polyunsaturated fatty acids in the management of gastrointestinal cancer. *Clinical Nutrition*. vol. 36, no. 1, pp. 65-78. 2017. <https://doi.org/10.1016/j.clnu.2016.01.007>
- [14] P.C. Haycock, M.C. Borges, K. Burrows, *et al.*, "The association between genetically elevated polyunsaturated fatty acids and risk of cancer". *eBiomedicine*. vol.91: 104510, 2023. <https://doi.org/10.1016/j.ebiom.2023.104510>
- [15] E.K. Aglago, I. Huybrechts, N. Murphy, *et al.*, "Consumption of Fish and Long-chain n-3 Polyunsaturated Fatty Acids Is Associated With Reduced Risk of Colorectal Cancer in a Large European Cohort". *Clinical Gastroenterology and Hepatology*. vol. 18, no. 3, pp. 654-666.e6. 2020. <https://doi.org/10.1016/j.cgh.2019.06.031>
- [16] H. Jiang, X. Shi, T.Y. Fan, *et al.*, "Dietary omega-3 polyunsaturated fatty acids and fish intake and risk of age-related macular degeneration". *Clinical Nutrition*. vol.40, no. 12, pp: 5662-5673. 2021. [https://www.clinicalnutritionjournal.com/article/S0261-5614\(21\)00474-X/fulltext](https://www.clinicalnutritionjournal.com/article/S0261-5614(21)00474-X/fulltext)
- [17] M.A. Rincón-Cervera, N.V. González-Barriga, R. Valenzuela, *et al.*, "Profile and distribution of fatty acids in edible parts of commonly consumed marine fishes in Chile", *Food Chemistry*, vol. 274, pp. 123-129, 2018. <https://doi.org/10.1016/j.foodchem.2018.08.113>
- [18] S. Sun, T. Ren, X. Li., *et al.*, "Polyunsaturated fatty acids synthesized by freshwater fish: A new insight to the roles of elovl2 and elovl5 in vivo", *Biochemical and Biophysical*

- Research Communications*, vol. 532, pp. 414-419, 2020. <https://doi.org/10.1016/j.bbr.2020.08.074>
- [19] M.B. Behyar, M. Hasanzadeh, F. Seidi, *et al.*, “Sensing of amino acids: Critical role of nanomaterials for the efficient biomedical analysis”, *Microchemical Journal*, vol. 188, pp. 108452, 2023 <https://doi.org/10.1016/j.microc.2023.108452>
- [20] P. Shi, K. Liao, J. Xu, *et al.*, “Eicosapentaenoic acid mitigates palmitic acid-induced heat shock response, inflammation and repair processes in fish intestine”, *Fish & Shellfish Immunology*, vol. 124, pp. 362-371, 2022. <https://doi.org/10.1016/j.fsi.2022.04.011>
- [21] J. Wen, L. Zeng, Y. Xu, *et al.*, Proximate composition, amino acid and fatty acid composition of fish maws, *Natural Product Research: Formerly*, vol. 30, no. 2, pp. 214-217, 2015. <https://doi.org/10.1080/14786419.2015.1040790>
- [22] F.O.S. Duarte, F.G. de Paula, C.S. Prado, *et al.*, “Better fatty acids profile in fillets of Nile Tilapia (*Oreochromis niloticus*) supplemented with fish oil”, *Aquaculture*, Vol. 534, pp. 736241, 2021. <https://doi.org/10.1016/j.aquaculture.2020.736241>
- [23] K.L. Weaver, P. Ivester, J.A. Chilton, *et al.*, “The Content of Favorable and Unfavorable Polyunsaturated Fatty Acids Found in Commonly Eaten Fish”, *Journal of the American Dietetic Association*, vol. 108, No. 7, pp. 1178–1185, 2008. <https://doi.org/10.1016/j.jada.2008.04.023>
- [24] AOAC, “Official Methods of Analysis. 15th Edn., Association of Official Analytical Chemists (AOAC)”, Washington, DC., USA.1990. Reference Source
- [25] S.A. Cohen, “Amino acid analysis using pre-column derivatization with 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate”, *Protein Sequencing Protocols*, vol. 211, pp. 143-54, 2003. <https://doi.org/10.1385/1-59259-047-0:039>
- [26] M.A. Rajion, J.G. McLean, R.N. Cahill, “Essential fatty acid metabolism in the fetal and neonatal lamb”, *Australian Journal of Biological Sciences*, vol. 38, no. 1, pp. 33-40, 1985. [Essential fatty acids in the fetal and newborn lamb - PubMed \(nih.gov\)](https://pubmed.ncbi.nlm.nih.gov/10160000/)
- [27] T.L. Ulbricht, D.A. Southgate, “Coronary heart disease: Seven dietary factors”, *The Lancet*, Vol 338, no. 8773, pp. 985–992, 1991. [https://doi.org/10.1016/0140-6736\(91\)91846-M](https://doi.org/10.1016/0140-6736(91)91846-M)
- [28] D.B. Duncan, “Multiple ranges and multiple F tests”, *Biometrics*, vol. 11, pp. 1–42, 1955. <https://doi.org/10.2307/3001478>
- [29] R. Froese, “Cube law, condition factor and weight–length relationships: History, meta-analysis and recommendations”. *Journal of Applied Ichthyology*, vol. 22, no. 4, pp. 241-253, 2006. <https://doi.org/10.1111/j.1439-0426.2006.00805.x>
- [30] O.O. Famofo, & W.O. Abdul, “Biometry, condition factors and length-weight relationships of sixteen fish species in Iwopin fresh-water ecotype of Lekki Lagoon, Ogun State, Southwest Nigeria”, *Heliyon*, vol. 6, no. 1, 2020. <https://doi.org/10.1016/j.heliyon.2019.e02957>
- [31] T.B. Bagenal, F.W. Tesch, “Age and Growth. In: Bagenal, T., Ed., Methods for Assessment of Fish Production in Fresh Waters”, *Blackwell Science Publications*, pp. 101-136, 1978.
- [32] E.O. Ahmed, M.E. Ali, A.A. Aziz, “Length-weight relationships and Condition factors of six fish species in Atbara River and Khashm el-girba Reservoir, Sudan”, *International Journal of Agriculture Sciences*, vol. 3, no.1, pp. 65–70, 2011. <http://dx.doi.org/10.9735/0975-3710.3.1.65-70>

- [33] N.C. Ujjania, M.P.S. Kohli, L.L. Sharma, "Length-weight relationship and condition factors of Indian major carps (*C. catla*, *L. rohita* and *C. mrigala*) in Mahi Bajaj Sagar, India", *Research Journal of Biology*, vol. 2, no.1, pp. 30-36, 2012. <https://doi.org/10.21077/ijf.2017.64.special-issue.76263-27>
- [34] J.P. Bonjour, "Calcium and phosphate: A duet of ions playing for bone health", *Journal of the American College of Nutrition*, vol. 30, no. 5, pp. 438S-48S, 2011. <https://doi.org/10.1080/07315724.2011.10719988>
- [35] B.P. Mohanty, T.V. Sankar, S. Ganguly, *et al.*, "Micronutrient composition of 35 food fishes from India and their significance in human nutrition", *Biological Trace Element Research*, vol. 174, pp. 448-458, 2016. <https://doi.org/10.1007/s12011-016-0714-3>
- [36] F. Jabeen, A.A. Chudhry, "Chemical composition and fatty acid profiles of three freshwater species", *Food Chemistry*, vol. 125, pp. 991-996, 2011. <https://doi.org/10.1016/j.foodchem.2010.09.103>
- [37] A.A. Nurnadia, A. Azrina, I. Amin, "Proximate composition and energetic value of selected marine fish and shellfish from the west coast of peninsular Malaysia". *International Food Research Journal*. Vol 18, pp. 137-148, 2011. <https://core.ac.uk/download/pdf/153807129.pdf>
- [38] S.C. Dyall, L. Balas, N.G. Bazan, *et al.*, "Polyunsaturated fatty acids and fatty acid-derived lipid mediators: Recent advances in the understanding of their biosynthesis, structures and functions", *Progress in Lipid Research*. vol. 86, pp. 101165, 2022. <https://doi.org/10.1016/j.plipres.2022.101165>
- [39] N. Kim, U.D.Sohn, V. Mangannan, *et al.*, "Leukotrienes in Acetylcholine-Induced Contraction of Esophageal Circular Smooth Muscle in Experimental", *Gastroenterology*, vol. 112, no.5, pp. 1548-1558, 1997. [https://doi.org/10.1016/S0016-5085\(97\)70036-2](https://doi.org/10.1016/S0016-5085(97)70036-2)
- [40] D. Swanson, R. Block, S.A.Mousa, "Omega-3 Fatty Acids EPA and DHA: Health Benefits Throughout Life". *Advances in Nutrition*, vol. 3, no.1, pp. 1-7,2012. <https://doi.org/10.3945/an.111.000893>
- [41] R.P. Mason, S.C.R. Sherratt, L.H. Eckel, "Omega-3-fatty acids: Do they prevent cardiovascular disease?", *Best Practice & Research Clinical Endocrinology & Metabolism*, vol.37, no 3, 101681, 2023. <https://doi.org/10.1016/j.beem.2022.101681>.
- [42] J. M. Seddon, J. Cote, N. Davis, *et al.*, "Progression of age-related macular degeneration. Progression of age-related macular degeneration". *Arch Ophthalmol*. Vol. 121, no. 6, pp. 785-792, 2003. <https://doi.org/10.1001/archophth.121.6.785>
- [43] FAO/WHO: Fat and Oils in Human Nutrition: Report of a Joint Expert Consultation. FAO/WHO, Rome, Italy.
- [44] S.M. Pawar, S.R. Sonaware, "Fish muscle protein highest source of energy". *International Journal of Biodiversity and Conservation*. Vol. 5, no. 7, pp. 433-435, 2013. <https://doi.org/10.5897/IJBC12.043>
- [45] D. Tomé, C. Bos, "Lysine requirement through the human life cycle", *The Journal of Nutrition*, vol. 137, no.6, pp. 1642S-1645S, 2007. <https://doi.org/10.1093/jn/137.6.1642S>
- [46] M. Watford, "Glutamine and glutamate: Nonessential or essential amino acids?", *Animal Nutrition*, vol. 1, no. 3, pp. 119-122, 2015. <https://doi.org/10.1016/j.aninu.2015.08.008>
- [47] X. Li, S. Zheng, G. Wu, "Nutrition and metabolism of glutamate and glutamine in fish", *Amino Acids*. vol.52, no.5, pp. 671-691, 2020.

<https://doi.org/10.1007/s00726-020-02851-2>

- [48] M. Li, Y. Wu, L. Ye, “ The Role of Amino Acids in Endothelial Biology and Function”, *Cells*, vol.11, no.8, pp. 1372, 2022. <https://doi.org/10.3390/cells11081372>

On Fri, 8 Dec at 9:40 PM , hafrizal syandri <syandri_1960@bunghatta.ac.id> wrote:

[Quoted text hidden]

The contents of this email and any attachments are confidential and intended only for the person or entity to whom it is addressed. If you are not the intended recipient, any use, review, distribution, reproduction or any action taken in reliance upon this message is strictly prohibited. If you received this message in error, please immediately notify the sender and permanently delete all copies of the email and any attachments.

[Click here for translations of this disclaimer.](#)

Research Article

Analysis of Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security

Azrita Azrita¹, Hafrijal Syandri^{1*}, Hazlina Ahamad Zakeri², Harfiandri Damanhuri¹, Netti Aryani³

¹Faculty of Fisheries and Marine Science, Universitas Bung Hatta, 25131, Padang-West Sumatra Province, Indonesia

²Faculty of Science and Marine Environment, Universiti Malaysia Terengganu, Malaysia.

³Faculty of Fisheries and Marine Science, Universitas Riau, 28293, Pekanbaru-Riau Province, Indonesia

*Corresponding author: syandri_1960@bunghatta.ac.id

Abstract

Fish have become an irreplaceable dietary source of animal protein, especially among households with low socioeconomic status in rural and urban areas of Indonesia. This study aimed to analyse the proximate composition, minerals, fatty acids, and amino acids of three local Bagridae fish species in Indonesia. The standard AOAC method was employed to examine the proximate composition of the carcass, and the analysis of amino acids and fatty acids was conducted through HPLC and GC-MS, respectively. The mineral content was determined using AAS. The nutrient composition results of *Hemibagrus nemurus*, *Hemibagrus wyckii*, and *Mystus nigriceps* revealed that the protein content was 24.26%, 22.57%, and 21.39% (%dry weight), whereas the total lipid content was 6.64%, 7.47% and 7.75%, respectively. Regarding mineral contents, the calcium levels ranged from 1.49–1.66 mg/g, iron levels from 28.35–40.36 µg/g, and zinc levels from 24.03–54.46 µg/g. Among the fatty acids, palmitic acid was the most abundant in all three species, accounting for 25.59–30.70% of the total fatty acids. Additionally, significant amounts of C18:1 (oleic acid), C18:0 (stearic acid), and C20:4 (arachidonic acid) were also detected as primary fatty acids. The calculated atherogenic index values in the three species of Bagridae fish ranged from 0.73–0.99, while the thrombogenic index values varied between 0.54–0.75. The predominant amino acids found in the three species of Bagridae fish were glutamic acid with their concentrations ranging from 9.10–24.34%. These results indicate that consuming the meat of these three freshwater Bagridae fish species caught in the wild does not pose any health risks to consumers. They can be considered a safe and suitable food source with good nutritional quality.

Keywords: Amino acids, Fatty acids, Indonesia, Kampar Kanan River, Minerals

Commented [S1]: The abstract has been added with the study objectives and revised with yellow highlighting

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

1. Introduction

Fish serve as a high-quality protein source that is essential for satisfying dietary protein requirements, especially in areas where other protein sources are few or difficult to obtain [1,2]. Their accessibility can aid in ensuring a steady supply of food, particularly for communities whose main source of income is fishing or aquaculture [3]. Fish can contribute to a more varied selection of protein sources in communities, resulting in a diet that is more nutritious and balanced. Dietary fish can treat nutrient deficits and enhance general nutritional health. Fish are generally considered safe and offer a lower risk of contamination with harmful substances compared to some land-based animal products. A reliable indicator of a fish's quality, nutritional value, physiological status, and environment is its chemical makeup, which is found in fish flesh [4].

Hemibagrus nemurus, locally known as "baung", *Hemibagrus wyckii*, known as "geso" and *Mystus nigriceps*, known as "ingir-ingir," are three species of catfish found in the Kampar Kanan River, Indonesia [5,6]. Due to their high market value, these species have been identified as significant economic resources for rural communities [7]. Therefore, the analysis of the nutritional quality of fish meat in the study area is very important to determine whether it is beneficial to human health.

Knowing the composition of fatty acids and amino acids in fish meat is an essential factor that should not be ignored [8]. This information provides valuable insight into the nutritional quality of fish and the values of more nutritious and healthy foods [9]. In addition, a better understanding of the composition of fatty acids and amino acids can assist in selecting a suitable feed [10], developing a balanced diet [11], and maintaining optimal fish meat nutrition [12].

Omega-3 polyunsaturated fatty acids (PUFAs), particularly docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA) derived from fish oil, exhibit anti-inflammatory and anti-gastrointestinal cancer properties, making them crucial as immune-boosting nutrients [13,14]. Studies have demonstrated that ω -3 and ω -6 PUFAs exhibit beneficial effects in the management of cardiovascular diseases and cancers [15,16]. The fatty acid composition of PUFAs can differ among different types of fish, including both freshwater and marine species

[17, 18]. Amino acids (AAs), such as cysteine, arginine, tyrosine, glycine, proline, and serine, play a vital role in illness and stress situations and in the prevention of inflammation and repair processes in fish intestines [19, 20].

Furthermore, knowledge of the atherogenicity index (AI) and thrombogenicity index (TI) of fatty acids in the three local freshwater fish species of Bagridae can contribute to improving dietary guidelines. The AI and TI can be used to develop recommendations aimed at improving heart health [21,22]. These recommendations may involve selecting fish with favourable fatty acid profiles, allowing individuals to choose the most suitable fish species based on their specific needs [17, 23]. Therefore, this study aims to examine the nutritional composition profile, mineral content, amino acids, and fatty acids, and AI and TI of three freshwater fish species, namely, *H. nemurus*, *H. wyckii*, and *M. nigriceps*. These three fish species are generally consumed by people who live around the Kampar Kanan River in Kampar Regency, Riau Province, Indonesia.

2. Material and methods

2.1. Animal material

In this study, a total of 27 Bagridae fish belonging to three distinct species (*H. nemurus*, *H. wyckii*, and *M. nigriceps*) were examined. The specimens were obtained from local fishers operating in the upper stretches of the Kampar Kanan River in Kampar Regency, Riau Province (Figure 1). Sampling took place between June and August 2022 and was verified by the Fisheries Department, Faculty of Fisheries and Marine Science, Universitas Bung Hatta, Indonesia. The samples were placed in plastic bags and transported to the laboratory in an insulated icebox. The samples were then grouped based on their respective sampling locations.

2.2. Biometry measurement

Upon reaching the laboratory within 10 hours, the fish samples were subjected to gutting and washing. Each specimen was weighed (TW) and measured to determine the standard length (SL) as well as the depth or maximum height (H). TW was measured using balance scale (OHAUS Model CT 6000 USA) with a precision of 0.01 g. The length measurement was taken from the tip of the mouth to the end of the upper lobe of the caudal fin, representing the total body length, using a metre ruler with an accuracy of 1 mm. The height measurement involved a vertical assessment of the body's maximum height measured using a

Commented [S2]: The author has been revised

Formatted: Highlight

Digital Sekhmet Sigma Vernier calliper with an accuracy of 1 mm. The condition factor (CF) was calculated using the formula $CF = (TW/SL^3) \times 100$.

2.3. Proximate, amino acid, and mineral content analysis

The proximate carcass composition was assessed following the AOAC standard methods [24]. The samples were dried to a constant weight at 105 °C. The crude protein content was analysed using the standard Kjeldahl method, which involves multiplying the nitrogen content by a factor of 6.25. The Soxhlet method with ether extraction was employed to analyse crude lipids. The ash content was obtained by incinerating the samples at 550 °C for 16 h. The amino acid composition was determined using a high-performance liquid chromatography (HPLC) system, which consisted of a Waters® 1525 binary HPLC pump, Waters® 717 autosamplers, and Waters® 2475 multi- λ fluorescence detector (with excitation at 250 nm and emission at 395 nm). The samples were hydrolysed in triplicate with 6 N hydrochloric acid for 24 h at 11 °C [25].

For mineral data composition (Na, Mg, Ca, K, P, Fe, Cu, Mn, and Zn), the ashed feed sample and carcass were dissolved in 1 ml of hydrochloric acid (35% v/v Suprapur®, Merck) and then filtered with cellulose filter paper (Watchman No 1, International Ltd; Maidstone, UK) and diluted to an appropriate concentration for each elemental mineral. P levels were analysed with a Perkin-Elmer AA spectrophotometer mod 3110 (Norwalk, CT, USA).

2.4. Fatty acid analysis

The fish meat was examined utilizing the gas chromatography–mass spectrometry (GC–MS) method. The method of Folch et al. (1957) modified by Rajion et al. [26] was employed to perform the total fat extraction. This involved using a solvent system consisting of chloroform and methanol at a 2:1 (v/v) ratio. The process of transmethylation was conducted using a solution of 14% methanolic boron trifluoride. The fatty acid composition of the meat was then analysed at the SIG Laboratory, Accredited Testing Laboratory-LP-184-IDN.

The nutritional quality of lipids (AI and TI) was calculated using the following equations [27]:

$$AI = \frac{[(C12:0 + 4 \times C14:0 + C16:0)]}{\sum MUFA + \sum n-6 + \sum n-3}$$

$$TI = \frac{[(C14:0 + C16:0)]}{[(0.5 \times \sum MUFA) + (0.5 \times \sum n-6) + (3 \times \sum n-3) + (\sum n-3 / \sum n-6)]}$$

where:

AI = Atherogenic index

TI = Thrombogenic index

C12:0 = lauric acid

C14:0 = myristic acid

C16:0 = palmitic acid

\sum MUFA = sum of the concentrations of all monosaturated fatty acids

\sum n-6 = sum of the concentrations of n-6 polyunsaturated fatty acids

\sum n-3 = sum of the concentrations of n-3 polyunsaturated fatty acids

2.5. Data analysis

Data analysis was performed using the SPSS 16.0 software package (SPSS; Chicago, IL). The homogeneity of variances was assessed using Levene's test. One-way ANOVA was carried out to determine the parameters of proximate and amino acid composition, including the composition of fatty acids and nutritional quality of lipids (AI and TI) for each species, followed by the post hoc Duncan's multiple range test [28]. The results are reported as the mean values \pm standard errors for each parameter.

Commented [S3]: We appreciate the reviewer's contribution regarding the perspective that C18:0 should not be considered a pro-atherogenic fatty acid. However, to ensure alignment with the Thrombogenic Index analysis, we would like to request further clarification as to whether the authors expected to remove C18:0 from our data

The author has recalculated the Thrombogenic Index (TI) without using stearic fatty acid (C18:0).

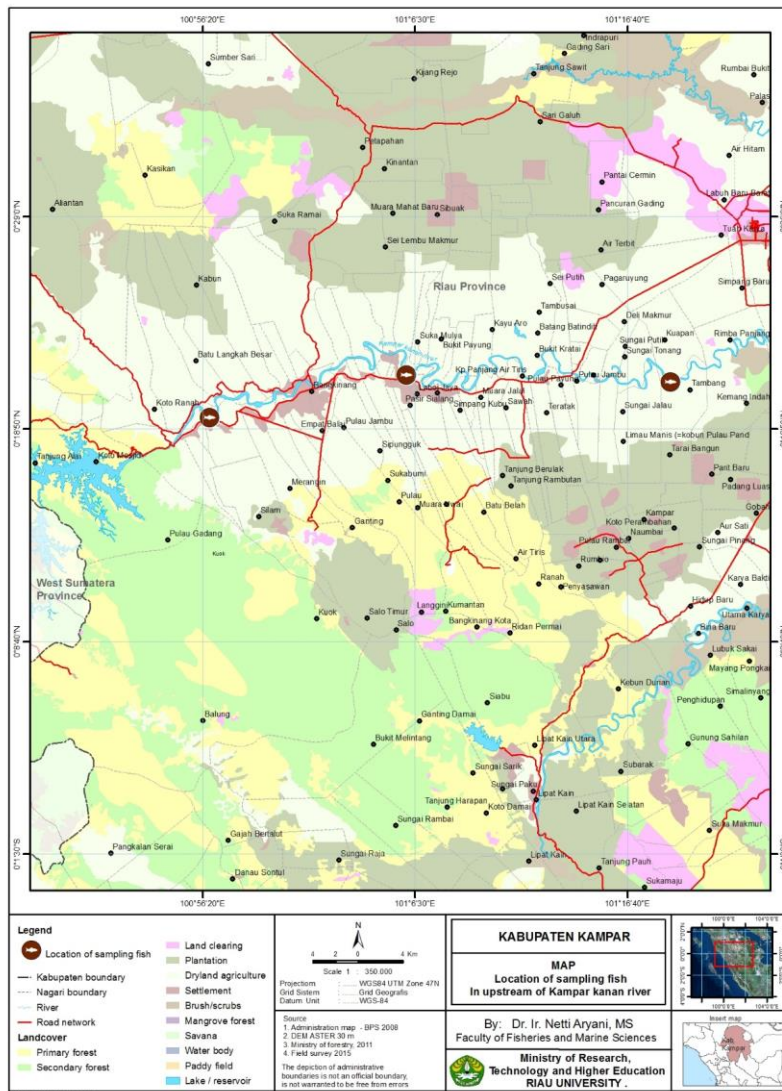


Figure 1. Map of Kampar Regency, Riau Province, and sampling locations of the three species of Bagridae fish.

Formatted: Highlight

Commented [S4]: Figure 1 has been revised

3. Results

Table 1 presents the average wet weight, standard length, height, and the results of meat nutritional composition analysis for three indigenous Indonesian freshwater Bagridae species. The moisture content (% wet weight) ranged from 82.40– 85.39%. Among the species examined, *M. nigriceps* exhibited the lowest protein content at 21.39%, while *H. nemurus* displayed the highest value at 24.26%. However, *M. nigriceps* had a higher mineral content, including of iron and zinc.

Table 2 summarizes the fatty acid composition (%total fatty acid) for the three Bagridae fish species. C16:0 was the predominant fatty acid in the three species of Bagridae fish, with percentages ranging from 25.09–30.71%. The additional prominent fatty acids found were C18:1, C18:0, and C22:6. *H. nemurus* exhibited a ω -6/ ω -3 ratio below 1 (0.82), while *H. wyckii* and *M. nigriceps* had ratios of 1.11 and 1.02, respectively. The obtained PUFA/saturated fatty acid (SFA) (P/S) ratio varied between 0.61 and 0.76. The AI values varied between 0.76 and 0.99 and the TI between 0.40 and 0.63 (**Table 3**).

Table 4 presents the amino acid composition (% of total protein) of the three Bagridae species. Glutamic acid was the dominant amino acid, ranging from 19.29–24.34%, followed by aspartic acid, which ranged from 9.21–11.27%. The lysine content was consistent, ranging from 9.67–9.87%. In *H. nemurus*, the levels of various amino acids ranged from 0.86–24.35%. Similarly, in *H. wyckii*, the amino acid levels ranged from 0.76–21.58%, while in *M. nigriceps*, the levels ranged from 0.74–19.29%.

Table 1. Results of biometric, proximate and mineral composition of three species of Bagridae fish

	Species		
	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
<i>Biometric measurements</i>			
Total weight (g)	389.99 ± 24.96 ^a	1,390.33 ± 168.29 ^b	17.57 ± 1.53 ^c
Standard length (cm)	28.16 ± 0.53 ^a	44.14 ± 1.98 ^b	10.29 ± 0.15 ^c
Height (cm)	8.44 ± 0.16 ^a	8.82 ± 0.39 ^b	2.57 ± 0.03 ^c
Condition factor	1.63 ± 0.08	1.58 ± 0.07	1.56 ± 0.08
<i>Proximate composition</i>			
Crude protein (% DW)	24.26 ± 0.87 ^a	22.57 ± 0.37 ^b	21.39 ± 0.15 ^c
Crude fat (% DW)	6.64 ± 0.03 ^a	7.47 ± 0.02 ^b	7.75 ± 0.40 ^c
Crude ash (% DW)	1.94 ± 0.02 ^a	2.30 ± 0.09 ^b	1.59 ± 0.02 ^c
Moisture (% WW)	82.40 ± 2.51	85.39 ± 2.36	83.75 ± 2.21

Commented [S5]: The author has recalculated the Thrombogenic Index (TI) without using stearic fatty acid (C18:0), and the TI value changed.

Formatted: Highlight

<i>Mineral composition</i>			
Macrominerals (mg/g)			
Sodium	0.99 ± 0.00 ^a	1.61 ± 0.01 ^b	1.61 ± 0.02 ^c
Magnesium	1.12 ± 0.01 ^a	0.57 ± 0.02 ^b	1.15 ± 0.01 ^c
Calcium	1.66 ± 0.00 ^a	1.55 ± 0.02 ^b	1.49 ± 0.04 ^c
Potassium	0.71 ± 0.00 ^a	0.55 ± 0.02 ^b	0.43 ± 0.01 ^c
Phosphorous	7.03 ± 0.03 ^a	2.74 ± 0.02 ^b	6.45 ± 0.07 ^c
Microminerals (µg/g)			
Iron	28.30 ± 0.11 ^a	28.73 ± 0.08 ^b	40.36 ± 0.55 ^c
Copper	8.93 ± 0.03 ^a	7.46 ± 0.09 ^b	6.21 ± 0.32 ^c
Manganese	1.64 ± 0.02 ^a	2.64 ± 0.02 ^b	2.84 ± 0.02 ^c
Zinc	24.03 ± 0.45 ^a	24.61 ± 0.19 ^b	54.46 ± 0.17 ^c

Values are mean % ± SE; *Values in the same row followed by different letters are significantly different (p<0.05).

Table 2. Fatty acid composition of three species of Bagridae fish

Fatty acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
C12:0 (lauric acid)	1.97 ± 0.05 ^a	3.94 ± 0.01 ^b	4.27 ± 0.01 ^c
C14:0 (Myristic acid)	2.38 ± 0.03 ^a	2.72 ± 0.08 ^b	2.89 ± 0.01 ^c
C16:0 (Palmitic acid)	27.23 ± 0.06 ^a	30.70 ± 0.06 ^b	25.59 ± 0.01 ^c
C18:0 (Stearic acid)	16.17 ± 0.02 ^a	13.54 ± 0.02 ^b	10.64 ± 0.01 ^c
C20:0 (Arachidic acid)	0.23 ± 0.01 ^a	0.14 ± 0.01 ^b	0.37 ± 0.01 ^c
C16:1 (Palmitoleic acid)	1.78 ± 0.00 ^a	1.27 ± 0.04 ^b	4.93 ± 0.01 ^c
C18:1 (Oleic acid)	16.86 ± 0.01 ^a	16.18 ± 0.01 ^b	16.97 ± 0.01 ^c
C18:2 (Linoleic acid)	3.24 ± 0.02 ^a	3.84 ± 0.33 ^b	4.55 ± 0.001 ^c
C18:3 (Linolenic acid)	1.12 ± 0.01 ^a	0.82 ± 0.01 ^b	1.57 ± 0.000 ^c
C20:4 (Arachidonic acid)	12.13 ± 0.01 ^a	12.49 ± 0.09 ^b	12.41 ± 0.02 ^c
C20:5 (Eicosapentaenoic acid; EPA)	2.06 ± 0.08 ^a	2.34 ± 0.04 ^b	1.35 ± 0.01 ^c
C22:6 (Docosahexaenoic acid; DHA)	15.65 ± 0.01 ^a	11.56 ± 0.04 ^b	13.64 ± 0.02 ^c

Values are mean % ± SE; *Values in the same row with a different superscript are significantly different (p<0.05).

Table 3. Fatty acid ω-6/ω-3 and polyunsaturated/saturated fatty acid ratio of three species of Bagridae fish

	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
ω-6/ω-3 ratio	0.82 ± 0.00 ^a	1.11 ± 0.02 ^b	1.02 ± 0.01 ^c

P/S ratio	0.69 ± 0.01 ^a	0.61 ± 0.06 ^b	0.76 ± 0.00 ^c
AI	0.87 ± 0.01 ^a	0.99 ± 0.06 ^b	0.73 ± 0.00 ^c
TI	0.40 ± 0.01 ^a	0.63 ± 0.01 ^b	0.43 ± 0.00 ^c

P/S, polyunsaturated/saturated fatty acid; AI, atherogenic index; TI, thrombogenic index
Values are mean % ± SE; *Values in the same row followed by different letters are significantly different (p<0.05).

Commented [S6]: The author has recalculated the Thrombogenic Index (TI) without using stearic fatty acid (C18:0), and the TI value changed.

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Table 4. Amino acid composition of three species of Bagridae fish

Amino acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
Aspartic acid	10.39 ± 0.01 ^a	9.61 ± 0.04 ^b	11.28 ± 0.02 ^c
Glutamic acid	24.34 ± 0.00 ^a	21.58 ± 0.41 ^b	19.29 ± 0.03 ^c
Serine	3.74 ± 0.00 ^a	5.40 ± 0.03 ^b	7.50 ± 0.06 ^c
Glycine	4.02 ± 0.00 ^a	5.77 ± 0.04 ^b	4.69 ± 0.02 ^c
Histidine	2.39 ± 0.00 ^a	2.90 ± 0.03 ^b	2.21 ± 0.02 ^c
Arginine	6.17 ± 0.01 ^a	8.27 ± 0.03 ^b	6.22 ± 0.03 ^c
Threonine	4.49 ± 0.01 ^a	4.19 ± 0.03 ^b	4.39 ± 0.04 ^c
Alanine	7.03 ± 0.00 ^a	6.04 ± 0.02 ^b	7.89 ± 0.01 ^c
Proline	2.40 ± 0.00 ^a	2.71 ± 0.01 ^b	2.21 ± 0.02 ^c
Tyrosine	3.91 ± 0.01 ^a	3.29 ± 0.06 ^b	3.82 ± 0.04 ^c
Valine	3.73 ± 0.00 ^a	4.02 ± 0.01 ^b	3.85 ± 0.04 ^c
Methionine	3.42 ± 0.00 ^a	1.96 ± 0.02 ^b	1.27 ± 0.03 ^c
Cystine	0.87 ± 0.00 ^a	0.76 ± 0.02 ^b	0.75 ± 0.03 ^c
Isoleucine	3.82 ± 0.001 ^a	3.96 ± 0.003 ^b	3.66 ± 0.002 ^c
Leucine	6.15 ± 0.000 ^a	6.69 ± 0.002 ^b	6.28 ± 0.004 ^c
Phenylalanine	5.13 ± 0.000 ^a	6.03 ± 0.005 ^b	5.18 ± 0.027 ^c
Lysine	9.61 ± 0.001 ^a	9.82 ± 0.026 ^b	9.87 ± 0.003 ^c

Values are mean % ± SE; *Values in the same row followed by different letters are significantly different (p<0.05).

4. Discussion

The results of this study provide important new information about the nutritional makeup of the studied species of Bagridae fish. The biometric measures of the fish yielded important information about their physical traits and health. Researchers are able to determine the average weight of fish at various lengths by examining the link between length and weight for a particular species. This knowledge is essential for understanding the trends in fish population growth, identifying potential shifts in growth rates over time, and assessing the general health and welfare of fish [29]. A variety of factors, including sex, age, maturity level, size, stomach fullness level, sampling strategies, sample sizes, and environmental circumstances, have an impact on fish health and the parameters that determine length–weight connections in fish [30]. However, none of these factors were considered in our study. In general, according to the theory put forth by [Bagenal and Tesch \[31\]](#), heavier fish of a given length are in better physiological condition. The condition factor is also used in fisheries science to compare the health, fatness, and condition of fish [32]. A condition factor of > 1 is favourable, indicating a good feed nutrition level and suitable living habitat [33]. Thus, based on the findings in this study, the Kampar Kanan River provides a suitable habitat for the fish, ensuring their accessibility and availability for community consumption.

The fish samples exhibited good nutritional profiles, with high amounts of proteins and lipids as well as moisture content contributing to their overall nutritional value, according to the results of proximate analysis and mineral composition (see Table 1). These results imply that the fish species studied provide a possible source of nutrients to solve issues with food security. The study of the mineral composition revealed the existence of critical minerals such as phosphorus (P), calcium (Ca), magnesium (Mg), iron (Fe), and zinc (Zn) in the fish samples in addition to proteins and lipids. All three Bagridae fish significantly differed from each other in terms of their crude ash content and mineral profile (see Table 1). However, in general, P and Ca are the two highest minerals found in all fish. Both of these macro minerals are good for human bone and tooth health, as key components of hydroxyapatite, the mineral matrix that provides strength and structure to bones and teeth [34].

In the current study, the meat of three species of Bagridae fish from the Kampar Kanan River in Indonesia was found to contain an abundance of microminerals such as Fe and Zn. *M. nigriceps* has been reported to exhibit higher mineral content, including of Fe and Zn. This species is categorized as a small indigenous fish (SIF) with a maximum standard length

of approximately 10.29 cm. According to [35], SIFs are generally known to be rich in Ca, Fe, and Mn.

In this study, all three Bagridae fish had a total fat content of approximately 7–8% by weight, with a higher water content ranging from 82.40–85.39% (Table 1). These values are similar to those found in wild and cultured sea bass *Dicentrarchus labrax* [36]. Additionally, the fish species can be grouped into different categories, including high-fat fish (>8%), medium-fat fish (4–8%), low-fat fish (2–4%), and lean fish (<2%) [37]. According to the classification established by [37], the three indigenous Bagridae fish species are characterized as having a moderate fat content level, which is reflected by the high levels of DHA and EPA in the meat of *H. nemurus*, *H. wyckii*, and *M. nigriceps* (Table 2). Therefore, we recommend that consuming the meat of these three fish species can indirectly boost food security in rural and urban communities.

All three Bagridae fish species analysed in the study contained arachidonic acid (C20:4), at 12.13% for *H. nemurus*, 12.49% for *H. wyckii*, and 12.41% for *M. nigriceps*. Arachidonic acid serves as a precursor for synthesizing prostaglandins and leukotrienes [38]. Leukotrienes are crucial in allergic responses, inducing smooth muscle contraction and increasing vascular permeability [39]. Additionally, all three species also contained EPA ranging from 1.35–2.34% and DHA ranging from 11.56–15.65%. The increased consumption of omega-3 fatty acids, such as EPA and DHA, has been linked to a decreased risk of pain development in older individuals and to the prevention of Alzheimer's disease [40,41].

As determined by the current study, the ω -6/ ω -3 ratios of the three Bagridae fish species ranged from 1:0.8 to 1:0.97. It was found that all three fish species had ω -6/ ω -3 ratios within the recommended range for a typical Indonesian diet. In contrast, a typical Western diet is characterized by a high ω -6/ ω -3 ratio of 10:1 to 20:1, while a Mediterranean diet has a ratio of 4:1 to 5:1 [42]. Additionally, the P/S ratio analysis showed that the three Bagridae fish species are sources of PUFAs that meet the requirements for food safety.

The Food and Agriculture Organization (FAO) and the World Health Organization (WHO) have recommended AI and TI values ranging from 0.4 to 0.5 for promoting human health [43]. In the current study, the observed AI values in the three freshwater species of Bagridae ranged from 0.73 to 0.99, while the TI values varied between 0.40 and 0.63. This finding is associated with a notable disparity in the values of SFAs among the three Bagridae

Commented [S7]: The author has recalculated the Thrombogenic Index (TI) without using stearic fatty acid (C18:0), and the TI value changed.

Formatted: Highlight

fish species. Based on these findings, we confirm that consuming the meat of the three freshwater species of Bagridae fish caught in the wild does not endanger consumer health and can serve as a suitable source of food safety and nutritional quality.

Fish with a high crude protein content, as shown by the three studied Bagridae fish (Table 1), are regarded as being nutritionally useful for human health. The protein and amino acid contents can directly indicate the nutritional quality of meat [44]. As shown in Table 4, *H. wyckii* had a higher content of essential amino acids (EAAs) than *H. nemurus* and *M. nigriceps*. Among the EAAs, lysine (Lys) was found to be present in the highest amount in all three species. Lys is necessary for tissue growth and repair, reduced stress-induced anxiety, and the conversion of fatty acids into energy [45-47].

More than 60% of the amino acid composition in all three Brigade fish studied comes from nonessential amino acids (NEAAs), particularly glutamic acid (Table 4). Glutamic acid and its amine, glutamine, are highly abundant amino acids found in fish in the free and protein-bound forms [48]. Other amino acids found at abundant levels in the three fish also play important physiological and biochemical functions. The NEAA arginine, for example, is involved in protein synthesis, creatine formation, and various other biochemical processes within the body. The EAAs leucine, phenylalanine, and methionine play important roles in the synthesis of nitric oxide, a molecule that can expand blood vessels [48].

4. Conclusions

The present study provides insightful information about the fatty acid and amino acid compositions, biometric measurements, proximate analysis, and mineral composition of three local freshwater species of Indonesian Bagridae fish. The findings show their potential as a nutrient-dense, sustainable food source that can support efforts to increase food security. These fish species might be incorporated into the local diet to increase nutrient intake and alleviate nutritional inadequacies, which would ultimately improve Indonesia's overall food security status.

Data Availability

The data utilized in this research has not been previously released or published in any form. The datasets employed and/or analysed during the present study can be obtained by contacting the corresponding author.

Conflicts of Interest

The authors confirm that they do not have any conflicting interests.

Acknowledgements

The authors acknowledge and express their gratitude for the financial support received from the Ministry of Education, Culture, Research, and Technology of the Republic of Indonesia, specifically through grant no. 076/E5/PG. 02.00. PT/2022, which funded the research. The authors affirm that there is no conflict of interest associated with the utilization of these research funds. Furthermore, the authors extend their appreciation to the students and fishers involved in the research conducted in the upper reaches of the Kampar Kanan River, Indonesia.

References

- [1] A.G. Hasselberg, I.Aakre, J. Scholtens *et al.*, “Fish for food and nutrition security in Ghana: Challenges and opportunities,” *Global Food Security*. vol 26: 100380, 2020 <https://doi.org/10.1016/j.gfs.2020.100380>
- [2] N.Tran, U.P, Rodriguez, C.Y.Chan., *et al.*, “Future scenarios of fish supply and demand for food and nutrition security in Bangladesh: An analysis with the AsiaFish model”, *Aquaculture*. Vol.568: 739288, 2023. <https://doi.org/10.1016/j.aquaculture.2023.739288>
- [3] C.Z. Levkoe, K. Lowitt, C. Nelson. “Fish as food: Exploring a food sovereignty approach to small-scale fisheries”. *Marine Policy*. vol 85: pp.65-70, 2017. <https://doi.org/10.1016/j.marpol.2017.08.018>
- [4] M. Ravichandran, N.S. Hettiarachchy, V. Ganesh *et al.*, “Enhancement of antimicrobial activities of naturally occurring phenolic compounds by nanoscale delivery against listeria monocytogenes, escherichia coli O157:H7 and salmonella typhimurium in broth and chicken meat system”. *Journal of Food Safety*. vol 31, no. 4, pp. 462-471,2011. <https://doi.org/10.1111/j.1745-4565.2011.00322.x>
- [5] N. Aryani, I. Suharman, Azrita, *et al.*, “Diversity and distribution of fish fauna of upstream and downstream areas at Koto Panjang Reservoir, Riau Province, Indonesia”. *F1000 Research*. vol 8, pp. 1435. 2020. <https://f1000research.com/articles/8-1435/v2>
- [6] Syafraldi, Dahelmi, D.I. Roesma, *et al.*, “Length-weight relationships and condition factor of Two-Spot Catfish (*Mystus nigriceps* [Valenciennes, 1840]) Pisces, Bagridae], from Kampar Kanan River and Kampar Kiri River in Indonesia”. *Pakistan Journal of Biological Science*, vol. 23, no. 12, pp. 1636-1642, 2020. <https://scialert.net/fulltext/?doi=pjbs.2020.1636.1642>
- [7] N. Aryani, “Native species in Kampar Kanan River, Riau Province Indonesia. *International Journal of Fisheries and Aquatic Studies*”, vol.2, no.5, pp. 213-217. 2015. <https://www.fisheriesjournal.com/archives/2015/vol2issue5/PartD/2-5-63.pdf>
- [8] Azrita, H. Syandri, N. Aryani, *et al.*, “The utilization of new products formulated from water coconut, palm sap sugar, and fungus to increase nutritional feed quality, feed

- efficiency, growth, and carcass of gurami sago (*Osphronemus goramy* Lacepède, 1801) juvenile”. *F1000 Research*. vol. 10. :1121, pp. 1-16, 2021. <https://f1000research.com/articles/10-1121/v1>
- [9] C. Bene, R. Arthur, H. Norbury *et al.*, “Contribution of Fisheries and Aquaculture to Food Security and Poverty Reduction: Assessing the Current Evidence”. *World Development*, 79: 177–196. 2016. <http://dx.doi.org/10.1016/j.worlddev.2015.11.007>
- [10] N. Aryani, I. Suharman, S. Hasibuan, *et al.*, “Fatty acid composition on diet and carcasses, growth, body indices and profile serum of Asian redbtail catfish (*Hemibagrus nemurus*) fed a diet containing different levels of EPA and DHA”. *F1000Research*. 11:1409, pp.1-23, 2023. <https://doi.org/10.12688/f1000research.126487.2>
- [11] E.N. Kingsley, O.O. Cyril, O.I. Patience, “Potential contribution of selected wild fish species to the minerals intake of Pregnant and Lactating Women, Children and Adults in Rural Riverine Communities of Edo State: Insights and Outcomes”. *Measurement: Food*, 8, 100063 : 1-7, 2022. <https://doi.org/10.1016/j.meafoc.2022.100063>
- [12] A.A. Sausa, L. Nora, D.L.A. Lopes, *et al.*, Vegetable choline in feed for Nile tilapia (*Oreochromis niloticus*) raised in a biofloc technology system (BFT): Biofloc composition, chemical composition, and fatty acid profiles in meat“. *Aquaculture*. vol. 545. 737174, 2021. <https://doi.org/10.1016/j.aquaculture.2021.737174>
- [13] A.M. Eltweri, A.L. Thomas, M. Metcalfe, *et al.*, Potential applications of fish oils rich in omega-3 polyunsaturated fatty acids in the management of gastrointestinal cancer. *Clinical Nutrition*. vol. 36, no. 1, pp. 65-78. 2017. <https://doi.org/10.1016/j.clnu.2016.01.007>
- [14] P.C. Haycock, M.C. Borges, K. Burrows, *et al.*, “The association between genetically elevated polyunsaturated fatty acids and risk of cancer”. *eBiomedicine*. vol.91: 104510, 2023. <https://doi.org/10.1016/j.ebiom.2023.104510>
- [15] E.K. Aglago, I. Huybrechts, N. Murphy, *et al.*, “Consumption of Fish and Long-chain n-3 Polyunsaturated Fatty Acids Is Associated With Reduced Risk of Colorectal Cancer in a Large European Cohort”. *Clinical Gastroenterology and Hepatology*. vol. 18, no. 3, pp. 654-666.e6. 2020. <https://doi.org/10.1016/j.cgh.2019.06.031>
- [16] H. Jiang, X. Shi, T.Y. Fan, *et al.*, “Dietary omega-3 polyunsaturated fatty acids and fish intake and risk of age-related macular degeneration” . *Clinical Nutrition*. vol.40, no. 12, pp: 5662-5673. 2021. [https://www.clinicalnutritionjournal.com/article/S0261-5614\(21\)00474-X/fulltext](https://www.clinicalnutritionjournal.com/article/S0261-5614(21)00474-X/fulltext)
- [17] M.A. Rincón-Cervera, NV. González-Barriga, R. Valenzuela, *et al.*, “Profile and distribution of fatty acids in edible parts of commonly consumed marine fishes in Chile”, *Food Chemistry*, vol. 274, pp. 123-129, 2018. <https://doi.org/10.1016/j.foodchem.2018.08.113>
- [18] S. Sun, T. Ren, X. Li., *et al.*, “Polyunsaturated fatty acids synthesized by freshwater fish: A new insight to the roles of elovl2 and elovl5 in vivo”, *Biochemical and Biophysical Research Communications*, vol. 532, pp. 414-419, 2020. <https://doi.org/10.1016/j.bbrc.2020.08.074>
- [19] M.B. Behyar, M. Hasanzadeh, F. Seidi, *et al.*, “Sensing of amino acids: Critical role of nanomaterials for the efficient biomedical analysis”, *Microchemical Journal*, vol. 188, pp. 108452, 2023 <https://doi.org/10.1016/j.microc.2023.108452>

- [20] P. Shi, K. Liao, J. Xu, *et al.*, “Eicosapentaenoic acid mitigates palmitic acid-induced heat shock response, inflammation and repair processes in fish intestine”, *Fish & Shellfish Immunology*, vol. 124, pp. 362-371, 2022. <https://doi.org/10.1016/j.fsi.2022.04.011>
- [21] J. Wen, L. Zeng, Y. Xu, *et al.*, Proximate composition, amino acid and fatty acid composition of fish maws, *Natural Product Research: Formerly*, vol. 30, no. 2, pp. 214-217, 2015. <https://doi.org/10.1080/14786419.2015.1040790>
- [22] F.O.S. Duarte, F.G. de Paula, C.S. Prado, *et al.*, “Better fatty acids profile in fillets of Nile Tilapia (*Oreochromis niloticus*) supplemented with fish oil”, *Aquaculture*, Vol. 534, pp. 736241, 2021. <https://doi.org/10.1016/j.aquaculture.2020.736241>
- [23] K.L. Weaver, P. Ivester, J.A. Chilton, *et al.*, “The Content of Favorable and Unfavorable Polyunsaturated Fatty Acids Found in Commonly Eaten Fish”, *Journal of the American Dietetic Association*, vol. 108, No. 7, pp. 1178–1185, 2008. <https://doi.org/10.1016/j.jada.2008.04.023>
- [24] AOAC, “Official Methods of Analysis. 15th Edn., Association of Official Analytical Chemists (AOAC)”, Washington, DC., USA.1990. Reference Source
- [25] S.A. Cohen, “Amino acid analysis using pre-column derivatization with 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate”, *Protein Sequencing Protocols*, vol. 211, pp. 143-54, 2003. <https://doi.org/10.1385/1-59259-047-0:039>
- [26] M.A. Rajion, J.G. McLean, R.N. Cahill, “Essential fatty acid metabolism in the fetal and neonatal lamb”, *Australian Journal of Biological Sciences*, vol. 38, no. 1, pp. 33-40, 1985. [Essential fatty acids in the fetal and newborn lamb - PubMed \(nih.gov\)](https://pubmed.ncbi.nlm.nih.gov/2711111/)
- [27] T.L. Ulbricht, D.A. Southgate, “Coronary heart disease: Seven dietary factors”, *The Lancet*, Vol 338, no. 8773, pp. 985–992, 1991. [https://doi.org/10.1016/0140-6736\(91\)91846-M](https://doi.org/10.1016/0140-6736(91)91846-M)
- [28] D.B. Duncan, “Multiple ranges and multiple F tests”, *Biometrics*, vol. 11, pp. 1–42, 1955. <https://doi.org/10.2307/3001478>
- [29] R. Froese, “Cube law, condition factor and weight–length relationships: History, meta-analysis and recommendations”. *Journal of Applied Ichthyology*, vol. 22, no. 4, pp. 241-253, 2006. <https://doi.org/10.1111/j.1439-0426.2006.00805.x>
- [30] O.O. Famofofo, & W.O. Abdul, “Biometry, condition factors and length-weight relationships of sixteen fish species in Iwopin fresh-water ecotype of Lekki Lagoon, Ogun State, Southwest Nigeria”, *Heliyon*, vol. 6, no. 1, 2020. <https://doi.org/10.1016/j.heliyon.2019.e02957>
- [31] T.B. Bagenal, F.W. Tesch, “Age and Growth. In: Bagenal, T., Ed., Methods for Assessment of Fish Production in Fresh Waters”, *Blackwell Science Publications*, pp. 101-136, 1978.
- [32] E.O. Ahmed, M.E. Ali, A.A. Aziz, “Length-weight relationships and Condition factors of six fish species in Atbara River and Khashm el-girba Reservoir, Sudan”, *International Journal of Agriculture Sciences*, vol. 3, no.1, pp. 65–70, 2011. <http://dx.doi.org/10.9735/0975-3710.3.1.65-70>
- [33] N.C. Ujjania, M.P.S. Kohli, L.L. Sharma, “Length-weight relationship and condition factors of Indian major carps (*C. catla*, *L. rohita* and *C. mrigala*) in Mahi Bajaj Sagar, India”, *Research Journal of Biology*, vol. 2, no.1, pp. 30-36, 2012. <https://doi.org/10.21077/ijf.2017.64.special-issue.76263-27>

- [34] J.P. Bonjour, "Calcium and phosphate: A duet of ions playing for bone health", *Journal of the American College of Nutrition*, vol. 30, no. 5, pp. 438S-48S, 2011. <https://doi.org/10.1080/07315724.2011.10719988>
- [35] B.P. Mohanty, T.V. Sankar, S. Ganguly, *et al.*, "Micronutrient composition of 35 food fishes from India and their significance in human nutrition", *Biological Trace Element Research*, vol. 174, pp. 448-458, 2016. <https://doi.org/10.1007/s12011-016-0714-3>
- [36] F. Jabeen, A.A. Chudhry, "Chemical composition and fatty acid profiles of three freshwater species", *Food Chemistry*, vol. 125, pp. 991-996, 2011. <https://doi.org/10.1016/j.foodchem.2010.09.103>
- [37] A.A. Nurnadia, A. Azrina, I. Amin, "Proximate composition and energetic value of selected marine fish and shellfish from the west coast of peninsular Malaysia". *International Food Research Journal*. Vol 18, pp. 137-148, 2011. <https://core.ac.uk/download/pdf/153807129.pdf>
- [38] S.C. Dyall, L. Balas, N.G. Bazan, *et al.*, "Polyunsaturated fatty acids and fatty acid-derived lipid mediators: Recent advances in the understanding of their biosynthesis, structures and functions", *Progress in Lipid Research*. vol. 86, pp. 101165, 2022. <https://doi.org/10.1016/j.plipres.2022.101165>
- [39] N. Kim, U.D.Sohn, V. Mangannan, *et al.*, "Leukotrienes in Acetylcholine-Induced Contraction of Esophageal Circular Smooth Muscle in Experimental", *Gastroenterology*, vol. 112, no.5, pp. 1548-1558, 1997. [https://doi.org/10.1016/S0016-5085\(97\)70036-2](https://doi.org/10.1016/S0016-5085(97)70036-2)
- [40] D. Swanson, R. Block, S.A.Mousa, "Omega-3 Fatty Acids EPA and DHA: Health Benefits Throughout Life". *Advances in Nutrition*, vol. 3, no.1, pp. 1-7,2012. <https://doi.org/10.3945/an.111.000893>
- [41] R.P. Mason, S.C.R. Sherratt, L.H. Eckel, "Omega-3-fatty acids: Do they prevent cardiovascular disease?", *Best Practice & Research Clinical Endocrinology & Metabolism*, vol.37, no 3, 101681, 2023. <https://doi.org/10.1016/j.beem.2022.101681>.
- [42] J. M. Seddon, J. Cote, N. Davis, *et al.*, "Progression of age-related macular degeneration. Progression of age-related macular degeneration". *Arch Ophthalmol*. Vol. 121, no. 6, pp. 785-792, 2003. <https://doi.org/10.1001/archophth.121.6.785>
- [43] FAO/WHO: Fat and Oils in Human Nutrition: Report of a Joint Expert Consultation. FAO/WHO, Rome, Italy.
- [44] S.M. Pawar, S.R. Sonaware, "Fish muscle protein highest source of energy". *International Journal of Biodiversity and Conservation*. Vol. 5, no. 7, pp. 433-435, 2013. <https://doi.org/10.5897/IJBC12.043>
- [45] D. Tomé, C. Bos, "Lysine requirement through the human life cycle", *The Journal of Nutrition*, vol. 137, no.6, pp. 1642S-1645S, 2007. <https://doi.org/10.1093/jn/137.6.1642S>
- [46] M. Watford, "Glutamine and glutamate: Nonessential or essential amino acids?", *Animal Nutrition*, vol. 1, no. 3, pp. 119-122, 2015. <https://doi.org/10.1016/j.aninu.2015.08.008>
- [47] X. Li, S. Zheng, G. Wu, "Nutrition and metabolism of glutamate and glutamine in fish", *Amino Acids*. vol.52 , no.5, pp. 671-691, 2020. <https://doi.org/10.1007/s00726-020-02851-2>

- [48] M. Li, Y. Wu, L. Ye, “ The Role of Amino Acids in Endothelial Biology and Function”, *Cells*, vol.11, no.8, pp. 1372, 2022. <https://doi.org/10.3390/cells11081372>



Webmail
Univ. Bung Hatta

hafrizal syandri <syandri_1960@bunghatta.ac.id>

6639837: Revision requested

3 messages

International Journal of Food Science <ijfs@hindawi.com>
Reply-To: International Journal of Food Science <drajadurai@wiley.com>
To: Hafrijal Syandri <syandri_1960@bunghatta.ac.id>

Tue, Dec 12, 2023 at 12:15 AM

WILEY

Dear Dr. Hafrijal Syandri,

In order for your submission "Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security " to "International Journal of Food Science" to proceed further in the review process, you will need to revise your manuscript.

Reason & Details:

“

Dear authors,

The manuscript requires revision according to the reviewer's report. You can also refer to the following articles to enhance the discussion of the TI: <https://doi.org/10.3233/MNM-17195> ; <https://doi.org/10.5194/aab-63-471-2020> ; <https://doi.org/10.1155/2021/6633774>

When you have finished revising, follow the link below to submit your revision:

MANUSCRIPT DETAILS

Kind regards,
Deepanaa Rajadurai
International Journal of Food Science

Reviewer Comments:

“

Reviewer 1 Comments to the Author

Regarding my remark about C18:0, it is preferable to recalculate the TI without using stearic fatty acid (C18:0) as a proatherogenic fatty acid

This email was sent to syandri_1960@bunghatta.ac.id by John Wiley & Sons, Inc.

111 River Street, Hoboken, NJ 07030 USA. 877-762-2974.

For more information, please see our [privacy policy](#).

hafrizal syandri <syandri_1960@bunghatta.ac.id>
To: International Journal of Food Science <drajadurai@wiley.com>

Fri, Dec 15, 2023 at 9:26 AM

Dear
Deepanaa Rajadurai
International Journal of Food Science

Based on the reviewer's recommendation, the authors have recalculated the Thrombogenic Index (TI) without using stearic fatty acid (C18:0), and the TI value changed, highlighted in pink.

Best Regards

Hafrijal Syandri

[Quoted text hidden]

Phenom Emails <phenom.emails@hindawi.com>
Reply-To: Phenom Emails <phenom.emails@hindawi.com>
To: syandri_1960@bunghatta.ac.id

Fri, Dec 15, 2023 at 4:41 PM

Dear Dr. Syandri,

Thank you so much for your response.

Your revised manuscript has been submitted successfully in the system. When the manuscript reaches the final stage you will be notified.

Please do not hesitate to get back in touch if you have any further questions.

Best Regards,

Deepanaa

Deepanaa Rajadurai
Editorial Assistant



Hindawi

[Hindawi.com](https://www.hindawi.com) | [Twitter](#) | [Facebook](#) | [LinkedIn](#) | [YouTube](#)

[Quoted text hidden]

, hafrizal syandri <syandri_1960@bunghatta.ac.id> wrote:

[Quoted text hidden]

[Quoted text hidden]

The contents of this email and any attachments are confidential and intended only for the person or entity to whom it is addressed. If you are not the intended recipient, any use, review, distribution, reproduction or any action taken in reliance upon this message is strictly prohibited. If you received this message in error, please immediately notify the sender and permanently delete all copies of the email and any attachments.

[Click here for translations of this disclaimer.](#)

Research Article

Analysis of Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security

Azrita Azrita¹, Hafrijal Syandri^{1*}, Hazlina Ahamad Zakeri², Harfiandri Damanhuri¹, Netti Aryani³

¹Faculty of Fisheries and Marine Science, Universitas Bung Hatta, 25131, Padang-West Sumatra Province, Indonesia

²Faculty of Science and Marine Environment, Universiti Malaysia Terengganu, Malaysia.

³Faculty of Fisheries and Marine Science, Universitas Riau, 28293, Pekanbaru-Riau Province, Indonesia

*Corresponding author: syandri_1960@bunghatta.ac.id

Abstract

Fish have become an irreplaceable dietary source of animal protein, especially among households with low socioeconomic status in rural and urban areas of Indonesia. This study aimed to analyse the proximate composition, minerals, fatty acids, and amino acids of three local Bagridae fish species in Indonesia. The standard AOAC method was employed to examine the proximate composition of the carcass, and the analysis of amino acids and fatty acids was conducted through HPLC and GC-MS, respectively. The mineral content was determined using AAS. The nutrient composition results of *Hemibagrus nemurus*, *Hemibagrus wyckii*, and *Mystus nigriceps* revealed that the protein content was 24.26%, 22.57%, and 21.39% (%dry weight), whereas the total lipid content was 6.64%, 7.47% and 7.75%, respectively. Regarding mineral contents, the calcium levels ranged from 1.49–1.66 mg/g, iron levels from 28.35–40.36 µg/g, and zinc levels from 24.03–54.46 µg/g. Among the fatty acids, palmitic acid was the most abundant in all three species, accounting for 25.59–30.70% of the total fatty acids. Additionally, significant amounts of C18:1 (oleic acid), C18:0 (stearic acid), and C20:4 (arachidonic acid) were also detected as primary fatty acids. The calculated atherogenic index values in the three species of Bagridae fish ranged from 0.73–0.99, while the thrombogenic index values varied between 0.54–0.75. The predominant amino acids found in the three species of Bagridae fish were glutamic acid with their concentrations ranging from 9.10–24.34%. These results indicate that consuming the meat of these three freshwater Bagridae fish species caught in the wild does not pose any health risks to consumers. They can be considered a safe and suitable food source with good nutritional quality.

Keywords: Amino acids, Fatty acids, Indonesia, Kampar Kanan River, Minerals

Commented [S1]: The abstract has been added with the study objectives and revised with yellow highlighting

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

1. Introduction

Fish serve as a high-quality protein source that is essential for satisfying dietary protein requirements, especially in areas where other protein sources are few or difficult to obtain [1,2]. Their accessibility can aid in ensuring a steady supply of food, particularly for communities whose main source of income is fishing or aquaculture [3]. Fish can contribute to a more varied selection of protein sources in communities, resulting in a diet that is more nutritious and balanced. Dietary fish can treat nutrient deficits and enhance general nutritional health. Fish are generally considered safe and offer a lower risk of contamination with harmful substances compared to some land-based animal products. A reliable indicator of a fish's quality, nutritional value, physiological status, and environment is its chemical makeup, which is found in fish flesh [4].

Hemibagrus nemurus, locally known as "baung", *Hemibagrus wyckii*, known as "geso" and *Mystus nigriceps*, known as "ingir-ingir," are three species of catfish found in the Kampar Kanan River, Indonesia [5,6]. Due to their high market value, these species have been identified as significant economic resources for rural communities [7]. Therefore, the analysis of the nutritional quality of fish meat in the study area is very important to determine whether it is beneficial to human health.

Knowing the composition of fatty acids and amino acids in fish meat is an essential factor that should not be ignored [8]. This information provides valuable insight into the nutritional quality of fish and the values of more nutritious and healthy foods [9]. In addition, a better understanding of the composition of fatty acids and amino acids can assist in selecting a suitable feed [10], developing a balanced diet [11], and maintaining optimal fish meat nutrition [12].

Omega-3 polyunsaturated fatty acids (PUFAs), particularly docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA) derived from fish oil, exhibit anti-inflammatory and anti-gastrointestinal cancer properties, making them crucial as immune-boosting nutrients [13,14]. Studies have demonstrated that ω -3 and ω -6 PUFAs exhibit beneficial effects in the management of cardiovascular diseases and cancers [15,16]. The fatty acid composition of PUFAs can differ among different types of fish, including both freshwater and marine species

[17, 18]. Amino acids (AAs), such as cysteine, arginine, tyrosine, glycine, proline, and serine, play a vital role in illness and stress situations and in the prevention of inflammation and repair processes in fish intestines [19, 20].

Furthermore, knowledge of the atherogenicity index (AI) and thrombogenicity index (TI) of fatty acids in the three local freshwater fish species of Bagridae can contribute to improving dietary guidelines. The AI and TI can be used to develop recommendations aimed at improving heart health [21,22]. These recommendations may involve selecting fish with favourable fatty acid profiles, allowing individuals to choose the most suitable fish species based on their specific needs [17, 23]. Therefore, this study aims to examine the nutritional composition profile, mineral content, amino acids, and fatty acids, and AI and TI of three freshwater fish species, namely, *H. nemurus*, *H. wyckii*, and *M. nigriceps*. These three fish species are generally consumed by people who live around the Kampar Kanan River in Kampar Regency, Riau Province, Indonesia.

2. Material and methods

2.1. Animal material

In this study, a total of 27 Bagridae fish belonging to three distinct species (*H. nemurus*, *H. wyckii*, and *M. nigriceps*) were examined. The specimens were obtained from local fishers operating in the upper stretches of the Kampar Kanan River in Kampar Regency, Riau Province (Figure 1). Sampling took place between June and August 2022 and was verified by the Fisheries Department, Faculty of Fisheries and Marine Science, Universitas Bung Hatta, Indonesia. The samples were placed in plastic bags and transported to the laboratory in an insulated icebox. The samples were then grouped based on their respective sampling locations.

2.2. Biometry measurement

Upon reaching the laboratory within 10 hours, the fish samples were subjected to gutting and washing. Each specimen was weighed (TW) and measured to determine the standard length (SL) as well as the depth or maximum height (H). TW was measured using balance scale (OHAUS Model CT 6000 USA) with a precision of 0.01 g. The length measurement was taken from the tip of the mouth to the end of the upper lobe of the caudal fin, representing the total body length, using a metre ruler with an accuracy of 1 mm. The height measurement involved a vertical assessment of the body's maximum height measured using a

Commented [S2]: The author has been revised

Formatted: Highlight

Digital Sekhmet Sigma Vernier calliper with an accuracy of 1 mm. The condition factor (CF) was calculated using the formula $CF = (TW/SL^3) \times 100$.

2.3. Proximate, amino acid, and mineral content analysis

The proximate carcass composition was assessed following the AOAC standard methods [24]. The samples were dried to a constant weight at 105 °C. The crude protein content was analysed using the standard Kjeldahl method, which involves multiplying the nitrogen content by a factor of 6.25. The Soxhlet method with ether extraction was employed to analyse crude lipids. The ash content was obtained by incinerating the samples at 550 °C for 16 h. The amino acid composition was determined using a high-performance liquid chromatography (HPLC) system, which consisted of a Waters® 1525 binary HPLC pump, Waters® 717 autosamplers, and Waters® 2475 multi- λ fluorescence detector (with excitation at 250 nm and emission at 395 nm). The samples were hydrolysed in triplicate with 6 N hydrochloric acid for 24 h at 11 °C [25].

For mineral data composition (Na, Mg, Ca, K, P, Fe, Cu, Mn, and Zn), the ashed feed sample and carcass were dissolved in 1 ml of hydrochloric acid (35% v/v Suprapur®, Merck) and then filtered with cellulose filter paper (Watchman No 1, International Ltd; Maidstone, UK) and diluted to an appropriate concentration for each elemental mineral. P levels were analysed with a Perkin-Elmer AA spectrophotometer mod 3110 (Norwalk, CT, USA).

2.4. Fatty acid analysis

The fish meat was examined utilizing the gas chromatography–mass spectrometry (GC–MS) method. The method of Folch et al. (1957) modified by Rajion et al. [26] was employed to perform the total fat extraction. This involved using a solvent system consisting of chloroform and methanol at a 2:1 (v/v) ratio. The process of transmethylation was conducted using a solution of 14% methanolic boron trifluoride. The fatty acid composition of the meat was then analysed at the SIG Laboratory, Accredited Testing Laboratory-LP-184-IDN.

The nutritional quality of lipids (AI and TI) was calculated using the following equations [27]:

$$AI = \frac{[(C12:0 + 4 \times C14:0 + C16:0)]}{\sum MUFA + \sum n-6 + \sum n-3}$$

$$TI = \frac{[(C14:0 + C16:0)]}{[(0.5 \times \sum MUFA) + (0.5 \times \sum n-6) + (3 \times \sum n-3) + (\sum n-3 / \sum n-6)]}$$

where:

AI = Atherogenic index

TI = Thrombogenic index

C12:0 = lauric acid

C14:0 = myristic acid

C16:0 = palmitic acid

\sum MUFA = sum of the concentrations of all monosaturated fatty acids

\sum n-6 = sum of the concentrations of n-6 polyunsaturated fatty acids

\sum n-3 = sum of the concentrations of n-3 polyunsaturated fatty acids

2.5. Data analysis

Data analysis was performed using the SPSS 16.0 software package (SPSS; Chicago, IL). The homogeneity of variances was assessed using Levene's test. One-way ANOVA was carried out to determine the parameters of proximate and amino acid composition, including the composition of fatty acids and nutritional quality of lipids (AI and TI) for each species, followed by the post hoc Duncan's multiple range test [28]. The results are reported as the mean values \pm standard errors for each parameter.

Commented [S3]: We appreciate the reviewer's contribution regarding the perspective that C18:0 should not be considered a pro-atherogenic fatty acid. However, to ensure alignment with the Thrombogenic Index analysis, we would like to request further clarification as to whether the authors expected to remove C18:0 from our data

The author has recalculated the Thrombogenic Index (TI) without using stearic fatty acid (C18:0).

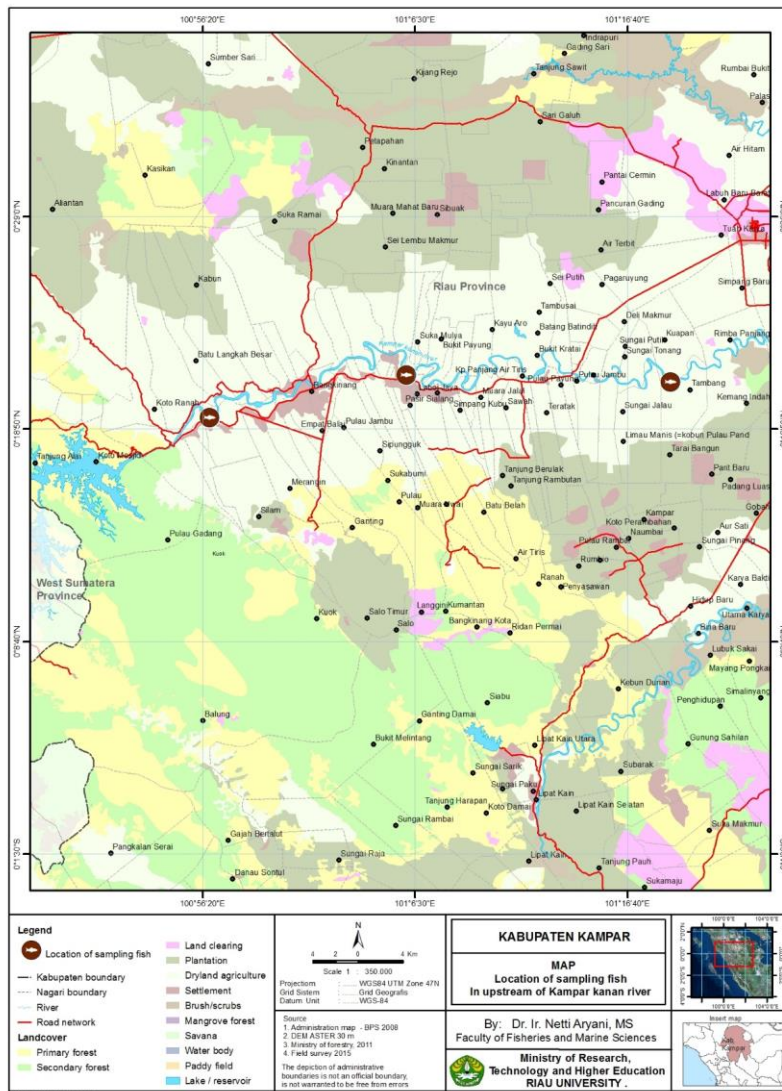


Figure 1. Map of Kampar Regency, Riau Province, and sampling locations of the three species of Bagridae fish.

Formatted: Highlight

Commented [S4]: Figure 1 has been revised

3. Results

Table 1 presents the average wet weight, standard length, height, and the results of meat nutritional composition analysis for three indigenous Indonesian freshwater Bagridae species. The moisture content (% wet weight) ranged from 82.40– 85.39%. Among the species examined, *M. nigriceps* exhibited the lowest protein content at 21.39%, while *H. nemurus* displayed the highest value at 24.26%. However, *M. nigriceps* had a higher mineral content, including of iron and zinc.

Table 2 summarizes the fatty acid composition (%total fatty acid) for the three Bagridae fish species. C16:0 was the predominant fatty acid in the three species of Bagridae fish, with percentages ranging from 25.09–30.71%. The additional prominent fatty acids found were C18:1, C18:0, and C22:6. *H. nemurus* exhibited a ω -6/ ω -3 ratio below 1 (0.82), while *H. wyckii* and *M. nigriceps* had ratios of 1.11 and 1.02, respectively. The obtained PUFA/saturated fatty acid (SFA) (P/S) ratio varied between 0.61 and 0.76. The AI values varied between 0.76 and 0.99 and the TI between 0.40 and 0.63 (**Table 3**).

Table 4 presents the amino acid composition (% of total protein) of the three Bagridae species. Glutamic acid was the dominant amino acid, ranging from 19.29–24.34%, followed by aspartic acid, which ranged from 9.21–11.27%. The lysine content was consistent, ranging from 9.67–9.87%. In *H. nemurus*, the levels of various amino acids ranged from 0.86–24.35%. Similarly, in *H. wyckii*, the amino acid levels ranged from 0.76–21.58%, while in *M. nigriceps*, the levels ranged from 0.74–19.29%.

Table 1. Results of biometric, proximate and mineral composition of three species of Bagridae fish

	Species		
	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
<i>Biometric measurements</i>			
Total weight (g)	389.99 ± 24.96 ^a	1,390.33 ± 168.29 ^b	17.57 ± 1.53 ^c
Standard length (cm)	28.16 ± 0.53 ^a	44.14 ± 1.98 ^b	10.29 ± 0.15 ^c
Height (cm)	8.44 ± 0.16 ^a	8.82 ± 0.39 ^b	2.57 ± 0.03 ^c
Condition factor	1.63 ± 0.08	1.58 ± 0.07	1.56 ± 0.08
<i>Proximate composition</i>			
Crude protein (% DW)	24.26 ± 0.87 ^a	22.57 ± 0.37 ^b	21.39 ± 0.15 ^c
Crude fat (% DW)	6.64 ± 0.03 ^a	7.47 ± 0.02 ^b	7.75 ± 0.40 ^c
Crude ash (% DW)	1.94 ± 0.02 ^a	2.30 ± 0.09 ^b	1.59 ± 0.02 ^c
Moisture (% WW)	82.40 ± 2.51	85.39 ± 2.36	83.75 ± 2.21

Commented [S5]: The author has recalculated the Thrombogenic Index (TI) without using stearic fatty acid (C18:0), and the TI value changed.

Formatted: Highlight

<i>Mineral composition</i>			
Macrominerals (mg/g)			
Sodium	0.99 ± 0.00 ^a	1.61 ± 0.01 ^b	1.61 ± 0.02 ^c
Magnesium	1.12 ± 0.01 ^a	0.57 ± 0.02 ^b	1.15 ± 0.01 ^c
Calcium	1.66 ± 0.00 ^a	1.55 ± 0.02 ^b	1.49 ± 0.04 ^c
Potassium	0.71 ± 0.00 ^a	0.55 ± 0.02 ^b	0.43 ± 0.01 ^c
Phosphorous	7.03 ± 0.03 ^a	2.74 ± 0.02 ^b	6.45 ± 0.07 ^c
Microminerals (µg/g)			
Iron	28.30 ± 0.11 ^a	28.73 ± 0.08 ^b	40.36 ± 0.55 ^c
Copper	8.93 ± 0.03 ^a	7.46 ± 0.09 ^b	6.21 ± 0.32 ^c
Manganese	1.64 ± 0.02 ^a	2.64 ± 0.02 ^b	2.84 ± 0.02 ^c
Zinc	24.03 ± 0.45 ^a	24.61 ± 0.19 ^b	54.46 ± 0.17 ^c

Values are mean % ± SE; *Values in the same row followed by different letters are significantly different (p<0.05).

Table 2. Fatty acid composition of three species of Bagridae fish

Fatty acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
C12:0 (lauric acid)	1.97 ± 0.05 ^a	3.94 ± 0.01 ^b	4.27 ± 0.01 ^c
C14:0 (Myristic acid)	2.38 ± 0.03 ^a	2.72 ± 0.08 ^b	2.89 ± 0.01 ^c
C16:0 (Palmitic acid)	27.23 ± 0.06 ^a	30.70 ± 0.06 ^b	25.59 ± 0.01 ^c
C18:0 (Stearic acid)	16.17 ± 0.02 ^a	13.54 ± 0.02 ^b	10.64 ± 0.01 ^c
C20:0 (Arachidic acid)	0.23 ± 0.01 ^a	0.14 ± 0.01 ^b	0.37 ± 0.01 ^c
C16:1 (Palmitoleic acid)	1.78 ± 0.00 ^a	1.27 ± 0.04 ^b	4.93 ± 0.01 ^c
C18:1 (Oleic acid)	16.86 ± 0.01 ^a	16.18 ± 0.01 ^b	16.97 ± 0.01 ^c
C18:2 (Linoleic acid)	3.24 ± 0.02 ^a	3.84 ± 0.33 ^b	4.55 ± 0.001 ^c
C18:3 (Linolenic acid)	1.12 ± 0.01 ^a	0.82 ± 0.01 ^b	1.57 ± 0.000 ^c
C20:4 (Arachidonic acid)	12.13 ± 0.01 ^a	12.49 ± 0.09 ^b	12.41 ± 0.02 ^c
C20:5 (Eicosapentaenoic acid; EPA)	2.06 ± 0.08 ^a	2.34 ± 0.04 ^b	1.35 ± 0.01 ^c
C22:6 (Docosahexaenoic acid; DHA)	15.65 ± 0.01 ^a	11.56 ± 0.04 ^b	13.64 ± 0.02 ^c

Values are mean % ± SE; *Values in the same row with a different superscript are significantly different (p<0.05).

Table 3. Fatty acid ω-6/ω-3 and polyunsaturated/saturated fatty acid ratio of three species of Bagridae fish

	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
ω-6/ω-3 ratio	0.82 ± 0.00 ^a	1.11 ± 0.02 ^b	1.02 ± 0.01 ^c

P/S ratio	0.69 ± 0.01 ^a	0.61 ± 0.06 ^b	0.76 ± 0.00 ^c
AI	0.87 ± 0.01 ^a	0.99 ± 0.06 ^b	0.73 ± 0.00 ^c
TI	0.40 ± 0.01 ^a	0.63 ± 0.01 ^b	0.43 ± 0.00 ^c

P/S, polyunsaturated/saturated fatty acid; AI, atherogenic index; TI, thrombogenic index
 Values are mean % ± SE; *Values in the same row followed by different letters are significantly different (p<0.05).

Commented [S6]: The author has recalculated the Thrombogenic Index (TI) without using stearic fatty acid (C18:0), and the TI value changed.

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Table 4. Amino acid composition of three species of Bagridae fish

Amino acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
Aspartic acid	10.39 ± 0.01 ^a	9.61 ± 0.04 ^b	11.28 ± 0.02 ^c
Glutamic acid	24.34 ± 0.00 ^a	21.58 ± 0.41 ^b	19.29 ± 0.03 ^c
Serine	3.74 ± 0.00 ^a	5.40 ± 0.03 ^b	7.50 ± 0.06 ^c
Glycine	4.02 ± 0.00 ^a	5.77 ± 0.04 ^b	4.69 ± 0.02 ^c
Histidine	2.39 ± 0.00 ^a	2.90 ± 0.03 ^b	2.21 ± 0.02 ^c
Arginine	6.17 ± 0.01 ^a	8.27 ± 0.03 ^b	6.22 ± 0.03 ^c
Threonine	4.49 ± 0.01 ^a	4.19 ± 0.03 ^b	4.39 ± 0.04 ^c
Alanine	7.03 ± 0.00 ^a	6.04 ± 0.02 ^b	7.89 ± 0.01 ^c
Proline	2.40 ± 0.00 ^a	2.71 ± 0.01 ^b	2.21 ± 0.02 ^c
Tyrosine	3.91 ± 0.01 ^a	3.29 ± 0.06 ^b	3.82 ± 0.04 ^c
Valine	3.73 ± 0.00 ^a	4.02 ± 0.01 ^b	3.85 ± 0.04 ^c
Methionine	3.42 ± 0.00 ^a	1.96 ± 0.02 ^b	1.27 ± 0.03 ^c
Cystine	0.87 ± 0.00 ^a	0.76 ± 0.02 ^b	0.75 ± 0.03 ^c
Isoleucine	3.82 ± 0.001 ^a	3.96 ± 0.003 ^b	3.66 ± 0.002 ^c
Leucine	6.15 ± 0.000 ^a	6.69 ± 0.002 ^b	6.28 ± 0.004 ^c
Phenylalanine	5.13 ± 0.000 ^a	6.03 ± 0.005 ^b	5.18 ± 0.027 ^c
Lysine	9.61 ± 0.001 ^a	9.82 ± 0.026 ^b	9.87 ± 0.003 ^c

Values are mean % ± SE; *Values in the same row followed by different letters are significantly different (p<0.05).

4. Discussion

The results of this study provide important new information about the nutritional makeup of the studied species of Bagridae fish. The biometric measures of the fish yielded important information about their physical traits and health. Researchers are able to determine the average weight of fish at various lengths by examining the link between length and weight for a particular species. This knowledge is essential for understanding the trends in fish population growth, identifying potential shifts in growth rates over time, and assessing the general health and welfare of fish [29]. A variety of factors, including sex, age, maturity level, size, stomach fullness level, sampling strategies, sample sizes, and environmental circumstances, have an impact on fish health and the parameters that determine length–weight connections in fish [30]. However, none of these factors were considered in our study. In general, according to the theory put forth by [Bagenal and Tesch \[31\]](#), heavier fish of a given length are in better physiological condition. The condition factor is also used in fisheries science to compare the health, fatness, and condition of fish [32]. A condition factor of > 1 is favourable, indicating a good feed nutrition level and suitable living habitat [33]. Thus, based on the findings in this study, the Kampar Kanan River provides a suitable habitat for the fish, ensuring their accessibility and availability for community consumption.

The fish samples exhibited good nutritional profiles, with high amounts of proteins and lipids as well as moisture content contributing to their overall nutritional value, according to the results of proximate analysis and mineral composition (see Table 1). These results imply that the fish species studied provide a possible source of nutrients to solve issues with food security. The study of the mineral composition revealed the existence of critical minerals such as phosphorus (P), calcium (Ca), magnesium (Mg), iron (Fe), and zinc (Zn) in the fish samples in addition to proteins and lipids. All three Bagridae fish significantly differed from each other in terms of their crude ash content and mineral profile (see Table 1). However, in general, P and Ca are the two highest minerals found in all fish. Both of these macro minerals are good for human bone and tooth health, as key components of hydroxyapatite, the mineral matrix that provides strength and structure to bones and teeth [34].

In the current study, the meat of three species of Bagridae fish from the Kampar Kanan River in Indonesia was found to contain an abundance of microminerals such as Fe and Zn. *M. nigriceps* has been reported to exhibit higher mineral content, including of Fe and Zn. This species is categorized as a small indigenous fish (SIF) with a maximum standard length

of approximately 10.29 cm. According to [35], SIFs are generally known to be rich in Ca, Fe, and Mn.

In this study, all three Bagridae fish had a total fat content of approximately 7–8% by weight, with a higher water content ranging from 82.40–85.39% (Table 1). These values are similar to those found in wild and cultured sea bass *Dicentrarchus labrax* [36]. Additionally, the fish species can be grouped into different categories, including high-fat fish (>8%), medium-fat fish (4–8%), low-fat fish (2–4%), and lean fish (<2%) [37]. According to the classification established by [37], the three indigenous Bagridae fish species are characterized as having a moderate fat content level, which is reflected by the high levels of DHA and EPA in the meat of *H. nemurus*, *H. wyckii*, and *M. nigriceps* (Table 2). Therefore, we recommend that consuming the meat of these three fish species can indirectly boost food security in rural and urban communities.

All three Bagridae fish species analysed in the study contained arachidonic acid (C20:4), at 12.13% for *H. nemurus*, 12.49% for *H. wyckii*, and 12.41% for *M. nigriceps*. Arachidonic acid serves as a precursor for synthesizing prostaglandins and leukotrienes [38]. Leukotrienes are crucial in allergic responses, inducing smooth muscle contraction and increasing vascular permeability [39]. Additionally, all three species also contained EPA ranging from 1.35–2.34% and DHA ranging from 11.56–15.65%. The increased consumption of omega-3 fatty acids, such as EPA and DHA, has been linked to a decreased risk of pain development in older individuals and to the prevention of Alzheimer's disease [40,41].

As determined by the current study, the ω -6/ ω -3 ratios of the three Bagridae fish species ranged from 1:0.8 to 1:0.97. It was found that all three fish species had ω -6/ ω -3 ratios within the recommended range for a typical Indonesian diet. In contrast, a typical Western diet is characterized by a high ω -6/ ω -3 ratio of 10:1 to 20:1, while a Mediterranean diet has a ratio of 4:1 to 5:1 [42]. Additionally, the P/S ratio analysis showed that the three Bagridae fish species are sources of PUFAs that meet the requirements for food safety.

The Food and Agriculture Organization (FAO) and the World Health Organization (WHO) have recommended AI and TI values ranging from 0.4 to 0.5 for promoting human health [43]. In the current study, the observed AI values in the three freshwater species of Bagridae ranged from 0.73 to 0.99, while the TI values varied between 0.40 and 0.63. This finding is associated with a notable disparity in the values of SFAs among the three Bagridae

Commented [S7]: The author has recalculated the Thrombogenic Index (TI) without using stearic fatty acid (C18:0), and the TI value changed.

Formatted: Highlight

fish species. Based on these findings, we confirm that consuming the meat of the three freshwater species of Bagridae fish caught in the wild does not endanger consumer health and can serve as a suitable source of food safety and nutritional quality.

Fish with a high crude protein content, as shown by the three studied Bagridae fish (Table 1), are regarded as being nutritionally useful for human health. The protein and amino acid contents can directly indicate the nutritional quality of meat [44]. As shown in Table 4, *H. wyckii* had a higher content of essential amino acids (EAAs) than *H. nemurus* and *M. nigriceps*. Among the EAAs, lysine (Lys) was found to be present in the highest amount in all three species. Lys is necessary for tissue growth and repair, reduced stress-induced anxiety, and the conversion of fatty acids into energy [45-47].

More than 60% of the amino acid composition in all three Brigade fish studied comes from nonessential amino acids (NEAAs), particularly glutamic acid (Table 4). Glutamic acid and its amine, glutamine, are highly abundant amino acids found in fish in the free and protein-bound forms [48]. Other amino acids found at abundant levels in the three fish also play important physiological and biochemical functions. The NEAA arginine, for example, is involved in protein synthesis, creatine formation, and various other biochemical processes within the body. The EAAs leucine, phenylalanine, and methionine play important roles in the synthesis of nitric oxide, a molecule that can expand blood vessels [48].

4. Conclusions

The present study provides insightful information about the fatty acid and amino acid compositions, biometric measurements, proximate analysis, and mineral composition of three local freshwater species of Indonesian Bagridae fish. The findings show their potential as a nutrient-dense, sustainable food source that can support efforts to increase food security. These fish species might be incorporated into the local diet to increase nutrient intake and alleviate nutritional inadequacies, which would ultimately improve Indonesia's overall food security status.

Data Availability

The data utilized in this research has not been previously released or published in any form. The datasets employed and/or analysed during the present study can be obtained by contacting the corresponding author.

Conflicts of Interest

The authors confirm that they do not have any conflicting interests.

Acknowledgements

The authors acknowledge and express their gratitude for the financial support received from the Ministry of Education, Culture, Research, and Technology of the Republic of Indonesia, specifically through grant no. 076/E5/PG. 02.00. PT/2022, which funded the research. The authors affirm that there is no conflict of interest associated with the utilization of these research funds. Furthermore, the authors extend their appreciation to the students and fishers involved in the research conducted in the upper reaches of the Kampar Kanan River, Indonesia.

References

- [1] A.E. Hasselberg, I.Aakre, J. Scholtens, R. Avera, J. Kolding, M.S. Bank, A. Atter, M. Kjellevold., "Fish for food and nutrition security in Ghana: Challenges and opportunities," *Global Food Security*, vol. 26, pp. 100380, 2020. <https://doi.org/10.1016/j.gfs.2020.100380>
- [2] N.Tran, U.P, Rodriguez, C.Y.Chan, Y. M. Auang, L. Chu, A.H.Md.S. Islam., "Future scenarios of fish supply and demand for food and nutrition security in Bangladesh: An analysis with the AsiaFish model", *Aquaculture*, vol.568, pp.739288, 2023. <https://doi.org/10.1016/j.aquaculture.2023.739288>
- [3] C.Z. Levkoe, K. Lowitt, C. Nelson. "Fish as food: Exploring a food sovereignty approach to small-scale fisheries". *Marine Policy*, vol. 85, pp.65-70, 2017. <https://doi.org/10.1016/j.marpol.2017.08.018>
- [4] M. Ravichandran, N.S. Hettiarachchy, V. Ganesh, S. Ricke., "Enhancement of antimicrobial activities of naturally occurring phenolic compounds by nanoscale delivery against listeria monocytogenes, escherichia coli O157:H7 and salmonella typhimurium in broth and chicken meat system". *Journal of Food Safety*, vol. 31, no. 4, pp. 462-471,2011. <https://doi.org/10.1111/j.1745-4565.2011.00322.x>
- [5] N. Aryani, I. Suharman, Azrita, H. Syandri., "Diversity and distribution of fish fauna of upstream and downstream areas at Koto Panjang Reservoir, Riau Province, Indonesia". *F1000 Research*, vol. 8, 1435.pp.1-16, 2020. <https://f1000research.com/articles/8-1435/v2>
- [6] Syafrialdi, Dahelmi, D.I. Roesma, H. Syandri., "Length-weight relationships and condition factor of Two-Spot Catfish (*Mystus nigriceps* [Valenciennes, 1840]) Pisces, Bagridae], from Kampar Kanan River and Kampar Kiri River in Indonesia". *Pakistan Journal of Biological Science*, vol. 23, no. 12, pp. 1636-1642, 2020. <https://scialert.net/fulltext/?doi=pjbs.2020.1636.1642>
- [7] N. Aryani, "Native species in Kampar Kanan River, Riau Province Indonesia". *International Journal of Fisheries and Aquatic Studies*, vol.2, no.5, pp. 213-217. 2015. <https://www.fisheriesjournal.com/archives/2015/vol2issue5/PartD/2-5-63.pdf>

Formatted: Highlight

Formatted: Highlight

Formatted: Font: Italic

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Font: Italic

Formatted: Font: Italic

Formatted: Font: Italic

Formatted: Font: Italic

- [8] Azrita, H. Syandri, N. Aryani, A. Mardiah, I. Suharman., “The utilization of new products formulated from water coconut, palm sap sugar, and fungus to increase nutritional feed quality, feed efficiency, growth, and carcass of gurami sago (*Osphronemus goramy* Lacepède, 1801) juvenile”. *F1000 Research*, vol. 10, 1121, pp. 1-16, 2021. <https://f1000research.com/articles/10-1121/v1>
- [9] C. Bene, R. Arthur, H. Norbury, E.H. Allison, M. Beveridge, S. Bush, L. Campling, W. Leschen, D. Little, D. Squires, S.H. Thilsted, M. Troell, M. Williams., “Contribution of Fisheries and Aquaculture to Food Security and Poverty Reduction: Assessing the Current Evidence”. *World Development*, vol. 79, pp. 177–196. 2016. <http://dx.doi.org/10.1016/j.worlddev.2015.11.007>
- [10] N. Aryani, I. Suharman, S. Hasibuan, N. Asia, H. Syandri., “Fatty acid composition on diet and carcasses, growth, body indices and profile serum of Asian redtail catfish (*Hemibagrus nemurus*) fed a diet containing different levels of EPA and DHA”. *F1000Research*, vol. 11:1409, pp. 1-23, 2023. <https://doi.org/10.12688/f1000research.126487.2>
- [11] E.N. Kingsley, O.O. Cyril, O.I. Patience, “Potential contribution of selected wild fish species to the minerals intake of Pregnant and Lactating Women, Children and Adults in Rural Riverine Communities of Edo State: Insights and Outcomes”. *Measurement: Food*, vol. 8, 100063, pp. 1-7, 2022. <https://doi.org/10.1016/j.meafoo.2022.100063>
- [12] A.A. Sausa, L. Nora, D.L.A. Lopes, T.G. Petrolli, V.J.M. Vurlan, R. Wagner, C.M. Giacomelli, M.D. Baldisera, A.S. Da Silva., Vegetable choline in feed for Nile tilapia (*Oreochromis niloticus*) raised in a biofloc technology system (BFT): Biofloc composition, chemical composition, and fatty acid profiles in meat”. *Aquaculture*, vol. 545, pp. 737174, 2021. <https://doi.org/10.1016/j.aquaculture.2021.737174>
- [13] A.M. Eltweri, A.L. Thomas, M. Metcalfe, P.C. Calder, A.R. Dennison, D.J. Bowrey., Potential applications of fish oils rich in omega-3 polyunsaturated fatty acids in the management of gastrointestinal cancer. *Clinical Nutrition*. vol. 36, no. 1, pp. 65-78. 2017. <https://doi.org/10.1016/j.clnu.2016.01.007>
- [14] P.C. Haycock, M.C. Borges, K. Burrows, L.N. Letmetre, S. Burgess, N.K. Khankari, K.K. Tslidis, et al., “The association between genetically elevated polyunsaturated fatty acids and risk of cancer”. *eBiomedicine*. vol.91, pp. 104510, 2023. <https://doi.org/10.1016/j.ebiom.2023.104510>
- [15] E.K. Aglago, I. Huybrechts, N. Murphy, C. Casagrande, G. Nicolas et al., “Consumption of Fish and Long-chain n-3 Polyunsaturated Fatty Acids Is Associated With Reduced Risk of Colorectal Cancer in a Large European Cohort”. *Clinical Gastroenterology and Hepatology*. vol. 18, no. 3, pp. 654-666.e6. 2020. <https://doi.org/10.1016/j.cgh.2019.06.031>
- [16] H. Jiang, X. Shi, T.Y. Fan, D. Wang, B. Li, J. Zhou, C. Pei, L. Ma., “Dietary omega-3 polyunsaturated fatty acids and fish intake and risk of age-related macular degeneration” . *Clinical Nutrition*. vol.40, no. 12, pp: 5662-5673. 2021. [https://www.clinicalnutritionjournal.com/article/S0261-5614\(21\)00474-X/fulltext](https://www.clinicalnutritionjournal.com/article/S0261-5614(21)00474-X/fulltext)
- [17] M.A. Rincón-Cervera, N.V. González-Barriga, R. Valenzuela, S. Lopez-Arana, J. Romero, A. Valenzuela., “Profile and distribution of fatty acids in edible parts of commonly consumed marine fishes in Chile”, *Food Chemistry*, vol. 274, pp. 123-129, 2018. <https://doi.org/10.1016/j.foodchem.2018.08.113>

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Font: Italic

Formatted: Font: Italic

Formatted: Highlight

Formatted: Font: Italic

Formatted: Highlight

Formatted: Highlight

Commented [S8]: The authors of this article are 35 researchers.

Commented [S9]: The authors of this article are 47 researchers.

Formatted: Highlight

Formatted: Font: Not Italic, Highlight

Formatted: Font: Not Italic

Formatted: Highlight

Formatted: Highlight

- [18] S. Sun, T. Ren, X. Li., X. Cao, J. Gao., “Polyunsaturated fatty acids synthesized by freshwater fish: A new insight to the roles of elov12 and elov15 in vivo”, *Biochemical and Biophysical Research Communications*, vol. 532, pp. 414-419, 2020. <https://doi.org/10.1016/j.bbrc.2020.08.074>
- [19] M.B. Behyar, M. Hasanzadeh, F. Seidi, N. Sadjou ., “Sensing of amino acids: Critical role of nanomaterials for the efficient biomedical analysis”, *Microchemical Journal*, vol. 188, pp. 108452, 2023 <https://doi.org/10.1016/j.microc.2023.108452>
- [20] P. Shi, K. Liao, J. Xu, Y. Wang, S. Xu, X. Yan., “Eicosapentaenoic acid mitigates palmitic acid-induced heat shock response, inflammation and repair processes in fish intestine”, *Fish & Shellfish Immunology*, vol. 124, pp. 362-371, 2022. <https://doi.org/10.1016/j.fsi.2022.04.011>
- [21] J. Wen, L. Zeng, Y. Xu, Y. Sun, Z. Chen, S. Fan., Proximate composition, amino acid and fatty acid composition of fish maws, *Natural Product Research: Formerly*, vol. 30, no. 2, pp. 214-217, 2015. <https://doi.org/10.1080/14786419.2015.1040790>
- [22] F.O.S. Duarte, F.G. de Paula, C.S. Prado, R.R. dos Santos, C.S. Minafra-Rezende, C. Gebara, M.E. Lage., “Better fatty acids profile in fillets of Nile Tilapia (*Oreochromis niloticus*) supplemented with fish oil”, *Aquaculture*, Vol. 534, pp. 736241, 2021. <https://doi.org/10.1016/j.aquaculture.2020.736241>
- [23] K.L. Weaver, P. Ivester, J.A.Chilton, M.D. Wilson, P. Pandey, F.H. Chilton, “The Content of Favorable and Unfavorable Polyunsaturated Fatty Acids Found in Commonly Eaten Fish”, *Journal of the American Dietetic Association*, vol. 108, No. 7 , pp. 1178–1185, 2008. <https://doi.org/10.1016/j.jada.2008.04.023>
- [24] AOAC, “Official Methods of Analysis. 15th Edn., Association of Official Analytical Chemists (AOAC)”, Washington, DC., USA.1990. Reference Source
- [25] S.A.Cohen, “Amino acid analysis using pre-column derivatization with6-aminoquinolyl-N-hydroxysuccinimidyl carbamate”, *Protein Sequencing Protocols*, vol. 211, pp. 143-54, 2003. <https://doi.org/10.1385/1-59259-047-0:039>
- [26] M.A. Rajion, J.G. McLean, R.N. Cahill, “Essential fatty acid metabolism in the fetal and neonatal lamb”, *Australian Journal of Biological Sciences*, vol. 38, no. 1, pp. 33-40, 1985. [Essential fatty acids in the fetal and newborn lamb - PubMed \(nih.gov\)](https://pubmed.ncbi.nlm.nih.gov/1985/1985.Essential+fatty+acids+in+the+fetal+and+newborn+lamb/)
- [27] T.L. Ulbricht, D.A. Southgate, “Coronary heart disease: Seven dietary factors”, *The Lancet*, Vol 338, no. 8773, pp. 985–992, 1991. [https://doi.org/10.1016/0140-6736\(91\)91846-M](https://doi.org/10.1016/0140-6736(91)91846-M)
- [28] D.B. Duncan, “Multiple ranges and multiple F tests”, *Biometrics*, vol. 11, pp. 1–42, 1955. <https://doi.org/10.2307/3001478>
- [29] R. Froese, “Cube law, condition factor and weight–length relationships: History, meta-analysis and recommendations”. *Journal of Applied Ichthyology*, vol. 22, no. 4, pp. 241-253, 2006. <https://doi.org/10.1111/j.1439-0426.2006.00805.x>
- [30] O.O.Famoofo, & W.O. Abdul, “Biometry, condition factors and length-weight relationships of sixteen fish species in Iwopin fresh-water ecotype of Lekki Lagoon, Ogun State, Southwest Nigeria”, *Heliyon*, vol. 6, no. 1, pp. e02957, 2020. <https://doi.org/10.1016/j.heliyon.2019.e02957>
- [31] T.B. Bagenal, F.W. Tesch, “Age and Growth. In: Bagenal, T., Ed., Methods for Assessment of Fish Production in Fresh Waters”, *Blackwell Science Publications*, pp. 101-136,1978.

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

- [32] E.O. Ahmed, M.E. Ali, A.A. Aziz, "Length-weight relationships and Condition factors of six fish species in Atbara River and Khashm el-girba Reservoir, Sudan", *International Journal of Agriculture Sciences*, vol. 3, no.1, pp. 65–70, 2011. <http://dx.doi.org/10.9735/0975-3710.3.1.65-70>
- [33] N.C. Ujjania, M.P.S. Kohli, L.L. Sharma, "Length-weight relationship and condition factors of Indian major carps (*C. catla*, *L. rohita* and *C. mrigala*) in Mahi Bajaj Sagar, India", *Research Journal of Biology*, vol. 2, no.1, pp. 30-36, 2012. <https://doi.org/10.21077/rjf.2017.64.special-issue.76263-27>
- [34] J.P. Bonjour, "Calcium and phosphate: A duet of ions playing for bone health", *Journal of the American College of Nutrition*, vol. 30, no. 5, pp. 438S-48S, 2011. <https://doi.org/10.1080/07315724.2011.10719988>
- [35] B.P. Mohanty, T.V. Sankar, S. Ganguly, A. Mahanty, R. Anandan, K. Chakraborty et al., "Micronutrient composition of 35 food fishes from India and their significance in human nutrition", *Biological Trace Element Research*, vol. 174, pp. 448-458, 2016. <https://doi.org/10.1007/s12011-016-0714-3>
- [36] F. Jabeen, A.A. Chudhry, "Chemical composition and fatty acid profiles of three freshwater species", *Food Chemistry*, vol. 125, pp. 991-996, 2011. <https://doi.org/10.1016/j.foodchem.2010.09.103>
- [37] A.A. Nurnadia, A. Azrina, I. Amin, "Proximate composition and energetic value of selected marine fish and shellfish from the west coast of peninsular Malaysia". *International Food Research Journal*. Vol 18, pp. 137-148, 2011. <https://core.ac.uk/download/pdf/153807129.pdf>
- [38] S.C. Dyall, L. Balas, N.G. Bazan, J.T. Brena, N. Chiang, F.C. Sauza, J. Dalli, T. Durand, J. Galano, P.J. Lein, C.N. Serhan, A.Y. Taha., "Polyunsaturated fatty acids and fatty acid-derived lipid mediators: Recent advances in the understanding of their biosynthesis, structures and functions", *Progress in Lipid Research*. vol. 86, pp. 101165, 2022. <https://doi.org/10.1016/j.plipres.2022.101165>
- [39] N. Kim, U.D.Sohn, V. Mangannan, H. Rich, M.K. Jain, J. Behar, P. Biancani., "Leukotrienes in Acetylcholine-Induced Contraction of Esophageal Circular Smooth Muscle in Experimental", *Gastroenterology*, vol. 112, no.5, pp. 1548–1558, 1997. [https://doi.org/10.1016/S0016-5085\(97\)70036-2](https://doi.org/10.1016/S0016-5085(97)70036-2)
- [40] D. Swanson, R. Block, S.A.Mousa, "Omega-3 Fatty Acids EPA and DHA: Health Benefits Throughout Life". *Advances in Nutrition*, vol. 3, no.1, pp. 1-7,2012. <https://doi.org/10.3945/an.111.000893>
- [41] R.P. Mason, S.C.R. Sherratt, L.H. Eckel, "Omega-3-fatty acids: Do they prevent cardiovascular disease?", *Best Practice & Research Clinical Endocrinology & Metabolism*, vol.37, no 3, 101681, 2023. <https://doi.org/10.1016/j.beem.2022.101681>.
- [42] J. M. Seddon, J. Cote, N. Davis, B. Rosner., "Progression of age-related macular degeneration. Progression of age-related macular degeneration". *Arch Ophthalmol*. Vol. 121, no. 6, pp. 785-792, 2003. <https://doi.org/10.1001/archophth.121.6.785>
- [43] FAO/WHO: Fat and Oils in Human Nutrition: Report of a Joint Expert Consultation. FAO/WHO, Rome, Italy.
- [44] S.M. Pawar, S.R. Sonaware, "Fish muscle protein highest source of energy". *International Journal of Biodiversity and Conservation*. Vol. 5, no. 7, pp. 433-435, 2013. <https://doi.org/10.5897/IJBC12.043>

Formatted: Font: Italic

Formatted: Font: Italic

Formatted: Font: Italic

Commented [S10]: The authors of this article are 20 researchers.

Commented [S11]:

Formatted: Highlight

Formatted: Font: Not Italic

Formatted: Highlight

Formatted: Highlight

Formatted: Highlight

- [45] D. Tomé, C. Bos, “Lysine requirement through the human life cycle”, *The Journal of Nutrition*, vol. 137, no.6, pp. 1642S–1645S, 2007. <https://doi.org/10.1093/jn/137.6.1642S>
- [46] M. Watford, “Glutamine and glutamate: Nonessential or essential amino acids?”, *Animal Nutrition*, vol. 1, no. 3, pp. 119-122, 2015. <https://doi.org/10.1016/j.aninu.2015.08.008>
- [47] X. Li, S. Zheng, G. Wu, “Nutrition and metabolism of glutamate and glutamine in fish”, *Amino Acids*. vol.52, no.5, pp. 671-691, 2020. <https://doi.org/10.1007/s00726-020-02851-2>
- [48] M. Li, Y. Wu, L. Ye, “The Role of Amino Acids in Endothelial Biology and Function”, *Cells*, vol.11, no.8, pp. 1372, 2022. <https://doi.org/10.3390/cells11081372>



Webmail
Univ. Bung Hatta

hafrizal syandri <syandri_1960@bunghatta.ac.id>

[6639837] - Revised Reference List Needed

3 messages

Chandrika Harikrishnan <chandrika.harikrishnan@hindawi.com>

Sat, Dec 16, 2023 at 11:53 AM

Reply-To: Chandrika Harikrishnan <chandrika.harikrishnan@hindawi.com>

To: syandri_1960@bunghatta.ac.id

Dear Dr. Syandri,

This is regarding the manuscript "**6639837**" titled "**Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security**" has been submitted to "**International Journal of Food Science**".

During our routine quality checks, we found that the manuscript consists of many et al references. Kindly update the reference section and provide a detailed reference list that includes all authors names (***explain et al.***), ***title, journal name, and published year*** in the final manuscript and send us the file in document format via email in order to proceed further.

An early reply would be appreciated.

Best regards,
Chandrika

Chandrika Harikrishnan
Quality Checker



Hindawi

[Hindawi.com](https://www.hindawi.com) | [Twitter](#) | [FaceBook](#) | [LinkedIn](#) | [YouTube](#)

hafrizal syandri <syandri_1960@bunghatta.ac.id>

Sun, Dec 17, 2023 at 5:51 PM

To: Chandrika Harikrishnan <chandrika.harikrishnan@hindawi.com>

Dear
Chandrika Harikrishnan
Quality Checker of International Journal of Food Science

Thank you for your email on December 16, 2023; The authors have revised the manuscript consisting of many et al on the references (attached).

Best Regards
Hafrijal Syandri
[Quoted text hidden]



Manuscript Int. J. Food Sci_Dec 16, 2023_ Revised 5.doc
1056K

Chandrika Harikrishnan <chandrika.harikrishnan@hindawi.com>
Reply-To: Chandrika Harikrishnan <chandrika.harikrishnan@hindawi.com>
To: syandri_1960@bunghatta.ac.id

Mon, Dec 18, 2023 at 3:18 PM

Dear Dr. Syandri,

Thank you for your email. We will check this and get back to you incase of any queries.

Best regards,
Chandrika

Chandrika Harikrishnan
Quality Checker



Hindawi

[Hindawi.com](https://www.hindawi.com) | [Twitter](https://twitter.com/hindawipub) | [FaceBook](https://www.facebook.com/hindawipub) | [LinkedIn](https://www.linkedin.com/company/hindawi) | [YouTube](https://www.youtube.com/channel/UC8vRtN9iIbQ1mK1mK1mK1mK1)

On Sun, 17 Dec at 10:56 AM , hafrizal syandri <syandri_1960@bunghatta.ac.id> wrote:

Dear

Chandrika Harikrishnan

Quality Checker of International Journal of Food Science

Thank you for your email on December 16, 2023; The authors have revised the manuscript consisting of many et al on the references (attached).

Best Regards
Hafrijal Syandri

On Sat, Dec 16, 2023 at 11:53 AM Chandrika Harikrishnan
<chandrika.harikrishnan@hindawi.com> wrote:

Dear Dr. Syandri,

This is regarding the manuscript "**6639837**" titled "**Fatty Acids and Amino Acids of Three Local Freshwater Species of Indonesian Bagridae Fish for Food Security**" has been submitted to "**International Journal of Food Science**".

During our routine quality checks, we found that the manuscript consists of many et al references. Kindly update the reference section and provide a detailed reference list that includes all authors names (***explain et al.***), ***title, journal name, and published year*** in the final manuscript and send us the file in document format via email in order to proceed further.

An early reply would be appreciated.

Best regards,
Chandrika

Chandrika Harikrishnan
Quality Checker



Hindawi

[Hindawi.com](https://www.hindawi.com) | [Twitter](#) | [FaceBook](#) | [LinkedIn](#) | [YouTube](#)

The contents of this email and any attachments are confidential and intended only for the person or entity to whom it is addressed. If you are not the intended recipient, any use, review, distribution, reproduction or any action taken in reliance upon this message is strictly prohibited. If you received this message in error, please immediately notify the sender and permanently delete all copies of the email and any attachments.

[Click here for translations of this disclaimer.](#)

Analysis of Fatty Acids and Amino Acids of Three Local Freshwater Bagridae Fish Species in the Kampar Kanan River, Indonesia for Food Security

**Azrita Azrita¹, Hafrijal Syandri^{1*}, Hazlina Ahamad Zakeri²,
Harfiandri Damanhuri¹, Netti Aryani³**

¹Faculty of Fisheries and Marine Science, Bung Hatta University, 25131, Padang-West Sumatra Province, Indonesia

²Faculty of Science and Marine Environment, Universiti Malaysia Terengganu, Malaysia.

³Faculty of Fisheries and Marine Science, Riau University, 28293, Pekanbaru-Riau Province, Indonesia

Analysis of Fatty Acids and Amino Acids of Three Local Freshwater Bagridae Fish Species in the Kampar Kanan River, Indonesia for Food Security
**Azrita Azrita¹, Hafrijal Syandri^{1*}, Hazlina Ahamad Zakeri², Harfiandri Damanhuri¹,
Netti Aryani³**

¹*Faculty of Fisheries and Marine Science, Bung Hatta University, 25131, Padang-West Sumatra Province, Indonesia*

²*Faculty of Science and Marine Environment, Universiti Malaysia Terengganu, Malaysia.*

³*Faculty of Fisheries and Marine Science, Riau University, 28293, Pekanbaru-Riau Province, Indonesia*

*Corresponding author: syandri_1960@bunghatta.ac.id

Abstract

Fish have become an irreplaceable dietary source of animal protein, especially among households with low socioeconomic status in rural and urban areas of Indonesia. This study aimed to analyse the proximate composition, minerals, fatty acids, and amino acids of three local Bagridae fish species in Indonesia. The standard AOAC method was employed to examine the proximate composition of the carcass, and the analysis of amino acids and fatty acids was conducted through HPLC and GC-MS, respectively. The mineral content was determined using AAS. The nutrient composition results of *Hemibagrus nemurus*, *Hemibagrus wyckii*, and *Mystus nigriceps* revealed that the protein content was 24.26%, 22.57%, and 21.39% (% dry weight), whereas the total lipid content was 6.64%, 7.47% and 7.75%, respectively. Regarding mineral contents, the calcium levels ranged from 1.49–1.66 mg/g, iron levels from 28.35–40.36 µg/g, and zinc levels from 24.03–54.46 µg/g. Among the fatty acids, palmitic acid was the most abundant in all three species, accounting for 25.59–30.70% of the total fatty acids. Additionally, significant amounts of C18:1 (oleic acid), C18:0 (stearic acid), and C20:4 (arachidonic acid) were also detected as primary fatty acids. The calculated atherogenic index values in the three species of Bagridae fish ranged from 0.73–0.99, while the thrombogenic index values varied between 0.54–0.75. The predominant amino acids found in the three species of Bagridae fish were glutamic acid with their concentrations ranging from 9.10–24.34%. These results indicate that consuming the meat of these three freshwater Bagridae fish species caught in the wild does not pose any health risks to consumers. They can be considered a safe and suitable food source with good nutritional quality.

Keywords: Amino acids, Fatty acids, Indonesia, Kampar Kanan River, Minerals

1. Introduction

Fish serve as a high-quality protein source that is essential for satisfying dietary protein requirements, especially in areas where other protein sources are few or difficult to obtain [1,2]. Their accessibility can aid in ensuring a steady supply of food, particularly for communities whose main source of income is fishing or aquaculture [3]. Fish can contribute to a more varied selection of protein sources in communities, resulting in a diet that is more nutritious and balanced. Dietary fish can treat nutrient deficits and enhance general nutritional health. Fish are generally considered safe and offer a lower risk of contamination with harmful substances compared to some land-based animal products. A reliable indicator of a fish's quality, nutritional value, physiological status, and environment is its chemical makeup, which is found in fish flesh [4].

Hemibagrus nemurus, locally known as "baung", *Hemibagrus wyckii*, known as "geso" and *Mystus nigriceps*, known as "ingir-ingir," are three species of catfish found in the Kampar Kanan River, Indonesia [5,6]. Due to their high market value, these species have been identified as significant economic resources for rural communities [7]. Therefore, the analysis of the nutritional quality of fish meat in the study area is very important to determine whether it is beneficial to human health.

Knowing the composition of fatty acids and amino acids in fish meat is an essential factor that should not be ignored [8]. This information provides valuable insight into the nutritional quality of fish and the values of more nutritious and healthy foods [9]. In addition, a better understanding of the composition of fatty acids and amino acids can assist in selecting a suitable feed [10], developing a balanced diet [11], and maintaining optimal fish meat nutrition [12].

Omega-3 polyunsaturated fatty acids (PUFAs), particularly docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA) derived from fish oil, exhibit anti-inflammatory and anti-gastrointestinal cancer properties, making them crucial as immune-boosting nutrients [13,14]. Studies have demonstrated that ω -3 and ω -6 PUFAs exhibit beneficial effects in the management of cardiovascular diseases and cancers [15,16]. The fatty acid composition of PUFAs can differ among different types of fish, including both freshwater and marine species [17, 18]. Amino acids (AAs), such as cysteine, arginine, tyrosine, glycine, proline, and serine,

play a vital role in illness and stress situations and in the prevention of inflammation and repair processes in fish intestines [19, 20].

Furthermore, knowledge of the atherogenicity index (AI) and thrombogenicity index (TI) of fatty acids in the three local freshwater fish species of Bagridae can contribute to improving dietary guidelines. The AI and TI can be used to develop recommendations aimed at improving heart health [21,22]. These recommendations may involve selecting fish with favourable fatty acid profiles, allowing individuals to choose the most suitable fish species based on their specific needs [17, 23]. Therefore, this study aims to examine the nutritional composition profile, mineral content, amino acids, and fatty acids, and AI and TI of three freshwater fish species, namely, *H. nemurus*, *H. wyckii*, and *M. nigriceps*. These three fish species are generally consumed by people who live around the Kampar Kanan River in Kampar Regency, Riau Province, Indonesia.

2. Material and methods

2.1. Animal material

In this study, a total of 27 Bagridae fish belonging to three distinct species (*H. nemurus*, *H. wyckii*, and *M. nigriceps*) were examined. The specimens were obtained from local fishers operating in the upper stretches of the Kampar Kanan River in Kampar Regency, Riau Province (Figure 1). Sampling took place between June and August 2022 and was verified by the Fisheries Department, Faculty of Fisheries and Marine Science, Bung Hatta University, Indonesia. The samples were placed in plastic bags and transported to the laboratory in an insulated icebox. The samples were then grouped based on their respective sampling locations.

2.2. Biometry measurement

Upon reaching the laboratory within 10 hours, the fish samples were subjected to gutting and washing. Each specimen was weighed (TW) and measured to determine the standard length (SL) as well as the depth or maximum height (H). TW was measured using balance scale (OHAUS Model CT 6000 USA) with a precision of 0.01 g. The length measurement was taken from the tip of the mouth to the end of the upper lobe of the caudal fin, representing the total body length, using a metre ruler with an accuracy of 1 mm. The height measurement involved a vertical assessment of the body's maximum height measured using a Digital Sekhmet Sigma Vernier calliper with an accuracy of 1 mm. The condition factor (CF) was calculated using the formula $CF = (TW/SL^3) \times 100$.

2.3. Proximate, amino acid, and mineral content analysis

The proximate carcass composition was assessed following the AOAC standard methods [24]. The samples were dried to a constant weight at 105 °C. The crude protein content was analysed using the standard Kjeldahl method, which involves multiplying the nitrogen content by a factor of 6.25. The Soxhlet method with ether extraction was employed to analyse crude lipids. The ash content was obtained by incinerating the samples at 550 °C for 16 h. The amino acid composition was determined using a high-performance liquid chromatography (HPLC) system, which consisted of a Waters® 1525 binary HPLC pump, Waters® 717 autosamplers, and Waters® 2475 multi- λ fluorescence detector (with excitation at 250 nm and emission at 395 nm). The samples were hydrolysed in triplicate with 6 N hydrochloric acid for 24 h at 11 °C [25].

For mineral data composition (Na, Mg, Ca, K, P, Fe, Cu, Mn, and Zn), the ashed feed sample and carcass were dissolved in 1 ml of hydrochloric acid (35% v/v Suprapur®, Merck) and then filtered with cellulose filter paper (Watchman No 1, International Ltd; Maidstone, UK) and diluted to an appropriate concentration for each elemental mineral. P levels were analysed with a Perkin-Elmer AA spectrophotometer mod 3110 (Norwalk, CT, USA).

2.4. Fatty acid analysis

The fish meat was examined utilizing the gas chromatography–mass spectrometry (GC–MS) method. The method of Folch et al. (1957) modified by Rajion et al. [26] was employed to perform the total fat extraction. This involved using a solvent system consisting of chloroform and methanol at a 2:1 (v/v) ratio. The process of transmethylation was conducted using a solution of 14% methanolic boron trifluoride. The fatty acid composition of the meat was then analysed at the SIG Laboratory, Accredited Testing Laboratory-LP-184-IDN.

The nutritional quality of lipids (AI and TI) was calculated using the following equations [27]:

$$AI = \frac{[(C12:0 + 4 \times C14:0 + C16:0)]}{[(\sum MUFA + \sum n - 6 + \sum n - 3)]}$$

$$TI = \frac{[(C14:0 + C16:0)]}{[(0.5 \times \sum MUFA) + (0.5 \times \sum n - 6) + (3 \times \sum n - 3) + (\sum n - 3 / \sum n - 6)]}$$

where:

AI = Atherogenic index

TI = Thrombogenic index

C12:0 = lauric acid

C14:0 = myristic acid

C16:0 = palmitic acid

\sum MUFA = sum of the concentrations of all monosaturated fatty acids

\sum n-6 = sum of the concentrations of n-6 polyunsaturated fatty acids

\sum n-3 = sum of the concentrations of n-3 polyunsaturated fatty acids

2.5. Data analysis

Data analysis was performed using the SPSS 16.0 software package (SPSS; Chicago, IL). The homogeneity of variances was assessed using Levene's test. One-way ANOVA was carried out to determine the parameters of proximate and amino acid composition, including the composition of fatty acids and nutritional quality of lipids (AI and TI) for each species, followed by the post hoc Duncan's multiple range test [28]. The results are reported as the mean values \pm standard errors for each parameter.

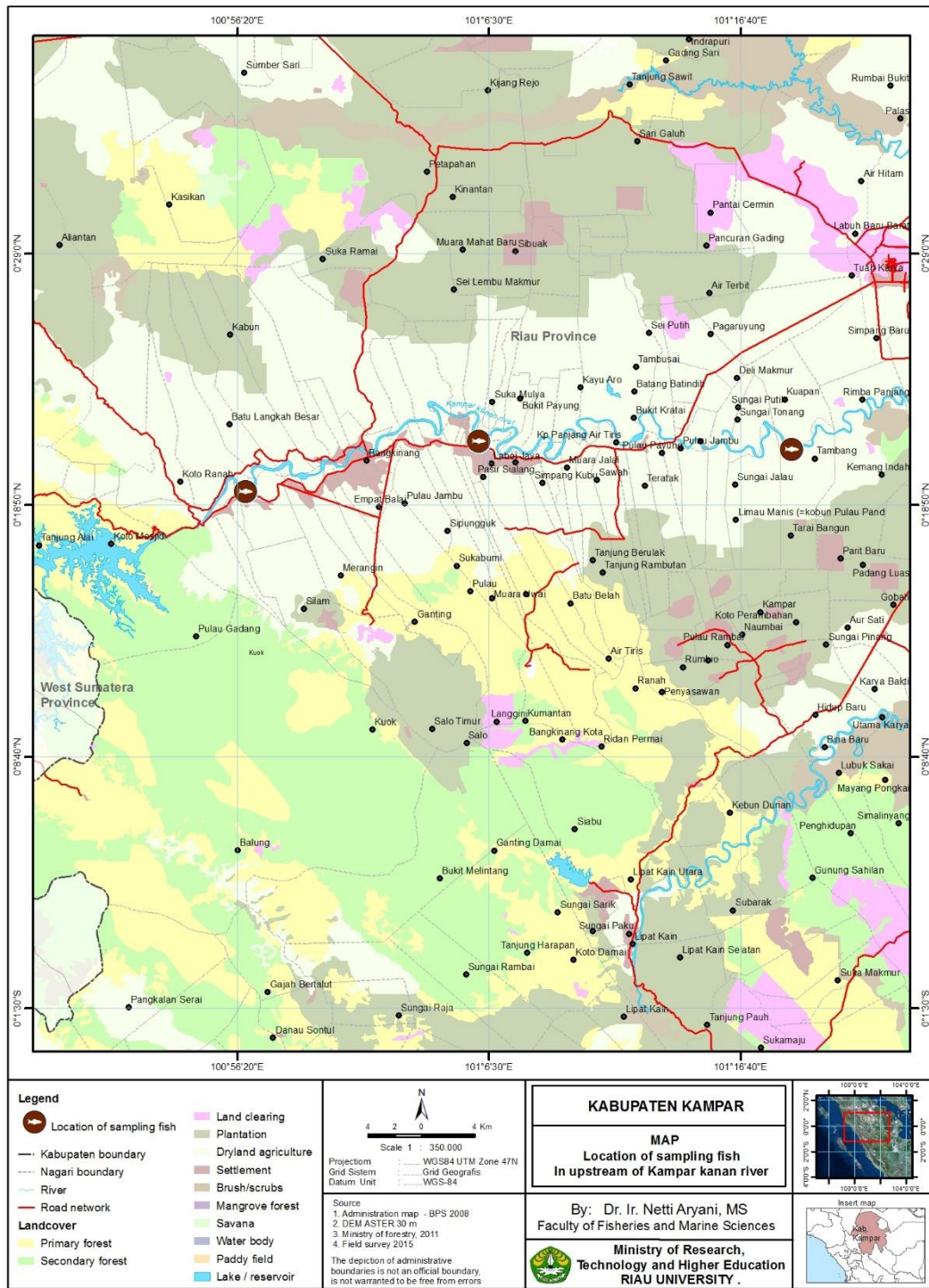


Figure 1. Map of Kampar Regency, Riau Province, and sampling locations of the three species of Bagridae fish.

3. Results

Table 1 presents the average wet weight, standard length, height, and the results of meat nutritional composition analysis for three indigenous Indonesian freshwater Bagridae species. The moisture content (% wet weight) ranged from 82.40– 85.39%. Among the species examined, *M. nigriceps* exhibited the lowest protein content at 21.39%, while *H. nemurus* displayed the highest value at 24.26%. However, *M. nigriceps* had a higher mineral content, including of iron and zinc.

Table 2 summarizes the fatty acid composition (%total fatty acid) for the three Bagridae fish species. C16:0 was the predominant fatty acid in the three species of Bagridae fish, with percentages ranging from 25.09–30.71%. The additional prominent fatty acids found were C18:1, C18:0, and C22:6. *H. nemurus* exhibited a ω -6/ ω -3 ratio below 1 (0.82), while *H. wyckii* and *M. nigriceps* had ratios of 1.11 and 1.02, respectively. The obtained PUFA/saturated fatty acid (SFA) (P/S) ratio varied between 0.61 and 0.76. The AI values varied between 0.76 and 0.99 and the TI between 0.40 and 0.63 (**Table 3**).

Table 4 presents the amino acid composition (% of total protein) of the three Bagridae species. Glutamic acid was the dominant amino acid, ranging from 19.29–24.34%, followed by aspartic acid, which ranged from 9.21–11.27%. The lysine content was consistent, ranging from 9.67–9.87%. In *H. nemurus*, the levels of various amino acids ranged from 0.86– 24.35%. Similarly, in *H. wyckii*, the amino acid levels ranged from 0.76–21.58%, while in *M. nigriceps*, the levels ranged from 0.74–19.29%.

Table 1. Results of biometric, proximate and mineral composition of three species of Bagridae fish

	Species		
	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
<i>Biometric measurements</i>			
Total weight (g)	389.99 ± 24.96 ^a	1,390.33 ± 168.29 ^b	17.57 ± 1.53 ^c
Standard length (cm)	28.16 ± 0.53 ^a	44.14 ± 1.98 ^b	10.29 ± 0.15 ^c
Height (cm)	8.44 ± 0.16 ^a	8.82 ± 0.39 ^b	2.57 ± 0.03 ^c
Condition factor	1.63 ± 0.08	1.58 ± 0.07	1.56 ± 0.08
<i>Proximate composition</i>			
Crude protein (% DW)	24.26 ± 0.87 ^a	22.57 ± 0.37 ^b	21.39 ± 0.15 ^c
Crude fat (% DW)	6.64 ± 0.03 ^a	7.47 ± 0.02 ^b	7.75 ± 0.40 ^c
Crude ash (% DW)	1.94 ± 0.02 ^a	2.30 ± 0.09 ^b	1.59 ± 0.02 ^c

Moisture (% WW)	82.40 ± 2.51	85.39 ± 2.36	83.75 ± 2.21
<i>Mineral composition</i>			
<i>Macrominerals (mg/g)</i>			
Sodium	0.99 ± 0.00 ^a	1.61 ± 0.01 ^b	1.61 ± 0.02 ^c
Magnesium	1.12 ± 0.01 ^a	0.57 ± 0.02 ^b	1.15 ± 0.01 ^c
Calcium	1.66 ± 0.00 ^a	1.55 ± 0.02 ^b	1.49 ± 0.04 ^c
Potassium	0.71 ± 0.00 ^a	0.55 ± 0.02 ^b	0.43 ± 0.01 ^c
Phosphorous	7.03 ± 0.03 ^a	2.74 ± 0.02 ^b	6.45 ± 0.07 ^c
<i>Microminerals (µg/g)</i>			
Iron	28.30 ± 0.11 ^a	28.73 ± 0.08 ^b	40.36 ± 0.55 ^c
Copper	8.93 ± 0.03 ^a	7.46 ± 0.09 ^b	6.21 ± 0.32 ^c
Manganese	1.64 ± 0.02 ^a	2.64 ± 0.02 ^b	2.84 ± 0.02 ^c
Zinc	24.03 ± 0.45 ^a	24.61 ± 0.19 ^b	54.46 ± 0.17 ^c

Values are mean % ± SE; *Values in the same row followed by different letters are significantly different (p<0.05).

Table 2. Fatty acid composition of three species of Bagridae fish

Fatty acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
C12:0 (lauric acid)	1.97 ± 0.05 ^a	3.94 ± 0.01 ^b	4.27 ± 0.01 ^c
C14:0 (Meristic acid)	2.38 ± 0.03 ^a	2.72 ± 0.08 ^b	2.89 ± 0.01 ^c
C16:0 (Palmitic acid)	27.23 ± 0.06 ^a	30.70 ± 0.06 ^b	25.59 ± 0.01 ^c
C18:0 (Stearic acid)	16.17 ± 0.02 ^a	13.54 ± 0.02 ^b	10.64 ± 0.01 ^c
C20:0 (Arachidic acid)	0.23 ± 0.01 ^a	0.14 ± 0.01 ^b	0.37 ± 0.01 ^c
C16:1 (Palmitoleic acid)	1.78 ± 0.00 ^a	1.27 ± 0.04 ^b	4.93 ± 0.01 ^c
C18:1 (Oleic acid)	16.86 ± 0.01 ^a	16.18 ± 0.01 ^b	16.97 ± 0.01 ^c
C18:2 (Linoleic acid)	3.24 ± 0.02 ^a	3.84 ± 0.33 ^b	4.55 ± 0.001 ^c
C18:3 (Linolenic acid)	1.12 ± 0.01 ^a	0.82 ± 0.01 ^b	1.57 ± 0.000 ^c
C20:4 (Arachidonic acid)	12.13 ± 0.01 ^a	12.49 ± 0.09 ^b	12.41 ± 0.02 ^c
C20:5 (Eicosapentaenoic acid; EPA)	2.06 ± 0.08 ^a	2.34 ± 0.04 ^b	1.35 ± 0.01 ^c
C22:6 (Docosahexaenoic acid; DHA)	15.65 ± 0.01 ^a	11.56 ± 0.04 ^b	13.64 ± 0.02 ^c

Values are mean % ± SE; *Values in the same row with a different superscript are significantly different (p<0.05).

Table 3. Fatty acid ω-6/ω-3 and polyunsaturated/saturated fatty acid ratio of three species of Bagridae fish

	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
--	---------------------------	--------------------------	-------------------------

ω -6/ ω -3 ratio	0.82 \pm 0.00 ^a	1.11 \pm 0.02 ^b	1.02 \pm 0.01 ^c
P/S ratio	0.69 \pm 0.01 ^a	0.61 \pm 0.06 ^b	0.76 \pm 0.00 ^c
AI	0.87 \pm 0.01 ^a	0.99 \pm 0.06 ^b	0.73 \pm 0.00 ^c
TI	0.40 \pm 0.01 ^a	0.63 \pm 0.01 ^b	0.43 \pm 0.00 ^c

P/S, polyunsaturated/saturated fatty acid; AI, atherogenic index; TI, thrombogenic index
 Values are mean % \pm SE; *Values in the same row followed by different letters are significantly different (p<0.05).

Table 4. Amino acid composition of three species of Bagridae fish

Amino acid	<i>Hemibagrus nemurus</i>	<i>Hemibagrus wyckii</i>	<i>Mystus nigriceps</i>
Aspartic acid	10.39 \pm 0.01 ^a	9.61 \pm 0.04 ^b	11.28 \pm 0.02 ^c
Glutamic acid	24.34 \pm 0.00 ^a	21.58 \pm 0.41 ^b	19.29 \pm 0.03 ^c
Serine	3.74 \pm 0.00 ^a	5.40 \pm 0.03 ^b	7.50 \pm 0.06 ^c
Glycine	4.02 \pm 0.00 ^a	5.77 \pm 0.04 ^b	4.69 \pm 0.02 ^c
Histidine	2.39 \pm 0.00 ^a	2.90 \pm 0.03 ^b	2.21 \pm 0.02 ^c
Arginine	6.17 \pm 0.01 ^a	8.27 \pm 0.03 ^b	6.22 \pm 0.03 ^c
Threonine	4.49 \pm 0.01 ^a	4.19 \pm 0.03 ^b	4.39 \pm 0.04 ^c
Alanine	7.03 \pm 0.00 ^a	6.04 \pm 0.02 ^b	7.89 \pm 0.01 ^c
Proline	2.40 \pm 0.00 ^a	2.71 \pm 0.01 ^b	2.21 \pm 0.02 ^c
Tyrosine	3.91 \pm 0.01 ^a	3.29 \pm 0.06 ^b	3.82 \pm 0.04 ^c
Valine	3.73 \pm 0.00 ^a	4.02 \pm 0.01 ^b	3.85 \pm 0.04 ^c
Methionine	3.42 \pm 0.00 ^a	1.96 \pm 0.02 ^b	1.27 \pm 0.03 ^c
Cystine	0.87 \pm 0.00 ^a	0.76 \pm 0.02 ^b	0.75 \pm 0.03 ^c
Isoleucine	3.82 \pm 0.001 ^a	3.96 \pm 0.003 ^b	3.66 \pm 0.002 ^c
Leucine	6.15 \pm 0.000 ^a	6.69 \pm 0.002 ^b	6.28 \pm 0.004 ^c
Phenylalanine	5.13 \pm 0.000 ^a	6.03 \pm 0.005 ^b	5.18 \pm 0.027 ^c
Lysine	9.61 \pm 0.001 ^a	9.82 \pm 0.026 ^b	9.87 \pm 0.003 ^c

Values are mean % \pm SE; *Values in the same row followed by different letters are significantly different (p<0.05).

4. Discussion

The results of this study provide important new information about the nutritional makeup of the studied species of Bagridae fish. The biometric measures of the fish yielded important information about their physical traits and health. Researchers are able to determine the average weight of fish at various lengths by examining the link between length and weight for a particular species. This knowledge is essential for understanding the trends in fish population growth, identifying potential shifts in growth rates over time, and assessing the general health and welfare of fish [29]. A variety of factors, including sex, age, maturity level, size, stomach fullness level, sampling strategies, sample sizes, and environmental circumstances, have an impact on fish health and the parameters that determine length–weight connections in fish [30]. However, none of these factors were considered in our study. In general, according to the theory put forth by [Bagenal and Tesch \[31\]](#), heavier fish of a given length are in better physiological condition. The condition factor is also used in fisheries science to compare the health, fatness, and condition of fish [32]. A condition factor of > 1 is favourable, indicating a good feed nutrition level and suitable living habitat [33]. Thus, based on the findings in this study, the Kampar Kanan River provides a suitable habitat for the fish, ensuring their accessibility and availability for community consumption.

The fish samples exhibited good nutritional profiles, with high amounts of proteins and lipids as well as moisture content contributing to their overall nutritional value, according to the results of proximate analysis and mineral composition (see Table 1). These results imply that the fish species studied provide a possible source of nutrients to solve issues with food security. The study of the mineral composition revealed the existence of critical minerals such as phosphorus (P), calcium (Ca), magnesium (Mg), iron (Fe), and zinc (Zn) in the fish samples in addition to proteins and lipids. All three Bagridae fish significantly differed from each other in terms of their crude ash content and mineral profile (see Table 1). However, in general, P and Ca are the two highest minerals found in all fish. Both of these macro minerals are good for human bone and tooth health, as key components of hydroxyapatite, the mineral matrix that provides strength and structure to bones and teeth [34].

In the current study, the meat of three species of Bagridae fish from the Kampar Kanan River in Indonesia was found to contain an abundance of microminerals such as Fe and Zn. *M. nigriceps* has been reported to exhibit higher mineral content, including of Fe and Zn. This

species is categorized as a small indigenous fish (SIF) with a maximum standard length of approximately 10.29 cm. According to [35], SIFs are generally known to be rich in Ca, Fe, and Mn. On the other hand, scale flour from three species of freshwater fish, such as *Osphronemus goramy*, *Cyprinus carpio*, and *Oreochromis niloticus*, also contains high levels of macrominerals and microminerals [36].

In this study, all three Bagridae fish had a total fat content of approximately 7–8% by weight, with a higher water content ranging from 82.40–85.39% (Table 1). These values are similar to those found in wild and cultured sea bass *Dicentrarchus labrax* [37]. Additionally, the fish species can be grouped into different categories, including high-fat fish (>8%), medium-fat fish (4–8%), low-fat fish (2–4%), and lean fish (<2%) [38]. According to the classification established by [38], the three indigenous Bagridae fish species are characterized as having a moderate fat content level, which is reflected by the high levels of DHA and EPA in the meat of *H. nemurus*, *H. wyckii*, and *M. nigriceps* (Table 2). Therefore, we recommend that consuming the meat of these three fish species can indirectly boost food security in rural and urban communities.

All three Bagridae fish species analysed in the study contained arachidonic acid (C20:4), at 12.13% for *H. nemurus*, 12.49% for *H. wyckii*, and 12.41% for *M. nigriceps*. Arachidonic acid serves as a precursor for synthesizing prostaglandins and leukotrienes [39]. Leukotrienes are crucial in allergic responses, inducing smooth muscle contraction and increasing vascular permeability [40]. Additionally, all three species also contained EPA ranging from 1.35–2.34% and DHA ranging from 11.56–15.65%. The increased consumption of omega-3 fatty acids, such as EPA and DHA, has been linked to a decreased risk of pain development in older individuals and to the prevention of Alzheimer's disease [41,42].

As determined by the current study, the ω -6/ ω -3 ratios of the three Bagridae fish species ranged from 1:0.8 to 1:0.97. It was found that all three fish species had ω -6/ ω -3 ratios within the recommended range for a typical Indonesian diet. In contrast, a typical Western diet is characterized by a high ω -6/ ω -3 ratio of 10:1 to 20:1, while a Mediterranean diet has a ratio of 4:1 to 5:1 [43]. Additionally, the P/S ratio analysis showed that the three Bagridae fish species are sources of PUFAs that meet the requirements for food safety.

The Food and Agriculture Organization (FAO) and the World Health Organization (WHO) have recommended AI and TI values ranging from 0.4 to 0.5 for promoting human health [44].

In the current study, the observed AI values in the three freshwater species of Bagridae ranged from 0.73 to 0.99, while the TI values varied between 0.40 and 0.63. This finding is associated with a notable disparity in the values of SFAs among the three Bagridae fish species. Based on these findings, we confirm that consuming the meat of the three freshwater species of Bagridae fish caught in the wild does not endanger consumer health and can serve as a suitable source of food safety and nutritional quality.

Fish with a high crude protein content, as shown by the three studied Bagridae fish (Table 1), are regarded as being nutritionally useful for human health. The protein and amino acid contents can directly indicate the nutritional quality of meat [45]. As shown in Table 4, *H. wyckii* had a higher content of essential amino acids (EAAs) than *H. nemurus* and *M. nigriceps*. Among the EAAs, lysine (Lys) was found to be present in the highest amount in all three species. Lys is necessary for tissue growth and repair, reduced stress-induced anxiety, and the conversion of fatty acids into energy [46-48].

More than 60% of the amino acid composition in all three Brigade fish studied comes from nonessential amino acids (NEAAs), particularly glutamic acid (Table 4). Glutamic acid and its amine, glutamine, are highly abundant amino acids found in fish in the free and protein-bound forms [49]. Other amino acids found at abundant levels in the three fish also play important physiological and biochemical functions. The NEAA arginine, for example, is involved in protein synthesis, creatine formation, and various other biochemical processes within the body. The EAAs leucine, phenylalanine, and methionine play important roles in the synthesis of nitric oxide, a molecule that can expand blood vessels [49].

4. Conclusions

The present study provides insightful information about the fatty acid and amino acid compositions, biometric measurements, proximate analysis, and mineral composition of three local freshwater species of Indonesian Bagridae fish. The findings show their potential as a nutrient-dense, sustainable food source that can support efforts to increase food security. These fish species might be incorporated into the local diet to increase nutrient intake and alleviate nutritional inadequacies, which would ultimately improve Indonesia's overall food security status.

Data Availability

The data utilized in this research has not been previously released or published in any form. The datasets employed and/or analysed during the present study can be obtained by contacting the corresponding author.

Conflicts of Interest

The authors confirm that they do not have any conflicting interests.

Acknowledgements

The authors acknowledge and express their gratitude for the financial support received from the Ministry of Education, Culture, Research, and Technology of the Republic of Indonesia, specifically through grant no. 076/E5/PG. 02.00. PT/2022, which funded the research. The authors affirm that there is no conflict of interest associated with the utilization of these research funds. Furthermore, the authors extend their appreciation to the students and fishers involved in the research conducted in the upper reaches of the Kampar Kanan River, Indonesia.

References

- [1] A.E. Hasselberg, I.Aakre, J. Scholtens, R. Avera, J. Kolding, M.S. Bank, A. Atter, M. Kjellevoid., “Fish for food and nutrition security in Ghana: Challenges and opportunities,” *Global Food Security*, vol. 26, pp. 100380, 2020. <https://doi.org/10.1016/j.gfs.2020.100380>
- [2] N.Tran, U.P, Rodriguez, C.Y.Chan, Y. M. Auang, L. Chu, A.H.Md.S. Islam., “Future scenarios of fish supply and demand for food and nutrition security in Bangladesh: An analysis with the AsiaFish model”, *Aquaculture*, vol.568, pp.739288, 2023. <https://doi.org/10.1016/j.aquaculture.2023.739288>
- [3] C.Z. Levkoe, K. Lowitt, C. Nelson. “Fish as food: Exploring a food sovereignty approach to small-scale fisheries”. *Marine Policy*, vol. 85, pp.65-70, 2017. <https://doi.org/10.1016/j.marpol.2017.08.018>
- [4] M. Ravichandran, N.S. Hettiarachchy, V. Ganesh, S. Ricke., “Enhancement of antimicrobial activities of naturally occurring phenolic compounds by nanoscale delivery against listeria monocytogenes, escherichia coli O157:H7 and salmonella typhimurium in broth and chicken meat system”. *Journal of Food Safety*, vol. 31, no. 4, pp. 462-471,2011. <https://doi.org/10.1111/j.1745-4565.2011.00322.x>
- [5] N. Aryani, I. Suharman, Azrita, H. Syandri., “Diversity and distribution of fish fauna of upstream and downstream areas at Koto Panjang Reservoir, Riau Province, Indonesia”. *F1000 Research*, vol. 8, 1435.pp.1-16, 2020. <https://f1000research.com/articles/8-1435/v2>
- [6] Syafrialdi, Dahelmi, D.I. Roesma, H. Syandri., “Length-weight relationships and condition factor of Two-Spot Catfish (*Mystus nigriceps* [Valenciennes, 1840]) Pisces, Bagridae], from Kampar Kanan River and Kampar Kiri River in Indonesia”. *Pakistan*

- Journal of Biological Science*, vol. 23, no. 12, pp. 1636-1642, 2020. <https://scialert.net/fulltext/?doi=pjbs.2020.1636.1642>
- [7] N. Aryani, "Native species in Kampar Kanan River, Riau Province Indonesia". *International Journal of Fisheries and Aquatic Studies*, vol.2, no.5, pp. 213-217. 2015. <https://www.fisheriesjournal.com/archives/2015/vol2issue5/PartD/2-5-63.pdf>
- [8] Azrita, H. Syandri, N. Aryani, A. Mardiah, I. Suharman., "The utilization of new products formulated from water coconut, palm sap sugar, and fungus to increase nutritional feed quality, feed efficiency, growth, and carcass of gurami sago (*Osphronemus goramy* Lacepède, 1801) juvenile". *F1000 Research*, vol. 10, 1121, pp. 1-16, 2021. <https://f1000research.com/articles/10-1121/v1>
- [9] C. Bene, R. Arthur, H. Norbury, E.H. Allison, M. Beveridge, S. Bush, L. Campling, W. Leschen, D. Little, D. Squires, S.H. Thilsted, M. Troell, M. Williams., "Contribution of Fisheries and Aquaculture to Food Security and Poverty Reduction: Assessing the Current Evidence". *World Development*, vol. 79, pp. 177–196. 2016. <http://dx.doi.org/10.1016/j.worlddev.2015.11.007>
- [10] N. Aryani, I. Suharman, S. Hasibuan, N. Asia, H. Syandri., "Fatty acid composition on diet and carcasses, growth, body indices and profile serum of Asian redtail catfish (*Hemibagrus nemurus*) fed a diet containing different levels of EPA and DHA". *F1000Research*, vol, 11:1409, pp. 1-23, 2023. <https://doi.org/10.12688/f1000research.126487.2>
- [11] E.N. Kingsley, O.O. Cyril, O.I. Patience, "Potential contribution of selected wild fish species to the minerals intake of Pregnant and Lactating Women, Children and Adults in Rural Riverine Communities of Edo State: Insights and Outcomes". *Measurement: Food*, vol. 8, 100063, pp. 1-7, 2022. <https://doi.org/10.1016/j.meafoo.2022.100063>
- [12] A.A. Sausa, L. Nora, D.L.A. Lopes, T.G. Petrolli, V.J.M. Vurlan, R. Wagner, C.M. Giacomelli, M.D. Baldisera, A.S. Da Silva., Vegetable choline in feed for Nile tilapia (*Oreochromis niloticus*) raised in a biofloc technology system (BFT): Biofloc composition, chemical composition, and fatty acid profiles in meat". *Aquaculture*, vol. 545, pp. 737174, 2021. <https://doi.org/10.1016/j.aquaculture.2021.737174>
- [13] A.M. Eltweri, A.L. Thomas, M. Metcalfe, P.C. Calder, A.R. Dennison, D.J. Bowrey., Potential applications of fish oils rich in omega-3 polyunsaturated fatty acids in the management of gastrointestinal cancer. *Clinical Nutrition*. vol. 36, no. 1, pp. 65-78. 2017. <https://doi.org/10.1016/j.clnu.2016.01.007>
- [14] P.C. Haycock, M.C. Borges, K. Burrows, L.N. Letmetre, S. Burgess, N.K. Khankari, K.K. Tslidis et al., "The association between genetically elevated polyunsaturated fatty acids and risk of cancer". *eBiomedicine*. vol.91, pp. 104510, 2023. <https://doi.org/10.1016/j.ebiom.2023.104510>
- [15] E.K. Aglago, I. Huybrechts, N. Murphy, C. Casagrande, G. Nicolas et al., "Consumption of Fish and Long-chain n-3 Polyunsaturated Fatty Acids Is Associated With Reduced Risk of Colorectal Cancer in a Large European Cohort". *Clinical Gastroenterology and Hepatology*. vol. 18, no. 3, pp. 654-666.e6. 2020. <https://doi.org/10.1016/j.cgh.2019.06.031>
- [16] H. Jiang, X. Shi, T.Y. Fan, D. Wang, B. Li, J. Zhou, C. Pei, L. Ma., "Dietary omega-3 polyunsaturated fatty acids and fish intake and risk of age-related macular degeneration". *Clinical Nutrition*. vol.40, no. 12, pp: 5662-5673. 2021. [https://www.clinicalnutritionjournal.com/article/S0261-5614\(21\)00474-X/fulltext](https://www.clinicalnutritionjournal.com/article/S0261-5614(21)00474-X/fulltext)

- [17] M.A. Rincón-Cervera, N.V. González-Barriga, R. Valenzuela, S. Lopez-Arana, J. Romero, A. Valenzuela., “Profile and distribution of fatty acids in edible parts of commonly consumed marine fishes in Chile”, *Food Chemistry*, vol. 274, pp. 123-129, 2018. <https://doi.org/10.1016/j.foodchem.2018.08.113>
- [18] S. Sun, T. Ren, X. Li., X. Cao, J. Gao., “Polyunsaturated fatty acids synthesized by freshwater fish: A new insight to the roles of elov12 and elov15 in vivo”, *Biochemical and Biophysical Research Communications*, vol. 532, pp. 414-419, 2020. <https://doi.org/10.1016/j.bbrc.2020.08.074>
- [19] M.B. Behyar, M. Hasanzadeh, F. Seidi, N. Sadjou ., “Sensing of amino acids: Critical role of nanomaterials for the efficient biomedical analysis”, *Microchemical Journal*, vol. 188, pp. 108452, 2023 <https://doi.org/10.1016/j.microc.2023.108452>
- [20] P. Shi, K. Liao, J. Xu, Y. Wang, S. Xu, X. Yan., “Eicosapentaenoic acid mitigates palmitic acid-induced heat shock response, inflammation and repair processes in fish intestine”, *Fish & Shellfish Immunology*, vol. 124, pp. 362-371, 2022. <https://doi.org/10.1016/j.fsi.2022.04.011>
- [21] J. Wen, L. Zeng, Y. Xu, Y. Sun, Z. Chen, S. Fan., Proximate composition, amino acid and fatty acid composition of fish maws, *Natural Product Research: Formerly*, vol. 30, no. 2, pp. 214-217, 2015. <https://doi.org/10.1080/14786419.2015.1040790>
- [22] F.O.S. Duarte, F.G. de Paula, C.S. Prado, R.R. dos Santos, C.S. Minafra-Rezende, C. Gebara, M.E. Lage., “Better fatty acids profile in fillets of Nile Tilapia (*Oreochromis niloticus*) supplemented with fish oil”, *Aquaculture*, Vol. 534, pp. 736241, 2021. <https://doi.org/10.1016/j.aquaculture.2020.736241>
- [23] K.L. Weaver, P. Ivester, J.A.Chilton, M.D. Wilson, P. Pandey, F.H. Chilton, “The Content of Favorable and Unfavorable Polyunsaturated Fatty Acids Found in Commonly Eaten Fish”, *Journal of the American Dietetic Association*, vol. 108, No. 7 , pp. 1178–1185, 2008. <https://doi.org/10.1016/j.jada.2008.04.023>
- [24] AOAC, “Official Methods of Analysis. 15th Edn., Association of Official Analytical Chemists (AOAC)”, Washington, DC., USA.1990. Reference Source
- [25] S.A.Cohen, “Amino acid analysis using pre-column derivatization with 6-aminoquinoly-N-hydroxysuccinimidyl carbamate”, *Protein Sequencing Protocols*, vol. 211, pp. 143-54, 2003. <https://doi.org/10.1385/1-59259-047-0:039>
- [26] M.A. Rajion, J.G. McLean, R.N. Cahill, “Essential fatty acid metabolism in the fetal and neonatal lamb”, *Australian Journal of Biological Sciences*, vol. 38, no. 1, pp. 33-40, 1985. [Essential fatty acids in the fetal and newborn lamb - PubMed \(nih.gov\)](https://pubmed.ncbi.nlm.nih.gov/)
- [27] T.L. Ulbricht, D.A. Southgate, “Coronary heart disease: Seven dietary factors”, *The Lancet*, Vol 338, no. 8773, pp. 985–992, 1991. [https://doi.org/10.1016/0140-6736\(91\)91846-M](https://doi.org/10.1016/0140-6736(91)91846-M)
- [28] D.B. Duncan, “Multiple ranges and multiple F tests”, *Biometrics*, vol. 11, pp. 1–42, 1955. <https://doi.org/10.2307/3001478>
- [29] R. Froese, “Cube law, condition factor and weight–length relationships: History, meta-analysis and recommendations”. *Journal of Applied Ichthyology*, vol. 22, no. 4, pp. 241-253, 2006. <https://doi.org/10.1111/j.1439-0426.2006.00805.x>
- [30] O.O.Famoofo, & W.O. Abdul, “Biometry, condition factors and length-weight relationships of sixteen fish species in Iwopin fresh-water ecotype of Lekki Lagoon, Ogun

- State, Southwest Nigeria”, *Heliyon*, vol. 6, no. 1, pp. e02957, 2020.
<https://doi.org/10.1016/j.heliyon.2019.e02957>
- [31] T.B. Bagenal, F.W. Tesch, “Age and Growth. In: Bagenal, T., Ed., *Methods for Assessment of Fish Production in Fresh Waters*”, *Blackwell Science Publications*, pp. 101-136, 1978.
- [32] E.O. Ahmed, M.E. Ali, A.A. Aziz, “Length-weight relationships and Condition factors of six fish species in Atbara River and Khashm el-girba Reservoir, Sudan”, *International Journal of Agriculture Sciences*, vol. 3, no.1, pp. 65–70, 2011.
<http://dx.doi.org/10.9735/0975-3710.3.1.65-70>
- [33] N.C. Ujjania, M.P.S. Kohli, L.L. Sharma, “Length-weight relationship and condition factors of Indian major carps (*C. catla*, *L. rohita* and *C. mrigala*) in Mahi Bajaj Sagar, India”, *Research Journal of Biology*, vol. 2, no.1, pp. 30-36, 2012.
<https://doi.org/10.21077/ijf.2017.64.special-issue.76263-27>
- [34] J.P. Bonjour, “Calcium and phosphate: A duet of ions playing for bone health”, *Journal of the American College of Nutrition*, vol. 30, no. 5, pp. 438S-48S, 2011.
<https://doi.org/10.1080/07315724.2011.10719988>
- [35] B.P. Mohanty, T.V. Sankar, S. Ganguly, A. Mahanty, R. Anandan, K. Chakraborty et al., “Micronutrient composition of 35 food fishes from India and their significance in human nutrition”, *Biological Trace Element Research*, vol. 174, pp. 448-458, 2016.
<https://doi.org/10.1007/s12011-016-0714-3>
- [36] H. Syandri, A. Azrita, A. Mardiah, N. Aryani, A. Diharmi, “The proximate composition, amino acid profile, fatty acid content, and mineral content of scale flour from three fish species as potential feeds for fish fry”, *F1000 Research*. Vol. 12, 1144, pp. 1-19, 2023.
<https://f1000research.com/articles/12-1144/v2>
- [37] F. Jabeen, A.A. Chudhry, “Chemical composition and fatty acid profiles of three freshwater species”, *Food Chemistry*, vol. 125, pp. 991-996, 2011.
<https://doi.org/10.1016/j.foodchem.2010.09.103>
- [38] A.A. Nurnadia, A. Azrina, I. Amin, “Proximate composition and energetic value of selected marine fish and shellfish from the west coast of peninsular Malaysia”. *International Food Research Journal*. Vol 18, pp. 137-148, 2011.
<https://core.ac.uk/download/pdf/153807129.pdf>
- [39] S.C. Dyllal, L. Balas, N.G. Bazan, J.T. Brena, N. Chiang, F.C. Sauza, J. Dalli, T. Durand, J. Galano, P.J. Lein, C.N. Serhan, A.Y. Taha., “Polyunsaturated fatty acids and fatty acid-derived lipid mediators: Recent advances in the understanding of their biosynthesis, structures and functions”, *Progress in Lipid Research*. vol. 86, pp. 101165, 2022.
<https://doi.org/10.1016/j.plipres.2022.101165>
- [40] N. Kim, U.D.Sohn, V. Mangannan, H. Rich, M.K. Jain, J. Behar, P. Biancani., “Leukotrienes in Acetylcholine-Induced Contraction of Esophageal Circular Smooth Muscle in Experimental”, *Gastroenterology*, vol. 112, no.5, pp. 1548–1558, 1997.
[https://doi.org/10.1016/S0016-5085\(97\)70036-2](https://doi.org/10.1016/S0016-5085(97)70036-2)
- [41] D. Swanson, R. Block, S.A.Mousa, “Omega-3 Fatty Acids EPA and DHA: Health Benefits Throughout Life”. *Advances in Nutrition*, vol. 3, no.1, pp. 1-7, 2012.
<https://doi.org/10.3945/an.111.000893>

- [42] R.P. Mason, S.C.R. Sherratt, L.H. Eckel, “Omega-3-fatty acids: Do they prevent cardiovascular disease?”, *Best Practice & Research Clinical Endocrinology & Metabolism*, vol.37, no 3, 101681, 2023. <https://doi.org/10.1016/j.beem.2022.101681>.
- [43] J. M. Seddon, J. Cote, N. Davis, B. Rosner., “Progression of age-related macular degeneration. Progression of age-related macular degeneration”. *Arch Ophthalmol*. Vol. 121, no. 6, pp. 785-792, 2003. <https://doi.org/10.1001/archophth.121.6.785>
- [44] FAO/WHO: Fat and Oils in Human Nutrition: Report of a Joint Expert Consultation. FAO/WHO, Rome, Italy.
- [45] S.M. Pawar, S.R. Sonaware, “Fish muscle protein highest source of energy”. *International Journal of Biodiversity and Conservation*. Vol. 5, no. 7, pp. 433-435, 2013. <https://doi.org/10.5897/IJBC12.043>
- [46] D. Tomé, C. Bos, “Lysine requirement through the human life cycle”, *The Journal of Nutrition*, vol. 137, no.6, pp. 1642S–1645S, 2007. <https://doi.org/10.1093/jn/137.6.1642S>
- [47] M. Watford, “Glutamine and glutamate: Nonessential or essential amino acids?”, *Animal Nutrition*, vol. 1, no. 3, pp. 119-122, 2015. <https://doi.org/10.1016/j.aninu.2015.08.008>
- [48] X. Li, S. Zheng, G. Wu, “Nutrition and metabolism of glutamate and glutamine in fish”, *Amino Acids*. vol.52, no.5, pp. 671-691, 2020. <https://doi.org/10.1007/s00726-020-02851-2>
- [49] M. Li, Y. Wu, L. Ye, “The Role of Amino Acids in Endothelial Biology and Function”, *Cells*, vol.11, no.8, pp. 1372, 2022. <https://doi.org/10.3390/cells11081372>



Webmail
Univ. Bung Hatta

hafrizal syandri <syandri_1960@bunghatta.ac.id>

6639837: Galley Proofs

2 messages

International Journal of Food Science <production.a@hindawi.com>

Wed, Dec 27, 2023 at 8:23 PM

To: syandri_1960@bunghatta.ac.id

Cc: hazlina@umt.edu.my, harfiandri@bunghatta.ac.id, netti.aryani@lecturer.unri.ac.id, azrita31@bunghatta.ac.id, mkrishnan@wiley.com, production.a@hindawi.com

Dear Dr. Hafrijal,

I am pleased to let you know that the first set of galley proofs of your Research Article 6639837 titled "Analysis of Fatty Acids and Amino Acids of Three Local Freshwater Bagridae Fish Species in the Kampar Kanan River, Indonesia for Food Security," is ready. You can apply your corrections directly to the manuscript with the Online Proofing System (OPS).

Using the OPS, you can quickly and easily make corrections directly to your galley proofs and submit these corrections with a single click.

<https://ops.hindawi.com/author/6639837/>

Please note, although all authors can view the proof, it is only the submitting author (the author addressed in this email) who has the ability to edit and submit the corrections. However, the submitting author can log in to the OPS and re-assign the proof to another author if necessary. The submitting author will need to log in with the email address included on this email.

If a new corresponding author is added, they must log into their manuscript tracking system account and add their ORCID ID. Any additional ORCID IDs added on during proofing will also need to be updated on that author's account. Delays can occur if this isn't done.

We encourage all authors to provide figures that are suitable for visually impaired readers. Please refer to the section "Are your figures accessible to all readers?" on our website <https://www.hindawi.com/publish-research/authors/ready-submit/> for advice on how to make your figures as accessible as possible, including guidelines on preferred colour combinations. Please upload any replacement figure files as attachments to the online proofing system.

Here are some suggested points for you to review when checking your manuscript proofs:

- i) Each section heading is correct and in sequence.
- ii) Any figure legends or table captions are correct and correspond to the correct figure/table.
- iii) Any figures or tables are in correct sequential order.
- iv) Any abbreviations are consistent through the manuscript.

To expedite the publication of your manuscript, please send us your corrected galley proofs within two days.

Please ensure that you read the proofs thoroughly and make all necessary corrections at this stage. A second round of proofs may be requested only for checking essential changes or major revisions.

Best regards,

--

Hindawi Production Team

Hindawi

<https://www.hindawi.com>

hafrizal syandri <syandri_1960@bunghatta.ac.id>

Fri, Dec 29, 2023 at 10:26 AM

To: International Journal of Food Science <production.a@hindawi.com>

Dear,

Hindawi Production Team

Thank you for your email on Dec 27, 2023. The author has checked the galley proofs Manuscript no. 6639837 and agree to publish.

Best regards
Hafrijal Syandri

[Quoted text hidden]

99+

Compose

Mail

Inbox 1,768

Chat

Starred

Spaces

Snoozed

Sent

Meet

Drafts 9

More

Labels



6639837: Your article has been published External Inbox x



Deepanaa Rajadurai <drajadurai@wiley.com>
to me

Dear Dr. Azrita,

I am pleased to let you know that your article has been published in its final form in "International Journal of Food Science, vol. 2024, Article ID 6639837, 8 pages, 2024. <https://doi.org/10.1155/2024/6639837>.

You can access this article from the Table of Contents of Volume 2024, which is located at the following link:

<https://www.hindawi.com/journals/ijfs/contents/>

Alternatively, you can access your article directly at the following location:

<https://www.hindawi.com/journals/ijfs/2024/6639837/>

"International Journal of Food Science" is an open access journal, meaning that the full-text of all published articles is not behind pay barriers.

If you would like to order reprints of this article please click here, <https://www.hindawi.com/journals/ijfs/2024/6639837/reprints>

Our [Science Communication guide](#) provides practical tips on how to maximize the visibility and impact of your article, in addition to communicating science in an engaging and effective way. Don't forget to make the most of your [exclusive discount](#)

We would love to know what you think about your experience publishing with us. Please share your feedback in this brief survey:

[Survey Link](#)

Thank you for publishing your article with Hindawi, and we hope that you continue to choose International Journal of Food Science.

99+

Compose

Mail

Inbox

1,768

Chat

Starred

Spaces

Snoozed

Sent

Meet

Drafts

9

More

Labels

_____ ✉

Welcome, and congratulations on your first published article in Internat

External Inbox x



International Journal of Food Science <IJFS@email.hindawi.com>
to me

Can't see this email? [View online](#)



- [Aims & Scope](#)
- [Recent Articles](#)
- [Indexing](#)

Dear Dr Azrita Azrita,

Congratulations on publishing your first article, '[Analysis Of Fatt Three Local Freshwater Bagridae Fish Species In The Kampar I Food Security](#)', with us in *International Journal of Food Science* you as part of our scholarly community, your contribution is valu journal and its readers, and we look forward to supporting your r

In this upcoming series of emails, we'll be providing you with ins to enhance the impact of your work. Stay tuned for exciting upd

If you have any questions or require any further support, please

Best wishes,

Research Article

Analysis of Fatty Acids and Amino Acids of Three Local Freshwater Bagridae Fish Species in the Kampar Kanan River, Indonesia, for Food Security

Azrita Azrita ¹, Hafrijal Syandri ¹, HazlinaAhamad Zakeri ²,
Harfiandri Damanhuri ¹ and Netti Aryani ³

¹Faculty of Fisheries and Marine Science, Bung Hatta University, 25131 Padang, West Sumatra Province, Indonesia

²Faculty of Science and Marine Environment, Universiti Malaysia Terengganu, Malaysia

³Faculty of Fisheries and Marine Science, Riau University, 28293 Pekanbaru, Riau Province, Indonesia

Correspondence should be addressed to Hafrijal Syandri; syandri_1960@bunghatta.ac.id

Received 13 July 2023; Revised 15 December 2023; Accepted 19 December 2023; Published 4 January 2024

Academic Editor: Farid Mansouri

Copyright © 2024 Azrita Azrita et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Fish have become an irreplaceable dietary source of animal protein, especially among households with low socioeconomic status in rural and urban areas of Indonesia. This study is aimed at analysing the proximate composition, minerals, fatty acids, and amino acids of three local Bagridae fish species in the Kampar Kanan river, Indonesia. The standard AOAC method was employed to examine the proximate composition of the carcass, and the analysis of amino acids and fatty acids was conducted through HPLC and GC-MS, respectively. The mineral content was determined using AAS. The nutrient composition results of *Hemibagrus nemurus*, *Hemibagrus wyckii*, and *Mystus nigriceps* revealed that the protein content was 24.26%, 22.57%, and 21.39% (% dry weight), whereas the total lipid content was 6.64%, 7.47%, and 7.75%, respectively. Regarding mineral contents, the calcium levels ranged from 1.49 to 1.66 mg/g, iron levels from 28.35 to 40.36 µg/g, and zinc levels from 24.03 to 54.46 µg/g. Among the fatty acids, palmitic acid was the most abundant in all three species, accounting for 25.59–30.70% of the total fatty acids. Additionally, significant amounts of C18:1 (oleic acid), C18:0 (stearic acid), and C20:4 (arachidonic acid) were also detected as primary fatty acids. The calculated atherogenic index values in the three species of Bagridae fish ranged from 0.73 to 0.99, while the thrombogenic index values varied between 0.54 and 0.75. The predominant amino acids found in the three species of Bagridae fish were glutamic acid with their concentrations ranging from 9.10 to 24.34%. These results indicate that consuming the meat of these three freshwater Bagridae fish species caught in the wild does not pose any health risks to consumers. They can be considered a safe and suitable food source with good nutritional quality.

1. Introduction

Fish serve as a high-quality protein source that is essential for satisfying dietary protein requirements, especially in areas where other protein sources are few or difficult to obtain [1, 2]. Their accessibility can aid in ensuring a steady supply of food, particularly for communities whose main source of income is fishing or aquaculture [3]. Fish can contribute to a more varied selection of protein sources in communities, resulting in a diet that is more nutritious and balanced. Dietary fish can treat nutrient deficits and enhance general nutritional health. Fish are generally considered safe

and offer a lower risk of contamination with harmful substances compared to some land-based animal products. A reliable indicator of a fish's quality, nutritional value, physiological status, and environment is its chemical makeup, which is found in fish flesh [4].

Hemibagrus nemurus, locally known as “baung,” *Hemibagrus wyckii*, known as “geso,” and *Mystus nigriceps*, known as “ingir-ingir,” are three species of catfish found in the Kampar Kanan River, Indonesia [5, 6]. Due to their high market value, these species have been identified as significant economic resources for rural communities [7]. Therefore, the analysis of the nutritional quality of fish meat in

the study area is very important to determine whether it is beneficial to human health.

Knowing the composition of fatty acids and amino acids in fish meat is an essential factor that should not be ignored [8]. This information provides valuable insight into the nutritional quality of fish and the value of more nutritious and healthy foods [9]. In addition, a better understanding of the composition of fatty acids and amino acids can assist in selecting a suitable feed [10], developing a balanced diet [11], and maintaining optimal fish meat nutrition [12].

Omega-3 polyunsaturated fatty acids (PUFAs), particularly docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA) derived from fish oil, exhibit anti-inflammatory and antigastrointestinal cancer properties, making them crucial as immune-boosting nutrients [13, 14]. Studies have demonstrated that ω -3 and ω -6 PUFAs exhibit beneficial effects in the management of cardiovascular diseases and cancers [15, 16]. The fatty acid composition of PUFAs can differ among different types of fish, including both freshwater and marine species [17, 18]. Amino acids (AAs), such as cysteine, arginine, tyrosine, glycine, proline, and serine, play a vital role in illness and stress situations and in the prevention of inflammation and repair processes in fish intestines [19, 20].

Furthermore, knowledge of the atherogenicity index (AI) and thrombogenicity index (TI) of fatty acids in the three local freshwater fish species of Bagridae can contribute to improving dietary guidelines. The AI and TI can be used to develop recommendations aimed at improving heart health [21, 22]. These recommendations may involve selecting fish with favourable fatty acid profiles, allowing individuals to choose the most suitable fish species based on their specific needs [17, 23]. Therefore, this study is aimed at examining the nutritional composition profile, mineral content, amino acids, fatty acids, and AI and TI of three freshwater fish species, namely, *H. nemurus*, *H. wyckii*, and *M. nigriceps*. These three fish species are generally consumed by people who live around the Kampar Kanan River in Kampar Regency, Riau Province, Indonesia.

2. Material and Methods

2.1. Animal Material. In this study, a total of 27 Bagridae fish belonging to three distinct species (*H. nemurus*, *H. wyckii*, and *M. nigriceps*) were examined. The specimens were obtained from local fishers operating in the upper stretches of the Kampar Kanan River in Kampar Regency, Riau Province (Figure 1). Sampling took place between June and August 2022 and was verified by the Fisheries Department, Faculty of Fisheries and Marine Science, Bung Hatta University, Indonesia. The samples were placed in plastic bags and transported to the laboratory in an insulated icebox. The samples were then grouped based on their respective sampling locations.

2.2. Biometry Measurement. Upon reaching the laboratory within 10 hours, the fish samples were subjected to gutting and washing. Each specimen was weighed (TW) and measured to determine the standard length (SL) as well as the

depth or maximum height (H). TW was measured using a balance scale (OHAUS Model CT 6000 USA) with a precision of 0.01 g. The length measurement was taken from the tip of the mouth to the end of the upper lobe of the caudal fin, representing the total body length, using a metre ruler with an accuracy of 1 mm. The height measurement involved a vertical assessment of the body's maximum height measured using a Digital Sekhmet Sigma Vernier calliper with an accuracy of 1 mm. The condition factor (CF) was calculated using the formula $CF = (TW/SL^3) \times 100$.

2.3. Proximate, Amino Acid, and Mineral Content Analysis.

The proximate carcass composition was assessed following the AOAC standard methods [24]. The samples were dried to a constant weight at 105°C. The crude protein content was analysed using the standard Kjeldahl method, which involves multiplying the nitrogen content by a factor of 6.25. The Soxhlet method with ether extraction was employed to analyse crude lipids. The ash content was obtained by incinerating the samples at 550°C for 16 h. The amino acid composition was determined using a high-performance liquid chromatography (HPLC) system, which consisted of a Waters® 1525 binary HPLC pump, Waters® 717 autosamplers, and Waters® 2475 multi- λ fluorescence detector (with excitation at 250 nm and emission at 395 nm). The samples were hydrolysed in triplicate with 6 N hydrochloric acid for 24 h at 11°C [25].

For mineral data composition (Na, Mg, Ca, K, P, Fe, Cu, Mn, and Zn), the ashed feed sample and carcass were dissolved in 1 ml of hydrochloric acid (35% v/v Suprapur®, Merck) and then filtered with cellulose filter paper (Watchman No. 1, International Ltd; Maidstone, UK) and diluted to an appropriate concentration for each elemental mineral. P levels were analysed with a Perkin-Elmer AA spectrophotometer mod 3110 (Norwalk, CT, USA).

2.4. Fatty Acid Analysis. The fish meat was examined utilizing the gas chromatography-mass spectrometry (GC-MS) method. The method of Folch et al. (1957) modified by Rajion et al. [26] was employed to perform the total fat extraction. This involved using a solvent system consisting of chloroform and methanol at a 2:1 (v/v) ratio. The process of transmethylation was conducted using a solution of 14% methanolic boron trifluoride. The fatty acid composition of the meat was then analysed at the SIG Laboratory, Accredited Testing Laboratory-LP-184-IDN.

The nutritional quality of lipids (AI and TI) was calculated using the following equations [27]:

$$AI = \frac{[(C12 : 0 + 4 \times C14 : 0 + C16 : 0)]}{[(\sum MUFA + \sum n - 6) + \sum n - 3]}$$

$$TI = \frac{[(C14 : 0 + C16 : 0)]}{[(0.5 \times \sum MUFA) + (0.5 \times \sum n - 6) + (3 \times \sum n - 3) + (\sum n - 3 / \sum n - 6)]}$$
(1)

where AI is the atherogenic index, TI is the thrombogenic index, C12 : 0 is the lauric acid, C14 : 0 is the myristic acid, C16 : 0 is the palmitic acid, $\sum MUFA$ is the sum of the concentrations of all monosaturated fatty acids, $\sum n - 6$ is the sum of the concentrations of n-6 polyunsaturated fatty acids,

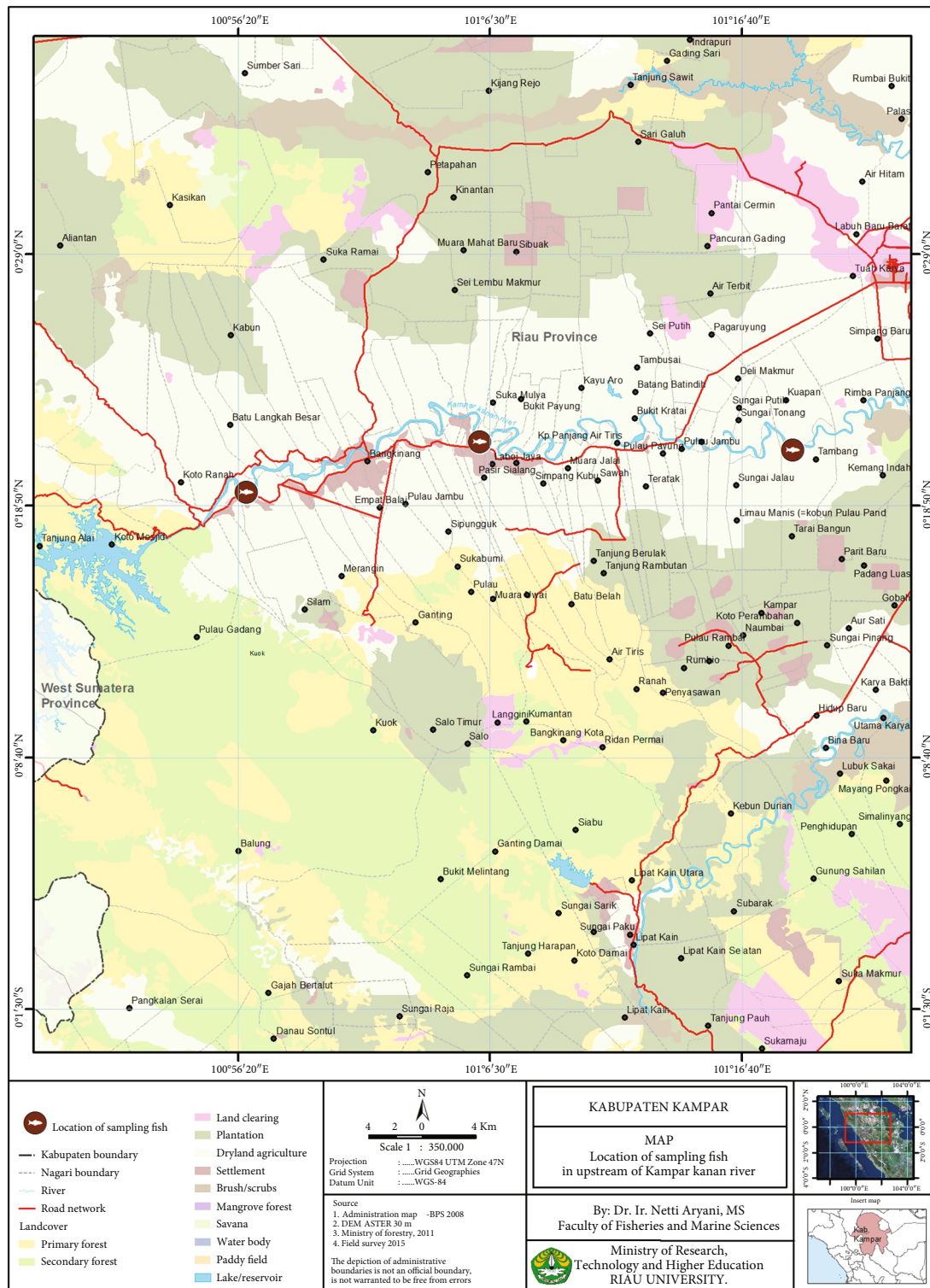


FIGURE 1: Map of Kampar Regency, Riau Province, and sampling locations of the three species of Bagridae fish.

and $\sum n - 3$ is the sum of the concentrations of n-3 polyunsaturated fatty acids.

2.5. Data Analysis. Data analysis was performed using the SPSS 16.0 software package (SPSS; Chicago, IL). The homogeneity of variances was assessed using Levene's test.

One-way ANOVA was carried out to determine the parameters of proximate and amino acid composition, including the composition of fatty acids and nutritional quality of lipids (AI and TI) for each species, followed by the post hoc Duncan's multiple range test [28]. The results are reported as the mean values \pm standard errors for each parameter.

3. Results

Table 1 presents the average wet weight, standard length, height, and the results of meat nutritional composition analysis for three indigenous freshwater Bagridae species in the Kampar Kanan river, Indonesian. The moisture content (% wet weight) ranged from 82.40 to 85.39%. Among the species examined, *M. nigriceps* exhibited the lowest protein content at 21.39%, while *H. nemurus* displayed the highest value at 24.26%. However, *M. nigriceps* had a higher mineral content, including iron and zinc.

Table 2 summarizes the fatty acid composition (% total fatty acid) for the three Bagridae fish species. C16:0 was the predominant fatty acid in the three species of Bagridae fish, with percentages ranging from 25.09 to 30.71%. The additional prominent fatty acids found were C18: 1, C18:0, and C22:6. *H. nemurus* exhibited a ω -6/ ω -3 ratio below 1 (0.82), while *H. wyckii* and *M. nigriceps* had ratios of 1.11 and 1.02, respectively. The obtained PUFA/saturated fatty acid (SFA) (P/S) ratio varied between 0.61 and 0.76. The AI values varied between 0.76 and 0.99 and the TI between 0.40 and 0.63 (Table 3).

Table 4 presents the amino acid composition (% of total protein) of the three Bagridae species. Glutamic acid was the dominant amino acid, ranging from 19.29 to 24.34%, followed by aspartic acid, which ranged from 9.21 to 11.27%. The lysine content was consistent, ranging from 9.67 to 9.87%. In *H. nemurus*, the levels of various amino acids ranged from 0.86 to 24.35%. Similarly, in *H. wyckii*, the amino acid levels ranged from 0.76 to 21.58%, while in *M. nigriceps*, the levels ranged from 0.74 to 19.29%.

4. Discussion

The results of this study provide important new information about the nutritional makeup of the studied species of Bagridae fish. The biometric measures of the fish yielded important information about their physical traits and health. Researchers are able to determine the average weight of fish at various lengths by examining the link between length and weight for a particular species. This knowledge is essential for understanding the trends in fish population growth, identifying potential shifts in growth rates over time, and assessing the general health and welfare of fish [29]. A variety of factors, including sex, age, maturity level, size, stomach fullness level, sampling strategies, sample sizes, and environmental circumstances, have an impact on fish health and the parameters that determine length-weight connections in fish [30]. However, none of these factors were considered in our study. In general, according to the theory put forth by Bagenal and Tesch [31], heavier fish of a given length are in better physiological condition. The condition factor is also used in fishery science to compare the health, fatness, and condition of fish [32]. A condition factor of >1 is favourable, indicating a good feed nutrition level and suitable living habitat [33]. Thus, based on the findings in this study, the Kampar Kanan River provides a suitable habitat for the fish, ensuring their accessibility and availability for community consumption.

The fish samples exhibited good nutritional profiles, with high amounts of proteins and lipids as well as moisture content contributing to their overall nutritional value, according to the results of proximate analysis and mineral composition (see Table 1). These results imply that the fish species studied provide a possible source of nutrients to solve issues with food security. The study of the mineral composition revealed the existence of critical minerals such as phosphorus (P), calcium (Ca), magnesium (Mg), iron (Fe), and zinc (Zn) in the fish samples in addition to proteins and lipids. All three Bagridae fish significantly differed from each other in terms of their crude ash content and mineral profile (see Table 1). However, in general, P and Ca are the two highest minerals found in all fish. Both of these macrominerals are good for human bone and tooth health, as key components of hydroxyapatite, the mineral matrix that provides strength and structure to bones and teeth [34].

In the current study, the meat of three species of Bagridae fish from the Kampar Kanan River in Indonesia was found to contain an abundance of microminerals such as Fe and Zn. *M. nigriceps* has been reported to exhibit higher mineral content, including Fe and Zn. This species is categorized as a small indigenous fish (SIF) with a maximum standard length of approximately 10.29 cm. According to [35], SIFs are generally known to be rich in Ca, Fe, and Mn. On the other hand, scale flour from three species of freshwater fish, such as *Osphronemus goramy*, *Cyprinus carpio*, and *Oreochromis niloticus*, also contains high levels of macrominerals and microminerals [36].

In this study, all three Bagridae fish had a total fat content of approximately 7–8% by weight, with a higher water content ranging from 82.40 to 85.39% (Table 1). These values are similar to those found in wild and cultured sea bass *Dicentrarchus labrax* [37]. Additionally, the fish species can be grouped into different categories, including high-fat fish ($>8\%$), medium-fat fish (4–8%), low-fat fish (2–4%), and lean fish ($<2\%$) [38]. According to the classification established by [38], the three indigenous Bagridae fish species are characterized as having a moderate fat content level, which is reflected by the high levels of DHA and EPA in the meat of *H. nemurus*, *H. wyckii*, and *M. nigriceps* (Table 2). Therefore, we recommend that consuming the meat of these three fish species can indirectly boost food security in rural and urban communities.

All three Bagridae fish species analysed in the study contained arachidonic acid (C20:4), at 12.13% for *H. nemurus*, 12.49% for *H. wyckii*, and 12.41% for *M. nigriceps*. Arachidonic acid serves as a precursor for synthesizing prostaglandins and leukotrienes [39]. Leukotrienes are crucial in allergic responses, inducing smooth muscle contraction and increasing vascular permeability [40]. Additionally, all three species also contained EPA ranging from 1.35 to 2.34% and DHA ranging from 11.56 to 15.65%. The increased consumption of omega-3 fatty acids, such as EPA and DHA, has been linked to a decreased risk of pain development in older individuals and to the prevention of Alzheimer's disease [41, 42].

As determined by the current study, the ω -6/ ω -3 ratios of the three Bagridae fish species ranged from 1:0.8 to

TABLE 1: Results of biometric, proximate, and mineral composition of three species of Bagridae fish.

	Species		
	<i>Hemibagrusnemurus</i>	<i>Hemibagruswyckii</i>	<i>Mystusnigriceps</i>
<i>Biometric measurements</i>			
Total weight (g)	389.99 ± 24.96 ^a	1390.33 ± 168.29 ^b	17.57 ± 1.53 ^c
Standard length (cm)	28.16 ± 0.53 ^a	44.14 ± 1.98 ^b	10.29 ± 0.15 ^c
Height (cm)	8.44 ± 0.16 ^a	8.82 ± 0.39 ^b	2.57 ± 0.03 ^c
Condition factor	1.63 ± 0.08	1.58 ± 0.07	1.56 ± 0.08
<i>Proximate composition</i>			
Crude protein (% DW)	24.26 ± 0.87 ^a	22.57 ± 0.37 ^b	21.39 ± 0.15 ^c
Crude fat (% DW)	6.64 ± 0.03 ^a	7.47 ± 0.02 ^b	7.75 ± 0.40 ^c
Crude ash (% DW)	1.94 ± 0.02 ^a	2.30 ± 0.09 ^b	1.59 ± 0.02 ^c
Moisture (% WW)	82.40 ± 2.51	85.39 ± 2.36	83.75 ± 2.21
<i>Mineral composition</i>			
Macrominerals (mg/g)			
Sodium	0.99 ± 0.00 ^a	1.61 ± 0.01 ^b	1.61 ± 0.02 ^c
Magnesium	1.12 ± 0.01 ^a	0.57 ± 0.02 ^b	1.15 ± 0.01 ^c
Calcium	1.66 ± 0.00 ^a	1.55 ± 0.02 ^b	1.49 ± 0.04 ^c
Potassium	0.71 ± 0.00 ^a	0.55 ± 0.02 ^b	0.43 ± 0.01 ^c
Phosphorous	7.03 ± 0.03 ^a	2.74 ± 0.02 ^b	6.45 ± 0.07 ^c
Microminerals (µg/g)			
Iron	28.30 ± 0.11 ^a	28.73 ± 0.08 ^b	40.36 ± 0.55 ^c
Copper	8.93 ± 0.03 ^a	7.46 ± 0.09 ^b	6.21 ± 0.32 ^c
Manganese	1.64 ± 0.02 ^a	2.64 ± 0.02 ^b	2.84 ± 0.02 ^c
Zinc	24.03 ± 0.45 ^a	24.61 ± 0.19 ^b	54.46 ± 0.17 ^c

Values are mean% ± SE. Values in the same row followed by different letters are significantly different ($p < 0.05$).

TABLE 2: Fatty acid composition of three species of Bagridae fish.

Fatty acid	<i>Hemibagrusnemurus</i>	<i>Hemibagruswyckii</i>	<i>Mystusnigriceps</i>
C12:0 (lauric acid)	1.97 ± 0.05 ^a	3.94 ± 0.01 ^b	4.27 ± 0.01 ^c
C14:0 (meristic acid)	2.38 ± 0.03 ^a	2.72 ± 0.08 ^b	2.89 ± 0.01 ^c
C16:0 (palmitic acid)	27.23 ± 0.06 ^a	30.70 ± 0.06 ^b	25.59 ± 0.01 ^c
C18:0 (stearic acid)	16.17 ± 0.02 ^a	13.54 ± 0.02 ^b	10.64 ± 0.01 ^c
C20:0 (arachidic acid)	0.23 ± 0.01 ^a	0.14 ± 0.01 ^b	0.37 ± 0.01 ^c
C16:1 (palmitoleic acid)	1.78 ± 0.00 ^a	1.27 ± 0.04 ^b	4.93 ± 0.01 ^c
C18:1(oleic acid)	16.86 ± 0.01 ^a	16.18 ± 0.01 ^b	16.97 ± 0.01 ^c
C18:2 (linoleic acid)	3.24 ± 0.02 ^a	3.84 ± 0.33 ^b	4.55 ± 0.001 ^c
C18:3 (linolenic acid)	1.12 ± 0.01 ^a	0.82 ± 0.01 ^b	1.57 ± 0.00 ^c
C20:4 (arachidonic acid)	12.13 ± 0.01 ^a	12.49 ± 0.09 ^b	12.41 ± 0.02 ^c
C20:5 (eicosapentaenoic acid; EPA)	2.06 ± 0.08 ^a	2.34 ± 0.04 ^b	1.35 ± 0.01 ^c
C22:6 (docosahexaenoic acid; DHA)	15.65 ± 0.01 ^a	11.56 ± 0.04 ^b	13.64 ± 0.02 ^c

Values are mean% ± SE. Values in the same row with different superscripts are significantly different ($p < 0.05$).

1:0.97. It was found that all three fish species had ω -6/ ω -3 ratios within the recommended range for a typical Indonesian diet. In contrast, a typical Western diet is characterized by a high ω -6/ ω -3 ratio of 10:1 to 20:1, while a Mediterranean diet has a ratio of 4:1 to 5:1 [43]. Additionally, the P/S ratio analysis showed that the three Bagridae fish species are sources of PUFAs that meet the requirements for food safety.

The Food and Agriculture Organization (FAO) and the World Health Organization (WHO) have recommended AI and TI values ranging from 0.4 to 0.5 for promoting human health [44]. In the current study, the observed AI values in the three freshwater species of Bagridae ranged from 0.73 to 0.99, while the TI values varied between 0.40 and 0.63. This finding is associated with a notable disparity

TABLE 3: Fatty acid ω -6/ ω -3 and polyunsaturated/saturated fatty acid ratio of three species of Bagridae fish.

	<i>Hemibagrusnemurus</i>	<i>Hemibagruswyckii</i>	<i>Mystusnigriceps</i>
ω -6/ ω -3 ratio	0.82 ± 0.00 ^a	1.11 ± 0.02 ^b	1.02 ± 0.01 ^c
P/S ratio	0.69 ± 0.01 ^a	0.61 ± 0.06 ^b	0.76 ± 0.00 ^c
AI	0.87 ± 0.01 ^a	0.99 ± 0.06 ^b	0.73 ± 0.00 ^c
TI	0.40 ± 0.01 ^a	0.63 ± 0.01 ^b	0.43 ± 0.00 ^c

P/S: polyunsaturated/saturated fatty acid; AI: atherogenic index; TI: thrombogenic index. Values are *mean* ± *SE*. Values in the same row followed by different letters are significantly different ($p < 0.05$).

TABLE 4: Amino acid composition of three species of Bagridae fish.

Amino acid	<i>Hemibagrusnemurus</i>	<i>Hemibagruswyckii</i>	<i>Mystusnigriceps</i>
Aspartic acid	10.39 ± 0.01 ^a	9.61 ± 0.04 ^b	11.28 ± 0.02 ^c
Glutamic acid	24.34 ± 0.00 ^a	21.58 ± 0.41 ^b	19.29 ± 0.03 ^c
Serine	3.74 ± 0.00 ^a	5.40 ± 0.03 ^b	7.50 ± 0.06 ^c
Glycine	4.02 ± 0.00 ^a	5.77 ± 0.04 ^b	4.69 ± 0.02 ^c
Histidine	2.39 ± 0.00 ^a	2.90 ± 0.03 ^b	2.21 ± 0.02 ^c
Arginine	6.17 ± 0.01 ^a	8.27 ± 0.03 ^b	6.22 ± 0.03 ^c
Threonine	4.49 ± 0.01 ^a	4.19 ± 0.03 ^b	4.39 ± 0.04 ^c
Alanine	7.03 ± 0.00 ^a	6.04 ± 0.02 ^b	7.89 ± 0.01 ^c
Proline	2.40 ± 0.00 ^a	2.71 ± 0.01 ^b	2.21 ± 0.02 ^c
Tyrosine	3.91 ± 0.01 ^a	3.29 ± 0.06 ^b	3.82 ± 0.04 ^c
Valine	3.73 ± 0.00 ^a	4.02 ± 0.01 ^b	3.85 ± 0.04 ^c
Methionine	3.42 ± 0.00 ^a	1.96 ± 0.02 ^b	1.27 ± 0.03 ^c
Cystine	0.87 ± 0.00 ^a	0.76 ± 0.02 ^b	0.75 ± 0.03 ^c
Isoleucine	3.82 ± 0.001 ^a	3.96 ± 0.003 ^b	3.66 ± 0.002 ^c
Leucine	6.15 ± 0.000 ^a	6.69 ± 0.002 ^b	6.28 ± 0.004 ^c
Phenylalanine	5.13 ± 0.000 ^a	6.03 ± 0.005 ^b	5.18 ± 0.027 ^c
Lysine	9.61 ± 0.001 ^a	9.82 ± 0.026 ^b	9.87 ± 0.003 ^c

Values are *mean* ± *SE*. Values in the same row followed by different letters are significantly different ($p < 0.05$).

in the values of SFAs among the three Bagridae fish species. Based on these findings, we confirm that consuming the meat of the three freshwater species of Bagridae fish caught in the wild does not endanger consumer health and can serve as a suitable source of food safety and nutritional quality.

Fish with a high crude protein content, as shown by the three studied Bagridae fish (Table 1), are regarded as being nutritionally useful for human health. The protein and amino acid contents can directly indicate the nutritional quality of meat [45]. As shown in Table 4, *H. wyckii* had a higher content of essential amino acids (EAAs) than *H. nemurus* and *M. nigriceps*. Among the EAAs, lysine (Lys) was found to be present in the highest amount in all three species. Lys is necessary for tissue growth and repair, reduced stress-induced anxiety, and the conversion of fatty acids into energy [46–48].

More than 60% of the amino acid composition in all three Brigade fish studied comes from nonessential amino acids (NEAAs), particularly glutamic acid (Table 4). Glutamic acid and its amine, glutamine, are highly abundant amino acids found in fish in the free and protein-bound

forms [49]. Other amino acids found at abundant levels in the three fish also play important physiological and biochemical functions. The NEAA arginine, for example, is involved in protein synthesis, creatine formation, and various other biochemical processes within the body. The EAAs leucine, phenylalanine, and methionine play important roles in the synthesis of nitric oxide, a molecule that can expand blood vessels [49].

5. Conclusions

The present study provides insightful information about the fatty acid and amino acid compositions, biometric measurements, proximate analysis, and mineral composition of three local freshwater species Bagridae fish in the Kampar Kanan river, Indonesian. The findings show their potential as a nutrient-dense, sustainable food source that can support efforts to increase food security. These fish species might be incorporated into the local diet to increase nutrient intake and alleviate nutritional inadequacies, which would ultimately improve Indonesia's overall food security status.

Data Availability

The data utilized in this research has not been previously released or published in any form. The datasets employed and/or analysed during the present study can be obtained by contacting the corresponding author.

Conflicts of Interest

The authors confirm that they do not have any conflicting interests.

Acknowledgments

The authors acknowledge and express their gratitude for the financial support received from the Ministry of Education, Culture, Research, and Technology of the Republic of Indonesia, specifically through grant no. 076/E5/PG. 02.00. PT/2022, which funded the research. Furthermore, the authors extend their appreciation to the students and fishers involved in the research conducted in the upper reaches of the Kampar Kanan River, Indonesia.

References

- [1] A. E. Hasselberg, I. Aakre, J. Scholtens et al., "Fish for food and nutrition security in Ghana: challenges and opportunities," *Global Food Security*, vol. 26, article 100380, 2020.
- [2] N. Tran, U. P. Rodriguez, C. Y. Chan, Y. M. Auang, L. Chu, and A. H. M. S. Islam, "Future scenarios of fish supply and demand for food and nutrition security in Bangladesh: an analysis with the AsiaFish model," *Aquaculture*, vol. 568, article 739288, 2023.
- [3] C. Z. Levkoe, K. Lowitt, and C. Nelson, "Fish as food: exploring a food sovereignty approach to small-scale fisheries," *Marine Policy*, vol. 85, pp. 65–70, 2017.
- [4] M. Ravichandran, N. S. Hettiarachchy, V. Ganesh, and S. Ricke, "Enhancement of antimicrobial activities of naturally occurring phenolic compounds by nanoscale delivery against *Listeria monocytogenes*, *Escherichia coli* O157: H7 and *Salmonella typhimurium* in broth and chicken meat system," *Journal of Food Safety*, vol. 31, no. 4, pp. 462–471, 2011.
- [5] N. Aryani, I. Suharman, H. Syandri, and A. Mardiah, "Diversity and distribution of fish fauna of upstream and downstream areas at Koto Panjang Reservoir, Riau Province, Indonesia," *F1000 Research*, vol. 8, no. 1435, pp. 1–16, 2019.
- [6] D. Syafrialdi, D. I. Roesma, and H. Syandri, "Length-weight relationships and condition factor of two-spot catfish (*Mystus nigriceps* [Valenciennes, 1840]) (Pisces, Bagridae), from Kampar Kanan River and Kampar Kiri River in Indonesia," *Pakistan Journal of Biological Science*, vol. 23, no. 12, pp. 1636–1642, 2020.
- [7] N. Aryani, "Native species in Kampar Kanan River, Riau Province Indonesia," *International Journal of Fisheries and Aquatic Studies*, vol. 2, no. 5, pp. 213–217, 2015.
- [8] A. Azrita, H. Syandri, N. Aryani, A. Mardiah, and I. Suharman, "The utilization of new products formulated from water coconut, palm sap sugar, and fungus to increase nutritional feed quality, feed efficiency, growth, and carcass of gurami sago (*Osphronemus goramy* Lacepède, 1801) juvenile," *F1000 Research*, vol. 10, no. 1121, pp. 1–16, 2021.
- [9] C. Bene, R. Arthur, H. Norbury et al., "Contribution of fisheries and aquaculture to food security and poverty reduction: assessing the current evidence," *World Development*, vol. 79, pp. 177–196, 2016.
- [10] N. Aryani, I. Suharman, S. Hasibuan, N. Asia, and H. Syandri, "Fatty acid composition on diet and carcasses, growth, body indices and profile serum of Asian redtail catfish (*Hemibagrus nemurus*) fed a diet containing different levels of EPA and DHA," *F1000 Research*, vol. 11, no. 1409, pp. 1–23, 2022.
- [11] E. N. Kingsley, O. O. Cyril, and O. I. Patience, "Potential contribution of selected wild fish species to the minerals intake of pregnant and lactating women, children and adults in rural riverine communities of Edo State: insights and outcomes," *Measurement: Food*, vol. 8, article 100063, pp. 1–7, 2022.
- [12] A. A. Sausa, L. Nora, D. L. A. Lopes et al., "Vegetable choline in feed for Nile tilapia (*Oreochromis niloticus*) raised in a biofloc technology system (BFT): biofloc composition, chemical composition, and fatty acid profiles in meat," *Aquaculture*, vol. 545, article 737174, 2021.
- [13] A. M. Eltweri, A. L. Thomas, M. Metcalfe, P. C. Calder, A. R. Dennison, and D. J. Bowrey, "Potential applications of fish oils rich in omega-3 polyunsaturated fatty acids in the management of gastrointestinal cancer," *Clinical Nutrition*, vol. 36, no. 1, pp. 65–78, 2017.
- [14] N. Tintle, T. Rice, I. Cheng et al., "The association between genetically elevated polyunsaturated fatty acids and risk of cancer," *eBioMedicine*, vol. 91, article 104510, 2023.
- [15] E. K. Aglago, I. Huybrechts, N. Murphy et al., "Consumption of fish and long-chain n-3 polyunsaturated fatty acids is associated with reduced risk of colorectal cancer in a large European cohort," *Clinical Gastroenterology and Hepatology*, vol. 18, no. 3, pp. 654–666.e6, 2020.
- [16] H. Jiang, X. Shi, T. Y. Fan et al., "Dietary omega-3 polyunsaturated fatty acids and fish intake and risk of age-related macular degeneration," *Clinical Nutrition*, vol. 40, no. 12, pp. 5662–5673, 2021.
- [17] M. A. Rincón-Cervera, N. V. González-Barriga, R. Valenzuela, S. Lopez-Arana, J. Romero, and A. Valenzuela, "Profile and distribution of fatty acids in edible parts of commonly consumed marine fishes in Chile," *Food Chemistry*, vol. 274, pp. 123–129, 2019.
- [18] S. Sun, T. Ren, X. Li, X. Cao, and J. Gao, "Polyunsaturated fatty acids synthesized by freshwater fish: a new insight to the roles of *elovl2* and *elovl5* in vivo," *Biochemical and Biophysical Research Communications*, vol. 532, no. 3, pp. 414–419, 2020.
- [19] M. B. Behyar, M. Hasanzadeh, F. Seidi, and N. Sadjou, "Sensing of amino acids: critical role of nanomaterials for the efficient biomedical analysis," *Microchemical Journal*, vol. 188, article 108452, 2023.
- [20] P. Shi, K. Liao, J. Xu, Y. Wang, S. Xu, and X. Yan, "Eicosapentaenoic acid mitigates palmitic acid-induced heat shock response, inflammation and repair processes in fish intestine," *Fish & Shellfish Immunology*, vol. 124, pp. 362–371, 2022.
- [21] J. Wen, L. Zeng, Y. Xu, Y. Sun, Z. Chen, and S. Fan, "Proximate composition, amino acid and fatty acid composition of fish maws," *Natural Product Research*, vol. 30, no. 2, pp. 214–217, 2016.
- [22] F. O. S. Duarte, F. G. de Paula, C. S. Prado et al., "Better fatty acids profile in fillets of Nile tilapia (*Oreochromis niloticus*) supplemented with fish oil," *Aquaculture*, vol. 534, article 736241, 2021.

- [23] K. L. Weaver, P. Ivester, J. A. Chilton, M. D. Wilson, P. Pandey, and F. H. Chilton, "The content of favorable and unfavorable polyunsaturated fatty acids found in commonly eaten fish," *Journal of the American Dietetic Association*, vol. 108, no. 7, pp. 1178–1185, 2008.
- [24] AOAC, *Official Methods of Analysis*, Association of Official Analytical Chemists (AOAC), Washington, DC., USA, 15th edition, 1990.
- [25] S. A. Cohen, "Amino acid analysis using pre-column derivatization with 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate," *Amino Acid Analysis Protocols*, vol. 211, pp. 143–154, 2000.
- [26] M. A. Rajion, J. G. McLean, and R. N. Cahill, "Essential fatty acid metabolism in the fetal and neonatal lamb," *Australian Journal of Biological Sciences*, vol. 38, no. 1, pp. 33–40, 1985.
- [27] T. L. Ulbricht and D. A. Southgate, "Coronary heart disease: seven dietary factors," *The Lancet*, vol. 338, no. 8773, pp. 985–992, 1991.
- [28] D. B. Duncan, "Multiple range and multiple F tests," *Biometrics*, vol. 11, no. 1, pp. 1–42, 1955.
- [29] R. Froese, "Cube law, condition factor and weight–length relationships: history, meta-analysis and recommendations," *Journal of Applied Ichthyology*, vol. 22, no. 4, pp. 241–253, 2006.
- [30] O. O. Famofofo and W. O. Abdul, "Biometry, condition factors and length-weight relationships of sixteen fish species in Iwopin fresh-water ecotype of Lekki Lagoon, Ogun State, Southwest Nigeria," *Heliyon*, vol. 6, no. 1, article e02957, 2020.
- [31] T. B. Bagenal and F. W. Tesch, "Age and growth," in *Methods for Assessment of Fish Production in Fresh Waters*, T. Bagenal, Ed., pp. 101–136, Blackwell Science Publications, 1978.
- [32] E. O. Ahmed, M. E. Ali, and A. A. Aziz, "Length-weight relationships and condition factors of six fish species in Atbara River and Khashm El-Girba reservoir, Sudan," *International Journal of Agriculture Sciences*, vol. 3, no. 1, pp. 65–70, 2011.
- [33] N. C. Ujjania, M. P. S. Kohli, and L. L. Sharma, "Length-weight relationship and condition factors of Indian major carps (*C. catla*, *L. rohita* and *C. mrigala*) in Mahi Bajaj Sagar, India," *Research Journal of Biology*, vol. 2, no. 1, pp. 30–36, 2012.
- [34] J. P. Bonjour, "Calcium and phosphate: a duet of ions playing for bone health," *Journal of the American College of Nutrition*, vol. 30, Supplement 5, pp. 438S–448S, 2011.
- [35] B. P. Mohanty, T. V. Sankar, S. Ganguly et al., "Micronutrient composition of 35 food fishes from India and their significance in human nutrition," *Biological Trace Element Research*, vol. 174, no. 2, pp. 448–458, 2016.
- [36] H. Syandri, A. Azrita, A. Mardiah, N. Aryani, and A. Diharmi, "The proximate composition, amino acid profile, fatty acid content, and mineral content of scale flour from three fish species as potential feeds for fish fry," *F1000 Research*, vol. 12, no. 1144, pp. 1–19, 2023.
- [37] F. Jabeen and A. A. Chudhry, "Chemical compositions and fatty acid profiles of three freshwater fish species," *Food Chemistry*, vol. 125, no. 3, pp. 991–996, 2011.
- [38] A. A. Nurnadia, A. Azrina, and I. Amin, "Proximate composition and energetic value of selected marine fish and shellfish from the west coast of peninsular Malaysia," *International Food Research Journal*, vol. 18, pp. 137–148, 2011.
- [39] S. C. Dyllal, L. Balas, N. G. Bazan et al., "Polyunsaturated fatty acids and fatty acid-derived lipid mediators: recent advances in the understanding of their biosynthesis, structures, and functions," *Progress in Lipid Research*, vol. 86, article 101165, 2022.
- [40] N. Kim, U. D. Sohn, V. Mangannan et al., "Leukotrienes in acetylcholine-induced contraction of esophageal circular smooth muscle in experimental esophagitis," *Gastroenterology*, vol. 112, no. 5, pp. 1548–1558, 1997.
- [41] D. Swanson, R. Block, and S. A. Mousa, "Omega-3 fatty acids EPA and DHA: health benefits throughout life," *Advances in Nutrition*, vol. 3, no. 1, pp. 1–7, 2012.
- [42] R. P. Mason, S. C. R. Sherratt, and L. H. Eckel, "Omega-3-fatty acids: do they prevent cardiovascular disease?," *Best Practice & Research Clinical Endocrinology & Metabolism*, vol. 37, no. 3, article 101681, 2023.
- [43] J. M. Seddon, J. Cote, N. Davis, and B. Rosner, "Progression of age-related macular degeneration," *Archives of Ophthalmology*, vol. 121, no. 6, pp. 785–792, 2003.
- [44] FAO/WHO, *Fat and Oils in Human Nutrition: Report of a Joint Expert Consultation*, FAO/WHO, Rome, Italy, 1994.
- [45] S. M. Pawar and S. R. Sonaware, "Fish muscle protein highest source of energy," *International Journal of Biodiversity and Conservation*, vol. 5, no. 7, pp. 433–435, 2013.
- [46] D. Tomé and C. Bos, "Lysine requirement through the human life cycle," *The Journal of Nutrition*, vol. 137, no. 6, pp. 1642S–1645S, 2007.
- [47] M. Watford, "Glutamine and glutamate: nonessential or essential amino acids?," *Animal Nutrition*, vol. 1, no. 3, pp. 119–122, 2015.
- [48] X. Li, S. Zheng, and G. Wu, "Nutrition and metabolism of glutamate and glutamine in fish," *Amino Acids*, vol. 52, no. 5, pp. 671–691, 2020.
- [49] M. Li, Y. Wu, and L. Ye, "The role of amino acids in endothelial biology and function," *Cells*, vol. 11, no. 8, p. 1372, 2022.